



Caribbean Amphibians and Reptiles

Edited by

Brian I. Crother

Academic Press

**Caribbean
Amphibians
and Reptiles**

This Page Intentionally Left Blank

Caribbean Amphibians and Reptiles

Edited by

Brian I. Crother

Department of Biology
Southeastern Louisiana University



Academic Press

San Diego London Boston New York Sydney Tokyo Toronto

Front cover photograph: *Anolis smallwoodi* from Guantanamo Bay.
(For more details, see Color Plate 2, Figure D.)

This book is printed on acid-free paper.

Copyright © 1999 by ACADEMIC PRESS

All Rights Reserved.

No part of this publication may be reproduced or transmitted in any form or by any means, electronic or mechanical, including photocopy, recording, or any information storage and retrieval system, without permission in writing from the publisher.

Academic Press

a division of Harcourt Brace & Company

525 B Street, Suite 1900, San Diego, California 92101-4495, USA

<http://www.apnet.com>

Academic Press

24-28 Oval Road, London NW1 7DX, UK

<http://www.hbuk.co.uk/ap/>

Library of Congress Catalog Card Number: 98-87235

International Standard Book Number: 0-12-197955-5

PRINTED IN THE UNITED STATES OF AMERICA

99 00 01 02 03 04 QW 9 8 7 6 5 4 3 2 1

Contents

<i>Contributors</i>	<i>ix</i>
<i>Foreword</i>	<i>xi</i>
<i>Preface</i>	<i>xiii</i>
<i>Acknowledgments</i>	<i>xv</i>
<i>Legends to Color Plates</i>	<i>xxi</i>

1 Over 300 Years of Collecting in the Caribbean

Ernest Williams

Introduction: Where We Are	1
Periods in the History of Knowledge of the West	
Indian Herpetofauna	6
Period I	6
Period II	12
Period III	17
Period IV	20
Period V	24
Period VI	26
The Seventh Period?	30

2 A Review of Cuban Herpetology

Alberto R. Estrada and Rodolpho Ruibal

History	31
The Species	35

3 Jamaica

Ronald I. Crombie

Introduction	63
Location–Topography–Geology	67
Rivers	69
Climate and Vegetation	71
Historical Summary	73
Problems	89

4 The Hispaniolan Herpetofauna: Diversity, Endemism, and Historical Perspectives, with Comments on Navassa Island

**Robert Powell, Jose A. Ottenwalder,
and Sixto J. Inchaustegui**

Introduction	93
Geography and Physiography	93
Vegetation	98
Diversity and Endemism	102
Historical Perspectives	139
Fossil Record	163
Current Status of the Herpetofauna	164

5 The Puerto Rico Area

Richard Thomas

Introduction: The Region	169
A Brief History of Herpetology in the Puerto Rico Area	171
The Puerto Rican Herpetofauna within the Greater Antillean Assemblages	171
Patterns of Distribution: The Puerto Rico Bank	172
Distribution within Puerto Rico	172
The Incurative and Anthropochore Herpetofauna	178

6 The Lesser Antillean Fauna

Ellen J. Censky and Hinrich Kaiser

Introduction	181
Diversity and Taxonomic History	187
Biogeography	219
Conservation	220

7 West Indian Herpetoecology

Robert W. Henderson and Robert Powell

Introduction	223
History	224
Some Aspects of the Ecology of the West Indian Herpetofauna	231
Summary	263
Future Emphasis	264

8 Evolutionary Relationships

Brian I. Crother

Introduction	269
Anura	270
Sauria	278
Serpentes	306
Testudines	326
Crocodylia	328
Conclusions	329
Appendix	331

9 Ecological and Historical Biogeography

Gregory K. Pregill and Brian I. Crother

Introduction	335
--------------	-----

Historical Biogeography	336
Ecological Biogeography	344
Coda	356

10 The West Indies and Middle America: Contrasting Origins and Diversity

William E. Duellman

Introduction	357
Materials and Methods	358
Origins of the Herpetofaunas	358
Diversity of the Herpetofaunas	364
Discussion	369

References 371

Systematic Index 475

Contributors

Numbers in parentheses indicate the pages on which the authors' contributions begin.

Ellen J. Censky (181), *Section of Amphibians and Reptiles, Carnegie Museum of Natural History, Pittsburgh, Pennsylvania 15213*

Ronald I. Crombie (63), *Division of Amphibians and Reptiles, National Museum of Natural History, Washington, DC 20560*

Brian I. Crother (269, 335), *Department of Biology, Southeastern Louisiana University, Hammond, Louisiana 70402*

William E. Duellman (357), *Division of Herpetology, Museum of Natural History, University of Kansas, Lawrence, Kansas 66045*

Alberto R. Estrada (31), *Department of Biology, University of Puerto Rico, San Juan, Puerto Rico 00937*

Robert W. Henderson (223), *Section of Vertebrate Zoology, Milwaukee Public Museum, Milwaukee, Wisconsin 53233*

Sixto J. Inchuastegui (93), *Grupo Jaragua, El Vergel, Santo Domingo, Dominican Republic*

Hinrich Kaiser (181), *Institut für Humangenetik, Universität Würzburg, Biozentrum, am Hubland, 97074 Würzburg, Germany*

José A. Ottenwalder (93), *Conservation and Management of Biodiversity Project, United Nations Development Programme, Santo Domingo, Dominican Republic*

Robert Powell (93, 223), *Department of Natural Sciences, Avila College, Kansas City, Missouri 64145*

Gregory K. Pregill (335), *Department of Biology, The University of San Diego, San Diego, California 92110*

Rudolfo Ruibal (31), *Department of Biology, University of California, Riverside, California 92521*

Richard Thomas (169), *Biology Department, University of Puerto Rico, Rio Piedras, Puerto Rico 00931*

Ernest Williams[†] (1), *Museum of Comparative Zoology, Harvard University, Cambridge, Massachusetts 02138*

[†]Deceased.

Foreword¹

In honor of Albert Schwartz.

“Just when I get you to a point that you can intelligently discuss biology, you are gone!” That or a similar statement was Al’s common lament. However, if you listened carefully, it was said with a note of pride. Al never had a graduate student and more than half of his teaching career was at a two-year college. Yet his imprint as an intellectual mentor is borne by dozens, perhaps hundreds, of us—biologists, medical doctors, dentists, architects, librarians, to name a few professions of his students.

As herpetologists, we cannot fail to recognize Al’s legacy to West Indian Herpetology, even though there were some of us who mumbled disparaging remarks about his constant description and splitting taxa. Whereas James Bond provided the foundation for ecological and behavioral studies of the bird fauna, Al’s studies provided an even broader framework for the study of amphibians and reptiles; his published studies and his voucher collections serve us and future generations of systematic herpetologists. Al’s meticulous attention to details make his collections and publications gold mines of information. There was nothing casual about his collecting; he was intent upon obtaining large samples and geographically closely spaced so that he could better discern speciation through an intimate knowledge of geographic variation. This understanding was gained by intense examination, a remarkable memory for details, and an ability to bring all data-bits into a coherent interpretation. He relied little on statistical analysis; his “taxonomic eye” was amazingly capable. Similarly, his herpetofaunal surveys are models of how biodiversity research needs to be pursued—thorough and repeated sampling (with retention of voucher specimens) of a geographic area and PROMPT study and publication of the results.

As herpetologists, many of you are unaware of Al as a student magnet. Oh yes, at the beginning of each school term, he would grumble something about staring out at all those blank faces. He would then “gird his loins” and like a gladiator march into the lecture arena to challenge

¹This contribution is a U.S. government work in the public domain.

those faces and minds to think. An imposing and demanding teacher, he excited your imagination, largely I believe through his enthusiasm for biology. For some of us, this enthusiasm attracted us to his lab and research. For me, he shook me out of my sophomore lethargy and impending dropout status. Amphibians and reptiles had had little fascination for me but that was to change and stay changed. I was neither the first nor the last to be so challenged and invigorated by Al. However, I may be unique in being his only academic student to retain a herpetological focus. But of his Albrightian students of the fifties, I was not the only one to owe my continuing fascination with biology to Al. Several of my dentist and M.D. classmates continued to join him in his West Indies research visits, until Al's legs could no longer tolerate field work.

Owing to the thriftiness of an uncle, Al was able to escape the northern winters in 1960 and, to the best of my knowledge, never again ventured north of Florida. His shift to Miami gave him ready access and importantly short flight time to the Antilles. For a number of years, he devoted himself entirely to field work and writing. Then among the student revolts of the late sixties, he returned to full-time teaching at Miami-Dade Community College and part time at Florida Atlantic University, again sharing his enthusiasm for biology with students, some who would continue in biology but most who would not. This book is an intellectual tribute from Al's herpetological colleagues. If I may be a bit presumptuous, I offer Al a tribute of appreciation and love from his students.

September 1997

George R. Zug, Department of Vertebrate Zoology,
National Museum of Natural History, Washington DC 20560

Preface

In a small token of appreciation for Dr. Schwartz's help and friendship, and in recognition of his incredible scientific accomplishments, this book reviews herpetology in the Caribbean. A letter was sent to Dr. Schwartz informing him of the plans and to ask him if he would like to write a short autobiography (a true biography of Dr. Schwartz would be a book unto itself). He never saw the letter. He never knew a tribute was in the offing. The letter had been mailed on a Friday and Dr. Schwartz peacefully passed away that weekend. I somehow suspect he now knows and I hope that he is pleased.

This book presents an overview of the herpetology of the West Indies–Caribbean region. Authoritative reviews have been prepared by herpetologists who have worked extensively in the Caribbean. Chapters cover particular islands and amphibian and reptile ecology, phylogeny, and biogeography. The final chapter is a perspective that places the West Indian herpetofauna in relation to other geographic regions.

The islands of the Caribbean continue to lure scientists interested in testing hypotheses on the manageable ecosystems (not to mention the balmy climate, beaches, rum, etc.). In 10 years another review of Caribbean herpetofauna could include separate chapters on ethology, physiology, conservation biology, and micro-evolution at the molecular level, as well as entirely new chapters on phylogeny, biogeography, and ecology. The latter three, while receiving much attention here, are far from well understood. Only a small fraction of the taxa have been examined in explicit phylogenetic studies, and of those, only a few can be considered well corroborated (stable). To add to the difficulty in estimating stable phylogenies, it seems that more species remain to be discovered and described, as exemplified by continued discoveries of Blair Hedges, Richard Thomas, and Orlando Garrido. Biogeographic explanations, both historical and ecological, remain in debate.

This incomplete understanding of the evolutionary history of the taxa and the areas has naturally resulted in differences in opinion on matters of taxonomy and origins of the herpetofauna in the West Indies. Even the casual reader will note differences among the authors in this book. Empirically, it can be argued that much of this is due to the lack of stable phylogenies (and consistent philosophies), and with increasing numbers

of stable phylogenies, differences of opinion on taxonomy and even origins should dwindle.

As for general ecological work, much has been accomplished (see Chapter 7), but far more needs to be done, especially at the most fundamental level of natural history. The natural history of the vast majority of the islands' herpetofauna remains completely unknown, or at best based on museum ecology (stomach contents studies). Given these gaps, I hope information will continue to accumulate at a high rate over the next decades.

Herpetology in the Caribbean region remains in a dynamic state. Opportunities for research on the West Indian herpetofauna abound, but rapid losses of forests and undisturbed areas in these natural laboratories are of great concern. So to all current and future researchers of the island herpetofauna, and to those engaged in preserving the beauty and nature of the islands, godspeed.

Brian I. Crother

Acknowledgments

EDITOR

In 1985 I had decided upon a dissertation project that required field work throughout the West Indies. To help the project along, my major professor, Jay Savage, arranged a meeting for me with Dr. Albert Schwartz. I was on time (*that* was important) for my meeting at Dr. Schwartz's home/library/lab/collection. He showed me his collection (I had no idea he kept such a collection in his garage), and then we sat in his library to talk. Throughout the conversation I prodded him about helping me contact Richard Thomas for help with field work. Finally, he stopped me and said, "Is there something wrong with me?" At first I had no idea what he was talking about and then it sunk in that I had essentially been slowly sticking both feet in my mouth. Here was the most famous Caribbean herpetologist to have ever lived sitting before me, kindly talking to a green graduate student/budding scientist, and I never thought to ask *him* for advice, help, etc.! The embarrassment of the moment will never be forgotten, but I will also never forget the tremendous help he provided over the years. And so, I gladly acknowledge Dr. Schwartz's (Al's) friendship and advice as the impetus for editing this volume in his honor.

Because of the goal and format of this volume, I had to solicit specific topics from the contributing authors: a proposition that turned out to be challenging. I am greatly indebted to the contributing authors for their expertise. My gratitude for their understanding and patience is inexpressible: it made this volume a reality. Jonathan Campbell, Charles Crumly, Roy McDiarmid, and Rich Seigel all responded to my questions concerning advice on assembling this volume. I am most grateful, even if I neglected to heed it.

Charles Crumly and Academic Press are thanked for publishing this book. Charles Crumly proved most patient and persistent throughout the project.

A number of colleagues provided reviews of chapters (some multiple chapters): Darrel Frost, Craig Guyer, Robert Henderson, Jonathan Losos, Thomas Ostertag, Kirsten Nicholson, Gregory Pregill, Phil Stouffer, Travis Taggart, Mary White. Their generosity of time and expertise increased the quality of the work herein.

Mary White showed great understanding and support of this project, even when it clearly conflicted with other research. I could not have made this volume a reality without her. “Thank you” is not quite enough a reciprocation. I am in her debt.

CHAPTER 1. History (Williams)

During the preparation of this volume, the most esteemed contributor to the work, Ernest Williams, passed away. I, and hopefully everyone who reads it, will cherish the experience and wisdom to be found in his chapter. Dr. Williams left no acknowledgments, but I do understand from a letter to me that John Cadle read and commented on his paper.

CHAPTER 2. Cuba (Estrada and Ruibal)

We thank Rafael de la Vega for the assistance he provided with publications and information that were crucial for the completion of this article.

CHAPTER 3. Jamaica (Crombie)

In the course of my Jamaican field work I enjoyed the company of many individuals who contributed materially to the success of our ventures in many ways. Alphabetically they are Frank Davis, Dick Franz, Linda (Crout) Gordon, Susan Hansen, Barb Harvey, Blair Hedges, Bill Hilgartner, George Jacobs, Alan Jaslow, Fran McCullough (now Irish), Dyer Moore, Greg Pregill, Miles Roberts, Dave Steadman, Richard Thomas, Fred Thompson, and Bob Tuck. Special appreciation is due Jeremy F. Jacobs, who was along on nearly every trip even though there are no salamanders in Jamaica. He endured the long hikes through the Cockpits and an even more perilous stroll through downtown Kingston, demonstrating great dedication and endurance, if not much common sense.

In Jamaica we received cooperation from and imposed on the hospitality of a great many people. Patrick Fairbairn and Ann Haynes (Department of Natural Resources, Kingston) facilitated our permits and made our visits to Kingston much more endurable. Tom Farr (Institute of Jamaica), Ivan Goodbody (University of the West Indies, Mona), and Jeremy Woodley (UWI and later Discovery Bay Marine Lab) helped in too many ways to list. For hospitality above and beyond the call of duty, Menocal Stephenson and family of Quick Step, the Sutton family of Marshall's Pen, and Patrick Tennison of Windsor deserve special mention. Early in my field work, Tom Jenssen and his family provided much-needed sanctuary in Mandeville, for which I am most grateful. The management and staff of several hotels tolerated many strange beasts (including us) in their rooms with equanimity if not enthusiasm. Most notable are the Malvern in Montego Bay, Mayfair Guest House in Mandeville, and the Port of Call in Black River.

I have swapped notes, data, and general chat on Jamaica with so many people that I apologize to any I have neglected to mention here, but those that distinguish themselves in my admittedly feeble mind are Kurt Auffenberg, Alex Cruz, Dick Franz, Les Garrick, Glen Goodfriend, Blair Hedges, Jeff Lang, Ross MacPhee, Gary Morgan, Storrs Olson, Greg Pregill, Al Schwartz, Dave Steadman, Richard Thomas, Becki Thompson, and Fred Thompson.

Brian Crother has demonstrated both the patience of Job and an unexpected streak of masochism in editing this volume, a commendable if puzzling combination. Thanks dude, but I still can't believe you wouldn't buy the story about evil space aliens stealing my manuscript.

I dedicate this chapter to all those who "went before" in Jamaica, with thanks for the contributions that made this summary possible. I am particularly grateful that I was able to meet and swap Jamaica stories with Gardner Lynn, Al Schwartz, Garth Underwood, and Ernest Williams.

Last but not least, I express my appreciation and admiration to George Proctor, botanist of the Institute of Jamaica, who I met briefly in Kingston many years ago. No matter where I went in Jamaica, from the deepest Cockpits to the peaks in the Blue Mountains, Dr. Proctor had been there first and was fondly remembered by everyone I encountered. I can only hope I have done half as well, George.

CHAPTER 4: Hispaniola (Powell *et al.*)

We thank Brian I. Crother for the invitation to contribute to this volume. The late Albert Schwartz helped each of us immeasurably and on numerous occasions over the years. Robert W. Henderson, S. Blair Hedges, and Brian I. Crother commented critically on early drafts of this chapter and kindly provided access to considerable data and unpublished manuscripts; without their help this chapter could not have been written. Kraig Adler helped with citations to the historical literature. John S. Parmerlee, Jr., assisted with figures. Directors and staff at the Departamento de Vida Silvestre and the Dirección Nacional de Parques have been supportive and helpful and have made available the permits necessary for us to engage in our studies. Andreas Schubert and Gloria Santana, both of the Departamento de Vida Silvestre, have facilitated opportunities for field work by R.P. in the Dominican Republic. We also thank the many colleagues and students who have accompanied us in the field over the years.

Powell's field work was supported by Avila College and via funding by a series of National Science Foundation Research Experiences for Undergraduates (REU) programs.

CHAPTER 6: Lesser Antilles (Censky and Kaiser)

We thank Gregory Pregill for his comments on our manuscript. Kaiser's Caribbean research was funded by grants from the Hans-Krueger-

Stiftung and the Studienstiftung des Deutschen Volkes, by Boehringer Ingelheim Fonds, and by the Deutsche Forschungsgemeinschaft.

CHAPTER 7: Ecology (Henderson and Powell)

We thank Brian Crother for inviting us to write this chapter, and Craig Guyer and Jonathan Losos for constructive, thought-provoking reviews. Blair Hedges and Hinrich Kaiser answered many questions about West Indian frogs, and Susan Otto (librarian, Milwaukee Public Museum [MPM]) was instrumental in procuring literature. We also are indebted to the many kind officials on many islands who granted permission to conduct research in their respective nations.

Funding for Henderson's field work in the West Indies came from a variety of sources: MPM, Zoology Trust Fund (MPM), the late Albert Schwartz, Institute of Museum Services, the Central Florida Herpetological Society, the Chicago Zoological Society, Gulf + Western Dominicana, and the Museo Nacional de Historia Natural in Santo Domingo.

Powell's field work has been made possible largely through opportunities presented by teaching a series of field biology classes at Avila College and via funding provided by the National Science Foundation Research Experience for Undergraduates (REU) Program.

CHAPTER 8: Phylogeny (Crother)

I thank the several people who responded to my inquiries about various groups: Aaron Bauer, James Dixon, Carl Gans, Ronald Heyer, Ulrich Joger, Ulrich Kuch, Jonathan Losos, John Lynch, William Presch, Steven Werman, and Wolfgang Wüster. I thank Tom Ostertag, Travis Taggart, Kevin Toal, and Brian Warren for making many library and copying runs. I am especially grateful to Travis Taggart for making all the figures. Mary White kindly read, edited, and discussed drafts of the manuscript. The research was partly funded by the National Science Foundation (Award DEB-9207751).

CHAPTER 9: Biogeography (Pregill and Crother)

We extend our gratitude to George Zug and Ronald Crombie for their hospitality and free access to the herpetological resources of the U.S. National Museum. We also thank Mary White and David Steadman for their insightful comments on the manuscript.

CHAPTER 10: Perspective (Duellman)

I am grateful to Brian I. Crother for inviting the manuscript and thereby providing me an opportunity to express my views on the subject. I am indebted to B. I. Crother, C. Guyer, and S. B. Hedges for sharing with me copies of manuscripts that they had submitted elsewhere and to J. A. Campbell for the same courtesy, as well as providing up-to-date

information on the herpetofauna of Guatemala. But foremost, I am indebted to the late Albert Schwartz for many inspiring discussions; although we frequently entertained the idea of each of us accompanying the other to the West Indies and to Middle America, we never did so. What a pity!

This Page Intentionally Left Blank

Legends to Color Plates*

The phenomenal diversity of the Antillean herpetofauna precludes illustrating even a small fraction of the many species of amphibians and reptiles worthy of inclusion. Instead, Plates 2–6 were designed to provide a glimpse of the total picture through a sampling of representative types from each of the major regions covered in this volume. Habitats illustrated in Plate 1 were chosen also as representative of the region, and we purposefully selected in nearly every instance habitats altered by human activities, both because these represent the reality of the modern era and because these tend to be less frequently illustrated than the increasingly rare and fragmented pristine environments that often attract the attention of biologists working in the region. In Plate 7, we illustrate diagrammatically the ecomorph concept as applied to Greater Antillean anoles, using representative species that also expand the coverage of diversity presented in previous plates. Plate 8 illustrates species whose very existence has become tenuous due to human activities in the region, although only some of the taxa included are officially recognized as “rare,” “threatened,” or “endangered.”

Captions list information about habitats and specimens illustrated, credits the photographers who so graciously provided slides, and summarize some pertinent aspects of natural history, with much of the latter extracted from Schwartz and Henderson (1991).

Plate 1: Representative Antillean Habitats. Unfortunately, few West Indian habitats remain pristine. Although hurricanes and other natural events often wreak havoc, it is primarily the need to sustain the rapidly growing human populations and the accompanying exploitation of resources, introduction of alien species, and alterations of all but the most “worthless” areas that have converted the islands into a very different reality than that experienced by the first Europeans over 500 years ago. “With this in mind, biologists must be willing to examine interactions in severely altered habitats (Powell *et al.*, 1996a), contrary to our inclination to focus on the remaining pristine areas” (Henderson and

*Compiled by Robert Powell.

Powell, Chapter 7). (A). Cactus scrub northeast of Gonaïves, Haiti. The removal of woody plants for charcoal production has resulted in many areas being dominated by cacti; also note the deforested hillside in the background. (Insert) Impenetrable cactus scrub near Lago Enriquillo, Provincia de Baoruco, Dominican Republic (photograph by Thomas A. Jenssen; insert by Robert Powell). (B) A recent (1995) clear-cut through formerly virgin cloud forest at Loma Remigio, Sierra de Baoruco, Dominican Republic. Even pristine stands of primary forest are vulnerable to clearing for subsistence agriculture and charcoal production (photograph by Robert Powell). (C) Damage on Guadeloupe from Hurricane Hugo, September 1989 (photograph taken March 1990 by Richard A. Sajdak). (D) The Cockpit Country in Trelawny Parish, Jamaica. To date, such “inhospitable” areas have largely escaped extensive development, but growing human populations are intruding on these areas to greater and greater extents [see, for example, comments by Vogel *et al.*, (1996) on the Hellshire Hills] (photograph by Richard A. Sajdak). (E) Secondary forest mixed with cultivated coconut, mango, cacao, and banana trees at Westerhall Estaste, St. David Parish, Grenada. This composite habitat is extensively exploited by *Corallus grenadensis* (Henderson *et al.*, 1996) (photograph by R. Allan Winstel). (F) The dry, high-elevation pine forests of the Sierra de Baoruco, Provincia de Barahona, Dominican Republic. These pine forests are almost absent from Haiti and many Dominican stands are being destroyed by fire, both inadvertent and intentional (photograph by Richard A. Sajdak). (G) Mangrove swamp near Cabo Rojo, Puerto Rico. Mangrove forests throughout the Antilles are disappearing in order to expose beaches for tourist developments (photograph by Thomas A. Jenssen). (H) Sea grape beach at Cabo Rojo, Puerto Rico. Many beaches used historically by sea turtles and a number of terrestrial lizards have become devoid of reptiles largely as the result of tourist development (photograph by Thomas A. Jenssen).

Plate 2: Selected Amphibians and Reptiles of Cuba. “. . . today’s Cuba still treasures much of its original, wild nature. More, without a doubt, than any other Caribbean island. The *mogotes* and steep-sided mountains, swamps and distant or inaccessible estuaries, the scattering of low and muddy keys, and the desert of the southeastern region have proven hard to surmount barriers to settlement so far. More accessible highlands were indeed invaded, a long time ago. Their main use, however, has been coffee and cacao cultivation, which requires deep shadows: This has allowed them to survive dressed up like a densely packed arboretum. The tall vegetation attenuates the impact of rainwater, provides havens to many minor life-forms, and pleases the eye” (Silva Lee, 1996). (A) *Bufo* (formerly *Peltophryne*) *peltoccephalus* from Guantanamo Bay, Provincia de Guantánamo. One of seven native Cuban toads, this

species is at home in a wide variety of habitats. Males call from April through December from flooded ditches and slowly moving streams (photograph by Richard A. Sajdak). (B) *Eleutherodactylus atkinsi* from Guantanamo Bay, Provincia de Guantánamo. Although largely mesophilic, this widely distributed species, one of more than nearly 40 native Cuban species in this genus, is sufficiently opportunistic to utilize a variety of specific habitats. Males usually call from ground litter or vegetation up to 1.5 m high (photograph by Richard A. Sajdak). (C) *Anolis* (formerly *Chamaeleolis*) *chamaeleonides* from the Península de Guanahacabibes, Provincia de Pinar del Río. These slow-moving lizards rely on crypsis and are also like true chamaeleons in that they can move their eyes independently (photograph courtesy of the late Albert Schwartz). (D) *Anolis smallwoodi* from Guantanamo Bay. One of about 55 native Cuban anoles, this crown giant is usually found at height ≥ 3 m but will descend to the ground on occasion (photograph by Richard A. Sajdak). (E) *Leiocephalus carinatus*, from Guantanamo Bay, Provincia de Guantánamo. One of six endemic Cuban species in this genus, this xerophilic sit-and-wait forager is common in xeric coastal areas. Functionally omnivorous, these lizards frequently consume flowers, buds, fruits, and seeds along with smaller lizards and a variety of invertebrates, including large quantities of ants (photograph by Richard A. Sajdak). (F) *Alsophis cantherigerus* from Guantanamo Bay, Provincia de Guantánamo. Like many of the other 11 endemic West Indian species, the Cuban Racer has become considerably less common after introduction of the mongoose. Adults are quite catholic in diet, consuming lizards, frogs, birds, mammals (including bats), and even turtles (photograph by Richard A. Sajdak). (G) *Epicrates angulifer* from Guantanamo Bay, Provincia de Guantánamo. This species is the largest West Indian snake, with snout-vent lengths (SVL) approaching 4 m, and there are unconfirmed reports of much larger specimens. The diet changes ontogenetically from lizards to birds and mammals (photograph by Richard A. Sajdak). (H) *Tropidophis melanurus* from Guantanamo Bay, Provincia de Guantánamo. One of nine native species in this genus, these snakes feed primarily on frogs and lizards, but will take birds and small mammals on occasion (photograph by Richard A. Sajdak).

Plate 3: Selected Amphibians and Reptiles of Jamaica. “The list of Jamaican species is by no means complete, however, and the rugged limestone terrain continues to hide surprises and make a mockery of “definitive” statements on the rarity or even extinction of some species” (Crombie, Chapter 3). (A) *Osteopilus brunneus* from near Clarkstown, Trelawney Parish. As in the other four endemic species of Jamaican hylids, both terrestrial and arboreal bromeliads are used for daytime retreats, calling sites, egg deposition, and larval development (photograph

by Karen Anderson). (B) *Calyptahyla crucialis* (possibly *Osteopilus*; Hedges, 1996b; see also Chapter 3) from a limestone hillock at Marshall's Pen, Mandeville Parish (700 m). These large tree frogs occur in mesic broadleaf forests. Males usually call late at night, after other frogs. Eggs and tadpoles are in bromeliads (photograph by Jeffrey W. Lang). (C) *Hyla wilderi* (possibly *Osteopilus*; Hedges, 1996b) from Barbecue Bottom, Trelawney Parish. These frogs inhabit mesic woodlands where they are frequently associated with bromeliads, in which eggs are laid and tadpoles develop (photograph by Karen Anderson). (D) *Anolis grahami* from Discovery Bay, St. Ann Parish. This trunk–crown anole, one of eight native Jamaican species, has taken full advantage of habitats provided by humans, frequenting buildings, walls, and fences. The diet is dominated by invertebrates but may occasionally include smaller anoles (photograph by Jonathan B. Losos). (E) *Sphaerodactylus semasiops* from the karst regions in the Cockpit Country. These diminutive lizards, representing one of seven native Jamaican species, are associated with bromeliads on trees, rocks, or the ground (photograph by Richard Thomas). (F) *Sphaerodactylus parkeri* from Clarendon near Freetown. These generally mesophilic lizards may survive habitat alterations rendering areas more xeric. Individuals have been found in association with caves (photograph by R. G. Tuck, Jr. Courtesy of the National Museum of Natural History). (G) *Celestus cruscus* from Montego Bay, St. James Parish. Like *Sphaerodactylus parkeri*, these mesophiles may be able to tolerate altered xeric conditions. Like most Antillean lizards, these anoles are primarily insectivorous (photograph by Ronald I. Crombie. Courtesy of the National Museum of Natural History). (H) *Typhlops jamaicensis* from Worthy Park. These scolecophidian burrowers are associated with a variety of habitats, including termitaria (photograph by R. G. Tuck, Jr. Courtesy of the National Museum of Natural History).

Plate 4: Selected Amphibians and Reptiles of Hispaniola. Once described by Albert Schwartz as an “island of islands,” Hispaniola is a mosaic of mountain ranges separated by rain-shadowed lowlands (see Chapter 4). Its known diversity is astounding and new discoveries are ongoing. (A) *Hyla heilprini* from along the road from Jarabacoa to Manabao, Provincia de La Vega, Dominican Republic. One of only four endemic Hispaniolan hylids, this species is associated with rapidly flowing or torrential streams in mesic broadleaf forests, and the tadpoles have thick, short, muscular tails (photograph by Karen Anderson). (B) *Anolis* (formerly *Chamaelinorops*) *barbouri* from the Sierra de Baoruco, Provincia de Barahona, República Dominicana. An unusual and exceedingly cryptic terrestrial anole, these lizards do not bask in their deeply shaded forest habitats, but nevertheless maintain above-ambient body temperatures (photograph by Thomas A. Jenssen). (C) *Anolis whitemani* from

Cayo Monte Grande, Siete Hermanos, Provincia de Monte Cristi, Dominican Republic. This trunk-ground anole is the most xerophilic member of the cybotoid group and typically is found in desert scrub. Animals from the northernmost part of the species' range are unusual in having yellow dewlaps (photograph by Robert Powell). (D) *Anolis bahorucoensis* from near Paraiso, Sierra de Baoruco, Dominican Republic. One of about 40 native Hispaniolan species, these lizards represent a variant of the bush-grass ecomorph. The apparently bright pattern effectively serves to break up the lizards' outlines in the shaded mesic forests where they occur. The dewlap is diminutive and even males display only rarely (photograph by Robert W. Henderson). (E) *Sphaerodactylus ladae* from arid foothills of the Sierra Martín García, Provincia de Barahona, Dominican Republic. Most specimens of this poorly known species have been taken under dead *Agave* in *Agave*-cactus associations. About 30 endemic Hispaniolan species of *Sphaerodactylus* are known (photograph by James D. Forester). (F) *Leiocephalus schreibersii* from near Monte Cristi, Provincia de Monte Cristi, Dominican Republic. One of 10 endemic Hispaniolan species, this terrestrial sit-and-wait forager effectively regulates its body temperature by shuttling between sun and shade (photograph by Thomas A. Jenssen). (G) *Uromacer catesbyi* from Santo Domingo, Distrito Nacional, Dominican Republic. One of three endemic Hispaniolan "vine snakes" that are highly arboreal but willingly and frequently forage on the ground. These snakes feed primarily on *Osteopilus* and anoles but will take other frogs and lizards as opportunities arise. Generally inoffensive, one reported bite from one of these rear-fanged snakes produced an instant "burning" pain that rapidly subsided (photograph by Robert Powell). (H) *Epicrates striatus* from near Paraiso, Sierra de Baoruco, Provincia de Barahona, Dominican Republic. The largest Hispaniolan snake with maximum known SVL of 233 cm, these snakes inhabit a variety of habitats but are most common in mesic forests. As in many of its West Indian congeners, the diet shifts ontogenetically from lizards and frogs to birds and mammals (photograph by Suzanne Collins. Courtesy of the North American Center for Amphibians and Reptiles).

Plate 5: Selected Amphibians and Reptiles of the Puerto Rico Bank. "Of the 80 or so species in the entire area, 57 occur on Puerto Rico, and, of these, 37 are confined to that island and its immediate offshore cays. As one progresses east of Puerto Rico into the Passage Islands and the Virgin Islands, there is a marked drop-off in species, no doubt due largely to the reduction in the area of the eastern islands but also due to the absence of high-elevation, moist habitat" (Thomas, Chapter 5). (A) *Eleutherodactylus coqui* from El Verde, Río Grande. Named for its distinctive two-note call, this little frog has become a "mascot" of Puerto Rico. Males call from elevated perches and may "parachute"

to the ground (Stewart, 1985). Nearly 20 species of *Eleutherodactylus* are known from the Puerto Rico Bank (photograph by Pamela T. Lopez). (B) *Eleutherodactylus locustus* from Pico del Este, El Yunque. Diurnal retreats of this frog are primarily underground litter. Males call from elevated perches (<1.5 m) on vegetation. At El Yunque, where *E. locustus* is sympatric with *E. eneidae*, the latter calls later at night than elsewhere in its range. Rivero (1978) suggested that this shift may have resulted from acoustic competition with *E. locustus*, which has a similar call (photograph by Rafael L. Joglar). (C) *Sphaerodactylus nicholsi* from Isabela. One of about a dozen native Puerto Rican Bank species, these xerophilic geckos are frequently found on beaches where they hide in *Cocos* trash in relatively exposed areas (Schwartz and Henderson, 1991) (photograph by Manuel Leal). (D) *Anolis cristatellus* from Aguirre Forest Reserve, Guayama. This common trunk-ground anole favors sunnier perches than most of the nearly 20 native species of Puerto Rican anoles. Introduced into the Dominican Republic, these lizards have displaced the ecologically similar *A. cybotes* in urban areas (Fitch *et al.*, 1989) (photograph by Manuel Leal). (E) *Anolis stratulus* from El Verde, Río Grande. One of the few trunk-crown anoles that are not primarily green, these well-camouflaged lizards remain primarily on the trunk and seldom venture onto smaller branches and leaves; however, they do wander onto the ground more frequently than most trunk-crown ecomorphs. Relying heavily on crypsis, mean approach distance is 50 cm (photograph by Manuel Leal). (F) *Diploglossus pleii* from Río Abajo Forest Reserve, Arecibo. The only anguid known from Puerto Rico, these small, mesophilic lizards (maximum known SVL, 125 mm) give live birth to two to four young (Greer, 1967) (photograph by Manuel Leal). (G) Hatchling *Epicrates inornatus* from Río Grande. This ecologically versatile snake occupies a variety of habitats and is "not uncommon in urban and suburban areas" (Schwartz and Henderson, 1991). Individuals have been known to aggregate at the mouths of caves in order to capture emerging bats (Rodríguez and Reagan, 1984) (photograph by Rafael L. Joglar). (H) *Arrhyton exiguum* from Cerro de las Cuevas. The presence of this genus on the Puerto Rico Bank as well as on Cuba and Jamaica lends credence to the possibility that Hispaniolan *Darlingtonia* should be included in *Arrhyton*. These small active foragers prey heavily on *Eleutherodactylus* (eggs, hatchlings, and adults), *Sphaerodactylus*, and *Anolis* (photograph by Howard W. Campbell. Courtesy of the National Museum of Natural History).

Plate 6: Selected Amphibians and Reptiles of the Lesser Antilles.

These islands form two distinct arcs, the inner of which still has active volcanos. Diversity varies considerably from island bank to bank and is

related to island size, diversity of habitat, and distance from the mainland or another large island (see Chapter 6). (A) *Eleutherodactylus barlagnei* from Guadeloupe. One of the many endemic West Indian species in this most diverse of all vertebrate genera, this aquatic species is associated with boulder-strewn mountain torrents (Schwartz, 1967) (photograph by Richard A. Sajdak). (B) *Eleutherodactylus martinicensis* from Dominica. This ecologically diverse species often is exceedingly common in a variety of habitats, including many that have been substantially altered. Nevertheless, some populations have been adversely affected by the recent introductions of *E. johnstonei* on some islands (photograph by Richard A. Sajdak). (C) *Anolis marmoratus* from Capesterre-Belle-Eau, Guadeloupe. These habitat generalists exhibit remarkable pattern diversity in on Guadeloupe and its satellites (Lazell, 1964a) (photograph by Richard A. Sajdak). (D) *Anolis sabanus* from Saba. The distinct disruptive color pattern of this highly saxicolous lizard (Lazell, 1972) is very effective (photograph by Richard A. Sajdak). (E) *Ameiva griswoldi* from Great Bird Island, Antigua. These actively foraging terrestrial lizards are less common on the main island where the mongoose is quite common. In contrast, individuals on Great Bird Island are almost tame (R. W. Henderson, personal communication) (photograph by Richard A. Sajdak). (F) *Corallus grenadensis* from Mt. Hartman (sea level), Grenada. These snakes exhibit the highest population densities in disturbed habitats but require contiguous canopies (Henderson *et al.*, 1996) (photograph by Robert W. Henderson). (G) *Sphaerodactylus elegantulus* from Barbuda. Eggs of this "dwarf gecko" have been found in moist debris under palm fronds and rocks (Schwartz and Henderson, 1991) (photograph by Richard A. Sajdak). (H) The fate of *Alsophis antillensis manselli*, from Montserrat, may not be determined for some time after the cessation of current pyroclastic events (photograph by Richard A. Sajdak).

Plate 7: Ecomorphs of Greater Antillean *Anolis* Lizards. Ecomorphs are species of different phyletic origins with similar morphological adaptations to similar niches (Williams, 1983). Crown giants (A, B) are large (SVL >100 mm), generally green, sometimes patterned lizards with large, often casqued heads and vertebral crests. These active and aggressive foragers are generally found high in the canopies of large trees. Twig dwarfs (C, D) are small (<50 mm), gray, or lichenate anoles with long heads and short bodies and limbs. Slow "crawlers," these lizards forage on small-diameter twigs in the canopy, relying on crypsis and immobility to protect them from predators. Trunk-crown anoles (E, F) are moderately large (>70 mm), generally green lizards with large heads, long bodies, and proportionately short limbs. Foraging on leaves

and branches in the canopy and along the upper portion of the trunk, these lizards almost invariably flee upward when threatened. Trunk anoles (G) are small (<50 mm) and range from green to variously patterned with browns and/or grays. Head and body are both short. Foraging almost exclusively on the trunk below perches favored by trunk-crown and above those frequented by trunk-ground ecomorphs, these lizards “squirrel” (run around to the opposite side of the trunk) when threatened. Trunk-ground anoles (H, I) are of moderate size (>60 mm) and have relatively short heads, short, stout bodies, and long limbs. These lizards typically face downward from perches low on tree trunks, searching for prey, often on the ground. When threatened, they will usually flee downward. Grass-bush anoles (J, K) are small (<50 mm), slender lizards with relatively long heads and very long tails. Although ground colors vary, patterns inevitably include lateral and/or dorsal stripes. These adept jumpers forage in bushes and grasses, fleeing toward the ground to denser vegetation when threatened. Additional habitat partitioning occurs between sun- and shade-tolerant species within ecomorph assemblies. (A) *Anolis cuvieri*, a crown giant from Puerto Rico (photograph by Jonathan B. Losos). (B) *Anolis luteogularis*, a crown giant from Cuba (photograph by Jonathan B. Losos). (C) *Anolis angusticeps*, a twig anole from Cuba and the Bahamas (photograph by Jonathan B. Losos). (D) *Anolis insolitus*, a twig anole from Hispaniola (photograph by Jonathan B. Losos). (E) *Anolis porcatius*, a trunk-crown anole from Cuba (and introduced to Hispaniola) (photograph by Robert Powell). (F) *Anolis evermanni*, a trunk-crown anole from Puerto Rico (photograph by Jonathan B. Losos). (G) *Anolis caudalis*, a trunk anole from Hispaniola (photograph by Thomas A. Jenssen). (H) *Anolis cybotes*, a trunk-ground anole from Hispaniola (photograph by Jonathan B. Losos). (I) *Anolis mestrei*, a trunk-ground anole from Cuba (photograph by Jonathan B. Losos). (J) *Anolis olssoni*, a grass-bush anole from Hispaniola (photograph by Jonathan B. Losos). (K) *Anolis pulchellus*, a grass-bush anole from Puerto Rico (photograph by Jonathan B. Losos).

Plate 8: Some Threatened and Endangered Amphibians and Reptiles of the West Indies. “West Indian environments have been subjected to the influence of nonnative Americans, almost always negatively, longer than any other land masses in the New World. Habitat destruction is rampant and forests are rapidly disappearing in order to produce charcoal and to plant crops; Haiti, admittedly the most extreme example, retains less than 1% of its original forest cover. The introduction of the mongoose is correlated with the decline or disappearance of several snake taxa and populations; the entire known range of several species is now less than 0.5 km². The introduction of cats, dogs, and rats has had a

catastrophic effect on some *Cyclura* populations. Human numbers are booming and development (largely for the tourist trade) continues unabated. Although some versatile frogs and reptiles respond favorably to habitat modification, far more species fare less well” (Powell and Henderson, 1996a). (A) *Cyclura cornuta*, from Isla Cabritos, Provincia de Independencia, Dominican Republic. This large Hispaniolan iguanid has seen its populations fragmented and many extirpated by human exploitation for food, habitat destruction, and introduced predators and competitors. (Insert) The Mona Island population, *C. cornuta stejnegeri*, considered by some to represent a distinct species, is similarly threatened. Males compete for mating territories by means of highly formalized tests of strength (photograph by Robert Powell; insert by Manuel Leal). (B) *Cyclura nubila* from Guantanamo Bay, Cuba. The Cuban Iguana has been similarly exploited throughout its range. (Insert) *Cyclura collei*, the Jamaican Iguana, was thought to be extinct until rediscovered in the Hellshire Hills in 1970 (Vogel *et al.*, 1996). (photograph by Robert W. Henderson; insert by Peter Vogel). (C) *Eleutherodactylus richmondi* from Monte del Estado, Maricao, Puerto Rico. Populations of this and other forest-dwelling species have declined or have been extirpated at El Yunque, although this area is considered to be the “best-conserved forest reserve in Puerto Rico” (Joglar and Burrowes, 1996). Why *E. richmondi* is almost extinct at El Yunque, but are still present in the Sierra de Cayey and Cordillera Central, is difficult to explain. (D) *Alsophis antiguae sajdaki* is restricted to tiny Great Bird Island off Antigua (Henderson, 1990); the nominate subspecies has long been extirpated from the main island of Antigua (Henderson *et al.*, 1996a) (photograph by Richard A. Sajdak). (E) *Celestus* (formerly *Diploglossus*) *carrau*, from the Dominican Republic, is known from only three localities (Henderson, 1988a). Its Hispaniolan congener, *C. anelpistus*, has not been found since it was formally described in 1979 (Schwartz *et al.*, 1979; Henderson, 1988) (photograph by Richard A. Sajdak). (F) Carcass of a *Crocodylus acutus* from Lago Enriquillo, Provincia de Pedernales, Dominican Republic. This relict population has been historically exploited for food, hides, and pharmaceuticals (Schubert and Santana, 1996) (photograph by Andreas Schubert). (G) A *Chelonia mydas* awaiting slaughter near the beach of Isla Saona, Dominican Republic (photograph by Robert W. Henderson). (H) “Jicotecas” (*Trachemys stejnegeri*) from Laguna de Saladillo, Parque Nacional Montecristi, Provincia de Monte Cristi, Dominican Republic, being sold by a roadside vender. All of the endemic freshwater sliders of the West Indies are being heavily exploited for food (photograph by Robert Powell). (I) *Epicrates monensis* from Isla Mona, Puerto Rico. A number of populations have been extirpated and others are threatened by introduced predators, habitat destruction, and stochastic

processes resulting from population fragmentation and isolation (Tolson, 1996) (photograph by Peter J. Tolson). (J) The mongoose, *Herpestes javanicus*, is implicated in the extirpation of many West Indian species and populations of terrestrial snakes and lizards (Henderson, 1992) (photograph by Richard A. Sajdak).

Over 300 Years of Collecting in the Caribbean

Ernest E. Williams

*Museum of Comparative Zoology
Harvard University
Cambridge, Massachusetts 02138*

Introduction: Where We Are

With Schwartz and Henderson's massive 1991 compendium on the West Indian herpetofauna, an epoch primarily devoted to collection and description has come to an end. The goals will be quite different in the future.

In 1914 Thomas Barbour published a comparable work titled "A Contribution to the Zoogeography of the West Indies, With Special Reference to the Amphibians and Reptiles." His effort at zoogeography was, in fact, primitive and elementary, but he did provide an overview that was quite accurate in regard to the knowledge of its time. He included *Spelerpes infuscata* Peters, allegedly from Haiti, which was a bolitoglossine salamander that Dunn (1926a) would regard only as a badly mislabeled *Oedipus* [now *Lineatriton*] *lineolus* (Cope), a species from Mexico; although anomalous – no other salamander had ever been collected in the West Indies – it was a record attested to by Wilhelm Peters, Curator at Berlin, a very reputable source. He listed only 22 frogs: 17 *Eleutherodactylus*, six *Hyla*, six *Bufo*, one or two *Leptodactylus* (depending what is counted as endemic), and *Phyllobates limbatus*, so identified by Cope.

In lizards he listed 158 species, of which seven were *Mabuya*, and 73 anoles, one of which was *Chamaeleolis (chamaeleonides)*, one *Xiphocercus (valencienni)*, and one *Norops (ophiolepis)*. In non-anoline lizards he reported 29 *Sphaerodactylus*, 15 *Ameiva*, 11 *Leiocephalus*, nine *Celestus*, six *Cyclura*, three *Aristelliger*, a *Gymnodactylus* that is probably a mislabeled specimen, and a smattering of genera of which there was only one endemic example in the West Indies

(*Tarentola*, *Gymnophthalmus*, and *Cricosaura*). Of amphisbaenians he knew *Amphisbaena punctata* Bell. In snakes he was aware of six *Typhlops*, only two *Leptotyphlops*, eight *Epicrates*, six *Tropidophis*, and one "*Lachesis*." In colubrid snakes he recorded 37 species.

It is inevitable to compare this list of the West Indian herpetofauna with that of Schwartz and Henderson (1991) (Table 1.1). Their compendium is an accurate picture of the state of the art today. There are some problems of name changes, but these are remarkably few, except in snakes.

Counting only endemic species, there are a total of 152 species of frogs, 125 of them *Eleutherodactylus*; one of these is the single species supposed to represent a mainland genus, the *Phyllobates* of Barbour (1914). An odd story is concealed behind this apparent suppression of a genus and species. The species *limbatus* was discovered by Cope (1863a) and was referred by him to the genus *Phyllobates*. When Barbour and Noble (1920) in the course of a revision of *Phyllobates* discovered that the shoulder girdle of Cuban *limbatus* did not resemble the shoulder girdle of *Phyllobates* they erected the genus *Sminthillus* for it. Later, G. K. Noble (1921) and then H. W. Parker (1926a) erected mainland species, respectively, *peruvianus* and *brasiliensis*. Several authors (including Noble) considered the Cuban animal had eleutherodactyline affinities. Finally, Hedges (1989a) submerged the species in an expanded definition of *Eleutherodactylus* without examining the Cuban animal's shoulder girdle or preparing allozymes from it. He relied primarily on Bogart's statement that its karyotype resembled the *ricordii* group of *Eleutherodactylus* in referring it to that group and also on (his own character and observation) the shape of its liver. *Sminthillus peruvianus* had already been referred to *Phrynopus* by Heyer, as had *S. brasiliensis*. *Euparkerella* was erected by Griffiths (1959).

There are 294 species of lizards, and 128 of these are *Anolis* [*Eleutherodactylus* is close to overtaking *Anolis* as the most numerous genus of the West Indian herpetofauna, but despite new (post-Schwartz and Henderson) descriptions of species of both genera, I believe *Anolis* is still slightly ahead.] Two genera, *Xiphocercus* and *Norops*, have been synonymized with *Anolis* (Etheridge, 1960). (However, see Guyer and Savage, 1986 and the resulting literature: Canatella and de Queiroz, 1988; Williams, 1989a; and Guyer and Savage, 1992.)

One new endemic anoline genus, *Chamaelinorops*, has been recorded. This is ecologically (Flores *et al.*, 1994) and morphologically

Table 1.1. A Comparison of the Known Herpetofauna in the West Indies as Listed by Barbour (1914) and Schwartz and Henderson (1991).

Barbour (1914) Species	No.	Species	Schwartz and Henderson (1991) No.
Amphibia			
<i>Spelerpes infuscata</i> X See text			
<i>Hyla</i>	6	<i>Calyptahyla</i> ^a	1
		<i>Osteopilus</i> ^a	3
		<i>Hyla</i>	5
<i>Bufo</i>	6	<i>Peltaphryne</i> ^a	10
<i>Eleutherodactylus</i>	17	<i>Eleutherodactylus</i>	125
<i>Leptodactylus</i>	2	<i>Leptodactylus</i>	3
<i>Phyllobates</i> See text			
Sauria			
<i>Gymnodactylus fasciatus</i> See text			
<i>Gonatodes</i>	1	[as subspecies of <i>G. albogularis</i>]	
<i>Phyllodactylus</i>	1	<i>Phyllodactylus</i>	2
<i>Aristelliger</i>	3	<i>Aristelliger</i>	6
<i>Tarentola</i>	1	<i>Tarentola</i>	1
<i>Sphaerodactylus</i>	29	<i>Sphaerodactylus</i>	71
<i>Chamaeleolis</i>	1	<i>Chamaeleolis</i>	3
<i>Xiphocercus</i>	1	[now a synonym of <i>Anolis</i>]	
<i>Anolis</i>	70	<i>Anolis</i>	128
<i>Norops</i>	1	[now a synonym of <i>Anolis</i>]	
		<i>Chamaelinorops</i> ^a	
<i>Iguana</i>	1	<i>Iguana</i>	1
<i>Cyclura</i>	6	<i>Cyclura</i> ^a	8
<i>Leiocephalus</i>	11	<i>Leiocephalus</i> ^a	14
<i>Celestus</i>	9	<i>Celestus</i>	13
		<i>Diploglossus</i>	6
		<i>Sauresia</i> ^a	2
		<i>Wetmorena</i> ^a	1
<i>Ameiva</i>	15	<i>Ameiva</i>	18
		<i>Cnemidophorus</i>	1
<i>Gymnophthalmus</i>	1	<i>Gymnophthalmus</i>	1

<i>Mabuya</i>	7	<i>Mabuya</i> See text	1
<i>Amphisbaena</i>	7	<i>Amphisbaena</i>	10
		<i>Cadea</i>	2
Serpentes			
<i>Typhlops</i>	6	<i>Typhlops</i>	20
<i>Leptotyphlops</i>	2	<i>Leptotyphlops</i>	8
<i>Epicrates</i>	8	<i>Epicrates</i>	9
<i>Chironius</i>	1	<i>Chironius</i>	1
<i>Hypsirhynchus</i>	1	<i>Hypsirhynchus</i> ^a	1
<i>Ialtris</i>	1	<i>Ialtris</i> ^a	3
<i>Tretanorhinus</i>	1	<i>Tretanorhinus</i>	1
<i>Uromacer</i>	3	<i>Uromacer</i> ^a	3
[<i>Mastigodryas</i> as <i>Alsophis</i>]		<i>Mastigodryas</i>	1
		<i>Darlingtonia</i> ^a	1
<i>Alsophis</i>	15	<i>Alsophis</i> ^a	11
		<i>Antillophis</i> ^a	2
<i>Arrhyton</i>	3	<i>Arrhyton</i> ^a	10
<i>Leimadophis</i>	12		
		<i>Liophis</i>	5
<i>Lachesis</i>	1	<i>Bothrops</i>	2

^aEndemic genus fide Schwartz and Henderson.

(Forsgaard, 1983) a very peculiar anole; it is both montane and cryptic, and primarily confined to the south island of Hispaniola, the area south of the Cul de Sac-Valle de Neiba trough. All these characteristics are reasons that it was not recorded in the pre-1900s. A single specimen is known from above Constanza in the Cordillera Central of the Dominican Republic.

One genus and species of gecko is missing in the new listing: *Gymnodactylus fasciatus*. Kluge (1964) discussed the relevant problem. Duméril and Bibron (1836) described the species on a single specimen with the locality "Martinique" from the Plee collection. It is not a *Gymnodactylus*, as now understood, but instead a *Homonota*, a genus known only from between 15 and 48°S latitude in Argentina. In view of the fact that the genus has not been re-collected in Martinique and that the known distribution is so remote from that island, the locality is regarded as erroneous.

Seventy-one of the lizards are *Sphaerodactylus* and 18 are *Ameiva*. One is a surprising discovery of a Greater Antillean species of a gecko, *Phyllodactylus*, previously known in the West Indies only from far south in the Lesser Antilles. Only one species (three subspecies) is reported despite the fact that the animal is known from two of the Greater Antilles, from both Haiti and the Dominican Republic in Hispaniola, and also from the dry south central coast of Puerto Rico.

Thirteen West Indian anguids are *Celestus* and six are *Diploglossus*; one of the latter was described from a single specimen by Underwood from Montserrat in the middle of the Lesser Antillean chain. Two are in a genus (*Sauresia*) erected by Gray because instead of being pentadactyle like the previous two genera, these had only four toes on each foot. The other is a montane pentadactyle but earless genus (*Wetmorena*) named for Alexander Wetmore, who searched the mountains for birds. All species of *Celestus* and *Diploglossus* are distinguished on the basis of the structure of the osteoscutes (Strahm and Schwartz, 1977; but see Savage and Lips, 1993). *Sauresia* and *Wetmorena* resemble *Celestus* in "reduced and absent radices."

Twelve amphisbaenians are listed, two of them in the genus *Cadea*, which Gray had erected for *punctata* Bell. Stejneger (1916) casually mentioned the generic change in a note indicating that *punctata* was preoccupied and he substituted *blanoides* for it. [He mentioned another preoccupied name, a generic name for a South American taxon; his excuse for publishing the two names together was a curious one ("it is necessary to use the names in a forthcoming lecture")]. Mary Dickerson provided in the same year the second species in the identical genus (and also a better explanation for the recognition of the genus). All other amphisbaenians remain in the genus *Amphisbaena*.

There are 99 snakes recognized: 21 *Typhlops*, eight *Leptotyphlops*, nine *Epicrates*, and 13 *Tropidophis*. The genera of colubrids, apart from *Hypsirhynchus* (one species) and *Uromacer* (three species), have been changed or added to. Most of them have shifted generic names on the basis of Maglio's (1970) review of cranial osteology and hemipenial morphology. *Leimadophis* has totally disappeared. Sense can be made of the other names only on the basis of Maglio's discussion. *Antillophis* is the only new Maglio genus.

However, one new genus, the distinctive montane genus *Darlingtonia* was added earlier by Cochran (1935). It was named for P. J. Darlington, the second of the first two recorded herpetological explorers of the mountains of Hispaniola. (His herpetological efforts,

like those of Wetmore, were peripheral to his main interests. Darlington's major concern was with carabid beetles.)

In place of "*Chrysemys palustris*" there are four freshwater turtles of the genus *Trachemys*; this is not a great change since Stejneger (1904) confessed that Barbour relied on not enough specimens to allow him to estimate the number of species in the total fauna. There is still just one endemic crocodile in the West Indies.

No more than Barbour in 1914 did Schwartz and Henderson (1991) achieve a zoogeography of the herpetofauna of the West Indies. Discreetly, they did not attempt it. They have produced another overview, possibly nearly complete.

Periods in the History of Knowledge of the West Indian Herpetofauna

How did we get where we are? There has been a long history of observation and collecting in the West Indies. The history of knowledge of the herpetofauna of the West Indies can be divided into six periods. (I can only comment on a few workers that stand out in each period and perhaps exemplify the period in which they did their work.)

In these six periods there has been a very visible progression from essentially folk knowledge to scientific knowledge. Initially there was a great deal of carelessness about locality and a great ignorance of the features (characters) that distinguished the kinds (the species) that were recognized. In every period there has been a very evident solidification of standards and a truly steady increase in the individual application of clarity, consistency, and rigor in systematic procedure and example. To be sure, there have been in each period some people ahead of their time and others who were definitely behind their time; most were sometimes ahead and sometimes behind. There was obviously a learning process in which some learned part of the totality of what needed to be learned and some did not.

Schwartz was part of that learning process, and he was very good at what he did. He, like everyone, will be superseded. This is unavoidable and part of the progression of Science.

Period I

The first period was a pre-Linnaean period which I will characterize, using Hans Sloane as the example. (I confess that my choice of

Sloane is not solely on his virtues but also because he wrote about Jamaica, where I had my first experience of the West Indies.)

Hans Sloane (1660-1753) can be considered as the real founder of the British Museum. His collections provided the base for the British Museum (Günther, 1980; de Beer, 1953; Brooks, 1954). (The collections were both botanical and zoological, birds and mammals, but, not on verifiable evidence, herpetological.)

Long before his bequest, he spent 15 months (in another account 20 months) in Jamaica beginning in 1687. In 1725 he reported and figured the amphibians and reptiles that he had seen but apparently not collected. They are all recognizable (certainly to genus) from the figures, except for one lizard and a snake which is only recognizable by its English name, the Yellow Boa of Jamaica, and by its purported size. (The figure is definitely a caricature of a snake; crisscrossing lines presumably indicate scales. The artist must not have seen a Yellow Boa.) The lizards at least show genuine observation and sometimes careful observation.

Sloane was primarily a physician, employed while he was in Jamaica as the personal physician of the Governor-General of Jamaica. By avocation he was a botanist and not in any way a specialist in amphibians and reptiles, but even so his figures show a very good sample of the species that were then conspicuous elements of the herpetofauna, including some animals that may now be extinct. He dissected some of the specimens but succeeded in bringing none of them back to England. (That story is told below.) There were certainly no specimens available when John Edward Gray started the herpetological collections long after the museum was founded (Boulenger, 1906, the internal evidence of Gray's 1845 "Catalogue of the Lizards in the British Museum").

All of Sloane's (1725) figured sample's of the Jamaican herpetofauna have been identified, most of them correctly (Ahrenfeldt, 1954). I comment on Sloane's or Ahrenfeldt's observations and, incidentally, modernize the nomenclature. (In quoting Sloane, I have faithfully reproduced his spelling and punctuation, eliminating only the capitalizations which will distract the reader.)

"*Rana arborea maxima*" (p. 331) = *Osteopilus brunneus* Gosse, 1851

Sloane's remarks are these: "These tree frogs are very frequent in the woods of Jamaica. They are larger than ordinary frogs, slenderer, and of a light brown or grey colour." The figure is quite good for this

period, and there seems no doubt of its identity. Note the date of the name it now bears.

“The land tortoise” (p. 331)

“Tis common in the woods between Guanaboa and town every where.” Whether the land tortoise was endemic or imported cannot be determined without a specimen. If it was imported, whether it was *Geochelone denticulata* or *G. carbonaria*, the same observation applies. The population is now extinct. If there was ever an endemic land tortoise, its discovery and naming now belongs to paleontology.

“*Lacertus Indicus senembi & Iguana dictus*” (p. 333, Plate 273) = *Cyclura collei* Gray, 1845

“Gwanas are very fat and good meat.” This large endemic lizard was thought extinct until very recently. It has recently been rediscovered on the Hellshire Hills. It is still very much an endangered species.

“*Lacertus major e viridi cinereus, dorso crista breviori donato*” (p. 333, Plate 273) = *Anolis garmani* Stejneger, 1899

Sloane says, “This is found frequently in the woods of Jamaica, it differs little from the Guana, but being greener, less, and having a shorter crest or comb along the back. It lays eggs less than a pigeon’s egg.” The characteristic toe pads of *Anolis* are at least suggested, but the figured dorsal crest and described greenish coloration confirms the identity of the species. It is still common in the relict woods of Jamaica and sometimes occurs along roadsides (E. E. Williams, personal observation).

“*Lacertus major cinereus maculatus*” (p. 333, Plate 273) = *Ameiva dorsalis* Gray, 1838

Sloane calls it “The Greatest Spotted Lizard” and describes it in detail: “This is about 8 inches long, of the shape of other lizards, reddish under the chaps, light brown on the back and tail, with one yellowish green line along it, it is blue all under in the abdomen, & c.” Duméril and Bibron (1839) described it as *Ameiva sloanei* the year after Gray provided the name *Ameiva dorsalis* from a specimen “donated by R. Heward, Esq.” In his 1845 “Catalogue of the Specimens of Lizards in the Collection of the British Museum” Gray recognized the reality of Sloane’s priority by placing the English name “Sloane’s *Ameiva*” alongside his own formal Latin name.

“*Lacertus cinereus minor*” (p. 333, Plate 273) = ??

Sloane calls this “the least light brown or grey lizard” and describes it as “two inches and a half long, of which the body is in measure but an inch, and not much bigger than a goose-quill, it has four legs, the hindermost pair is the longest, it is all over smooth, of a light brown or gray color, except the tail, which has brown marks on it, the belly and under side of the body is whitish and the tail very small. It loves moist places and stones, where it is to be found leaping from one to the other, not only here but through the Caribes.” This was identified by Ahrenfeldt, perhaps on the advice of Malcolm Smith, whom he consulted, as *Anolis opalinus*. I doubt this identification. The animal is the correct size, if it were a young specimen, although I hesitate because the phrase “not much bigger than a goose-quill” does not accord with the build of the animal (nor the figure!). It lacks the light line along the sides which enables the identification of *A. opalinus* in life and, more important, lacks the suggestion of adhesive pads seen in the figure of *A. garmani*. The behavior as observed is also anomalous. *Anolis opalinus* is arboreal on trees or bushes. It is not terrestrial, not fond of stones or moist places, and not “leaping from one to the other.” It is endemic, not found “not only here, but through the Caribes.” Indeed, on the basis of behavior, I know of nothing alive today that fits this description. (Perhaps the figure and the text confuse two animals, but I can make nothing of Sloane’s text.)

“*Lacertus minor laevis*” (333, Plate 273) = *Mabuya* sp.

Sloane cites this as “bigger than the former, having a great many brown spots, otherwise much the same, laying a very small white hard shel’d egg...and nesting in rotten holed trees, leaping from one bough to another. Tis very common among old Palisadoes, & c.” This is clearly a West Indian *Mabuya*, so much so that it has been the source of confusion: In describing *Scincus sloanei* Daudin (1802a) not only intended to honor Sloane but also quoted the latter’s short Latin description and praised his figure. Stejneger (1904), however, was emphatic that *S. sloanei* could only have been based on the same specimen in the Paris Museum that Duméril and Bibron, as they themselves asserted, described as *Eumeces sloanei*. Daudin apparently recognized this specimen as being very similar to Sloane’s “*Lacertus minor laevis*.” This specimen, twice described under the trivial name “*sloanei*,” unfortunately, was from St. Thomas and, by general agreement, the taxa of the two islands are at least subspecifically different. Fortunately, Schwartz and Henderson (1991) decided that the same subspecies name is applicable from the Puerto Rico Bank all the

way to Jamaica. *Mabuya mabouia sloanei* (Daudin, 1802a), in consequence, cannot be the animal from Jamaica.

“*Salamandra minima, fusca, maculis albis notata*” (p. 334, Plate 273) = *Sphaerodactylus argus*; “The Wood Slave”

“This is about an inch long from the end of the snout, to that of the tail. It has two small eyes, four legs, each of which three quarters of an inch long, with two joints of and five toes, and a thick, blunt tail, of reddish brown colour. The body is not near so thick as one’s little finger, ‘tis all above of a brown colour, with white little spots like stars, and a white in the belly, and is all over cover’d with scarce discernible scales. The figures are different, one being taken from the lizard when dry’d, and the other from one preserv’d in spirit of wine.” The toes in the two figures do not show the characteristic toe structure of the genus, but, given that the animal was so small, the artist might have missed it, as he did not in the much larger *Anolis garmani* and as he should not have in the larger “*Lacertus cinereus minor*” – if it is an *Anolis*.

“*Scincus maximus fuscus*” (pp. 334-335, Plate 273) = *Celestus ociduus* Shaw, 1802

Sloane called this “a Galliwasp,” the first usage in English of this name for the genus now called *Celestus*. He described it as follows: “it was 11 inches long from head to tail, and six inches about the middle, where biggest, it was almost round every where from a sharp round snout, increasing to the middle, and from thence decreasing to the tail, which ended in a point; the back was hard and a little compress’d, and so was the belly, it had two round spiracula or nostrils in the two corners of the snout. About half an inch behind them were the eyes, and near an inch further two oblong holes for the ears; about three inches from the beginning of the snout towards the tail were the fore feet, which had two joints, and were not over an inch and a half long, with five toes like those of a lizard, the hinder legs were of the same length, & c. and beyond them the anus, covered with a transverse flap, the back or upper parts were all covered over with rhomboidal small rows of scales of a brown colour, with spots of orange color, and belly with the latter only...It is found in marsh grounds, in several parts of the island.”

Sloane described three snakes, but only figured one. Given the absurdity of the figure, the artist must not have seen any snake at all.

“*Serpens major subflavus*” (pp. 336-337, Plate 274) = *Epicrates subflavus* Stejneger, 1901

Sloane called this “the Yellow Snake” and described it as follows: “This is in length about seven or eight feet, its head is not very large, having nostrils and phangs tho’ not long, the neck is small, being near two inches about, rather less then his body, which growes bigger, till it be about as big as ones wrist and continues so large to the anus, whence it diminishes by degrees to the tail; its head is of a dark brown colour, and the scales all over the body are of a dark brown, with some yellow streaks here and there, the belly is all yellow, and cover’d with larger and greater scales than the back....It is for the most part in the woody mountains, quailed up in the paths as ropes in a ship....They feed on birds, rats, & which they swallow whole, and therefore Nature has given them such a folded or rugous tunicle of the stomach, that it may eat things of large dimensions. Many of them have been kill’d with thirteen or fourteen rats in their bellies.”

“*Serpens major nigricans*” (p. 337) = *Alsophis ater* Gosse, 1851

“The black snake is only smaller, else in everything the same, although not venomous.” There are two black snakes in Jamaica, one large, *Alsophis ater* Gosse, one small *Arrhyton funereum* Cope. Sloane’s adjective “major” decides the issue. The species *ater* is probably extinct.

“*Serpens major cinereus*” (p. 337)

“There is a sort likewise of a light grey colour.” Color is reputed to be variable in *A. ater*; this is surely a color phase of the latter.

Sloane did, in fact, attempt to bring some these “uncommon Creatures” home to England. He intended to bring the Yellow Snake (“seven feet long”), the Guana, and also a crocodile back from Jamaica alive. There was a tragic conclusion to his effort. While all the animals survived for a while on the ship peaceably and successfully, the snake was shot by servants when it invaded their quarters, the Guana was frightened by a seaman and fell into the sea, and the crocodile died of unknown causes (de Beer, 1953, pp. 47-48). Sloane did not preserve any of these animals nor any of the small ones. The only record of all of these animals are the figures and the text appended.

If at least the snake had arrived alive in England, if it had been preserved or even skinned, and if it had been part of collections that Sloane gave to the British Museum, the Yellow Snake of Jamaica would not have had to wait for Stejneger (1901) to give it its formal

Latin name, *Epicrates subflavus*. Stejneger conspicuously did not mention Sloane's note on the Yellow Boa, although he usurped the very epithet that Sloane used.

Two of these figures were sufficiently appreciated that they were recognized and named from the figures. *Scincus* (= *Mabuya*) *sloanei* Daudin 1802 and *Ameiva sloanei* Duméril and Bibron were in obvious honor of the man who discovered them and had them drawn.

Two other species were adequately figured: the Guana and the Crested Anole. The formal naming of these had to wait. The Guana was named by Gray (1845) *Cyclura collei*, not from Sloane's figure but from a stuffed specimen "Presented by the Admiralty, from Haslar Hospital." (It is called "Colley's Cyclura" at the beginning of the description; who Colley was is not explained.)

"The Great Crested Anole of Jamaica," Stejneger's English name for this animal in the title page of his formal description, was not named until the end of the nineteenth century (1899; Stejneger did mention Sloane's figure) and was rescued from a misnomer under which it suffered for many long years. Stejneger (1899) expressly says: "Herpetological writers have shown a curious unanimity in misnaming the large crested *Anolis* of Jamaica *Anolis edwardsii* Merrem." He was able to demolish the relevance of this name for the Jamaican species very effectively and named the species after West Indian specialist Samuel Garman whom one of the same "herpetological writers" had done an injustice.

Period II

The next period begins with the *Systema Naturae* of Linnaeus (1758) and ends with the first volume of the "Erpétologie générale" by Duméril and Bibron (1834).

Not many people have examined even the reprint of the 10th edition of the *Systema Naturae*, especially for items from the West Indies. All cold-blooded tetrapods were called "Amphibia." Under "Reptilia" there were just four genera: *Testudo*, *Draco*, *Lacerta*, and *Rana*. "Serpentes" included *Crotalus*, *Boa*, *Coluber*, *Anguis*, *Amphisbaena*, and *Caecilia*. The single West Indian representative that Linnaeus knew is surprisingly the Bahaman *Typhlops lumbricalis*, recorded under the genus *Anguis*.

The entry is characteristic of this early time:

lumbricalis- 230-7. Gron. mus. 2. p. 52 n. 3

Brown. jam. 460. t. 44. f. 1
Seb. mus. 1. p. 137, t. 86. f. 2
Habitat in America.
Color ex albido flavescens.

The numbers to the right of the name itself are respectively counts of ventrals and subcaudals, testifying to the use of the counts for snakes at so early a period.

This and the preceding period were epochs of random discovery, collection, and description. Many aspects contributed to this phase. Frequently the collector and the describer were quite different individuals. Often the exact locality was never given or, if given, was soon lost. Often the collector and the describer never saw each other. Many specimens had long been in the Cabinets of Curiosities. Many figures or descriptions in travel books that were often much worse than Hans Sloane's were interpreted, regardless of geographical plausibility, as identical with the usually badly preserved specimens that were in hand. Indeed, the localities of many specimens were "unknown," wildly wrong, or extraordinarily imprecise as "America" in Linnaeus.

Laurenti was the first to describe a West Indian species after Linnaeus. It was *Iguana delicatissima*, the second and more obscure of the two species of *Iguana* and the only one endemic to the Lesser Antilles. (Laurenti adopted for the generic name the second part of Linnaeus' name, *Lacerta iguana*, and because he avoided use of tautonomous names, he called the Linnaean iguana, *I. tuberculata*.) In the matter of locality, both species were labeled, typically for the period, "Indiis" (the Indies).

It is clear from the many *Iguana* species that Laurenti cites, many of which are superficially similar to agamids, that he did not possess the restricted modern concept of the genus *Iguana*. Not until the next period did Gray (1845) introduce the modern concept of just two species.

Anders Sparrman described, unwittingly, the first West Indian *Anolis*, *Lacerta bimaculatus*, 1784 and the first West Indian *Sphaerodactylus*, *Lacerta sputator*, 1784. His Plate IV portrays both unmistakably.

The Paris Museum began to be active about this time; it was called at this time the Cabinet du Roi. Only in 1793 did the assembly pass a law that transformed the Cabinet du Roi into the Museum National d' Histoire Naturelle. Previously, the Comte de Lacépède, among other duties, was Keeper of the Cabinet du Roi and published two volumes in

1788 and 1789 (i.e., just before and in the first year of the French Revolution) that described four West Indian species, one from Patrick Browne's (1789) book on Jamaica and three from "Martinique," on the basis of specimens in the Cabinet du Roi. The one from Browne's book was a freshwater turtle from Jamaica now called *Trachemys terrapen*. The other three species are as follows: the anole *Anolis roquet*, the colubrid *Liophis cursor*, and the crotalid *Bothrops lanceolatus*. (I put "Martinique" in quotation marks because the Paris collection, despite the fact that three of these species coincidentally came from Martinique, has a poor record of accuracy in claiming species that actually came from "Martinique." I previously noted that "*Gymnodactylus fasciatus*, which is *Homonota*, was inferred to be from far south in South America.)

It is a curious fact that the holotype of *L. cursor* may be in the collection of the Academy of Natural Sciences in Philadelphia. Lacépède mentions a specific specimen that he describes in detail, including color pattern, and in another part of the book he includes a table giving ventral and subcaudal counts. There is no explicit mention of syntypes. There is therefore no credible reason to doubt the statement of Dixon (1981, p. 300) that ANSP 5580 is the holotype of Lacépède's *cursor*. (Cope described this exact specimen as his new species *Liophis putnamii* but obscurely admits that the specimen is from Paris "Habitat. Martinique. Mus. Acad. Phila.; Mus. Plants in ex." I presume "in ex" means "in exchange.")

Even 30 years after the 10th edition of the *Systema Naturae*, Lacépède did not use the binomial consistently. Even in his new West Indian species, *terrapen* or *roquet*, and *cursor*, he used just the single name. Only for his new *lanceolatus* does he use *C.* for *Coluber*, the Linnaean genus in which he categorized this very venomous snake. Even among the snakes, in which he is much more consistent, for *cursor* he uses the single name, latinized and italicized, but without the prefatory *C.* Because of this inconsistency Lacépède's names have been rejected as not binomial by the International Commission, and those names not expressly conserved are at least temporarily invalid, including all three of those discussed previously.

Daudin (1802a), in contrast, published during the French Revolution. His eight volume *Histoire Naturelle, Generale et Particuliere, des Reptiles* bears the odd date: an X for Year 10 of the revolution. He is the inventor of the genus *Anolis*, a native name which has survived many years but only recently has received its generotype (by fiat of the International Commission). He included both *Lacerta bimaculatus* Sparman and *Lacerta sputator* in *Anolis*. He described only one West

Indian reptile species: *Ameiva erythrocephala*. He described it from a very detailed letter sent by M. Badier to Lacépède. The letter included measurements and diet as well as colors in life. The species is mentioned more briefly just as "Tete Rouge," without the Latin name, in the Additions a l'Histoire Naturelle des Quadrepedes Ovipares beginning on p. 487 of the second volume of Histoire Naturelle des Serpents of Lacépède. The requirement for an actual specimen in the hands of the describer was not understood at this time, certainly not by Daudin, nor, apparently, in the case of "Terrapen", by Lacépède.

As mentioned previously, at long last the galliwasp of Sloane was described as *Lacerta occidua* by George Shaw in his 1802 General Zoology, the first book in English to use the Linnaean system.

That Sloane's was the animal named *L. occidua*, which translates to the Western *Lacerta*, by Shaw is certain. He does not refer explicitly to Sloane but rather to Browne. However, he uses the term galliwasp, a local name in Jamaica which was first brought by Sloane into more general currency. It is also certain that *C. occiduus* was misnamed. Shaw was under the impression that the lizard he called the Australasian Galliwasp, his *Lacerta scincoides*, actually a true giant skink, was just a variety of the anguid *L. occidua*. This is an example of both the extreme confusion of species that was prevalent during this period and especially the total lack of realization that the range of a species might be restricted to a single island. I shall discuss two other examples of this taxonomic and zoogeographic blindness that especially apply to Jamaica.

Shaw also described as *Anguis jamaicensis* the *Typhlops* Patrick Browne (1805) cited as "The Silver Snake." Again, as with Shaw's name *L. occidua*, the name is based entirely on Browne's description; there is no hint of a type.

Neither the *Cyclura* nor the *Anolis* which Hans Sloane had figured, were described during this period. This was partly because there was great and very natural confusion as to the real species, reflected in the synonymies which plague every description or discussion of any species. The characters were not yet known, inevitably, in this early period to distinguish the authentic species.

Nowhere is this more evident than in the West Indian names which consensus attributes to Blasius Merrem (1820): *Anolis equestris* and *Anolis cuvieri*. The characters that are supposed to distinguish the species cause any anole expert to stop and stare: they are definitely not the ones anyone would choose today. The first trivial name identifies it as the Knight anole, which presumably distinguishes the species as having a helmeted head, i.e., a casque. None of the other

presumed distinguishing characters listed on facing pages in Latin and in German could be understood from the text itself. There is, however, a footnote to Cuvier's "Le grand *Anolis* à Echarpe" in his *La Regne Animale* and to a figure that is distinctly more useful. Cuvier's epithet (echarpe = sling in French) translates to the equivalent of "bandirt" (= "in a sling") in German on the left-facing page of Merrem's book. A light streak above each shoulder does look vaguely like a sling.

The name *cuvieri* conveys no information except the intention to honor Cuvier. However, there is a footnote reference to Cuvier's "Le Grand *Anolis* à Crete" and a figure that shows a high tail crest. There is again no indication of types other than the figures, which are excellent and which explain the characters of the tail by which Merrem (1820) chose to define his species.

The phenomenon of providing a formal Latin name is the reason that the genotype of *Anolis* is *Anolis carolinensis* Voight, 1832. In all the several French editions, Cuvier in his *Regne Animale* used only the French "l'*Anolis* de la Caroline." Friedrich Siegmund Voight was responsible for a German translation as "Das Thierreich." In the course of a mostly literal translation, Voight transformed the French name used by Cuvier into a formal Latin name!

Only two species have been ascribed to Cuvier, the Cuban endemic crocodile, *Crocodylus rhombifer* 1807, three syntypes initially of unknown locality, and the first West Indian amphisbaenid, named *Amphisbaena caeca* Cuvier, 1829 and distinguished by being "totally blind" and erroneously originally attributed to Martinique. Later, *A. caeca* was determined to have come from Puerto Rico and to have eyes after all. The latter is named in barely a phrase, but it did have a type; the *rhombifer* syntypes are at the very least misplaced, probably lost.

The paper in which Cuvier described *C. rhombifer* provides a thorough discussion of the species of crocodiles then known, with figures of the skulls and also the patterns of the dorsal scutes that are still used as taxonomic characters. It was certainly extraordinarily valuable in its time. It is the "time" I complain about, and it is the reason I am making disrespectful comments about Cuvier and Merrem. The neglect of all the data that we now know to be indispensable was not only permissible but all but inevitable. Both were able men who entered herpetology and systematics when these "disciplines" were in their naive very first stage. It was a time when providing a recognizable Latin name, whether from a picture without a physical

type or a type without a picture, or merely a unambiguous type locality was itself a service.

Richard Harlan named the first *Cyclura*, *Cyclura carinata* Harlan, 1824, a southern Bahaman lizard. He also provided a long overlooked name, *Hyla crucialis* Harlan, 1829 for the very distinctive Jamaican species for which, long after, Trueb and Tyler (1974) erected the generic name *Calyptahyla* not on externals at all but by utilizing skull characters. (Actually, the generic name was proposed for *Hyla lichenata* Gosse, 1851, the name long used for the earlier *H. crucialis*. There was, in fact, a rather heated dispute between the advocates of the respective names.)

The period between the 10th edition of the *Systema Naturae* and the *Erpétologie générale* was unpredictable and, indeed, random regarding the description of species from the West Indies. Species large and small are commingled, as are species that would be expected to be cryptic or fossorial and those that would be expected to be conspicuous. Confusion regarding to identity of species is the most evident feature of this period. Local naturalists are the tale-tellers and museum men, if they are even that, provide formal names – binomial if they were to be accepted by future generations.

Period III

The third period is the entire time span (20 years) of the *Erpétologie Générale* by Constant Duméril and Gabriel Bibron from 1834 to 1854. (After Bibron's death in 1848 the title pages of the later volumes had three names: The third was Auguste Duméril, the son of Constant.)

This is the very first book, or set of books, which is a complete work with full synonymies, elaborate descriptions (not in Latin), and even dichotomous keys. (I discuss two – one to genera and one to species – by way of illustration; each includes at least one West Indian taxon.) The authors tried to include summaries of aspects of the taxa they treated. Not only the systematics but also what was then known (although it was not as much information as exists today) of the anatomy and physiology and even the habits and behavior of the animals were described. It was an immense step forward and it was a considerable advance over the previous *Synopsis Methodica* or *Table Methodique* of Lacépède or the pages of listings provided by Merrem (1820), who merely followed Linnaeus in this respect. In some respects it has not been superseded. No subsequent set of books has even attempted to summarize all the literature. This is no longer possible.

During this period, there was general agreement that it was no longer permissible to describe a species without a specimen in hand. Museums and their collections became more important. It is not especially relevant to the West Indies, but it is significant for the progress of herpetology that Duméril and Bibron (1834, volume I, p. xi) included a tabulation of the total species known from the time of Lacépède to 1834. I reproduce it here in translation:

	Lacépède in 1790	Daudin in 1805	Merrem in 1820	In the Museum collection in 1834
Turtles	24	62	62	97
Lizards	56	88	83	168
Snakes	172	315	348	391
Amphibians	40	91	87	190

Thus, the total number of species in the Paris Museum in 1834 was 846. At the end of the nine volumes in 1854, Schmidt (1955, p. 601) estimated that 1393 species were treated.

The British Museum started its herpetological collections during this period. John Edward Gray was their first curator.

Regarding Jamaica, in 1845 John Edward Gray began cataloging the lizards of the collections of the British Museum – sans any from Hans Sloane. He included *Cyclura collei*, *Ameiva dorsalis*, *Anolis grahami*, and *A. lineatopus*, all named by himself. He did acknowledge Hans Sloane's priority by printing the English name "Sloane's *Ameiva*" in front of his own formal Latin *A. dorsalis* in the 1845 catalog. He also used Daudin's *M. sloanei* but attributed it to Jamaica.

Gray's 1845 catalog is the first of a series in which a list of the herpetological specimens actually in the collection of the British Museum was intended to be authentically a checklist of the herpetofauna of the whole world. It lists the desiderata of the British Museum (Natural History) by italicizing the names of the species wanted.

Gray was completely a museum man (as were all his successor curators at the British Museum until Grandison and Arnold, recent curators).

In 1851 Philip Henry Gosse, not a museum man, wrote an extraordinarily pleasant book: *A Naturalist's Soujourn in Jamaica*. The first paragraph of its preface deserves to be quoted often:

Natural History is far too much a science of dead things; a necrology. It is mainly conversant with dry skins furred or feathered, blackened, shrivelled, and hay-stuffed; with objects, some admirably beautiful, some hideously ugly, impaled on pins, and arranged

in rows in cork drawers; with uncouth forms, disgusting to sight and smell, bleached and shrunken, suspended by threads and immersed in spirit (in defiance of the aphorism, that "he is born to be hanged will never be drowned") in glass bottles. These distorted things are described; their scales, plates, feathers counted; their forms copied, all shriveled and stiffened as they are; their colours, changed and modified by death or partial decay, carefully set down; their limbs, members, and organs measured, and the results recorded in thousandths of an inch; two names are given to every one; the whole is enveloped in a mystic cloud of Graeco-Latino-English phraseology (often barbaric enough); –and this is Natural History!

Gosse partly retracts this vivid diatribe immediately on the second succeeding page:

The author would not be misunderstood. He is far from despising the labours of those who describe and catalogue the specimens that travellers send to the cabinets of Europe. Careful and minute descriptions, accurate admeasurements, and distinctive names are absolutely indispensable to science; but they must not be confounded with science itself. Valuable as these details are,...they are the cumbrous machinery by which that knowledge is preserved and communicated to the world.

Gosse could not afford wholly to denigrate the "necrology" of biological collections. I have myself seen his syntypes of *Anolis iodurus* and *Placopsis ocellata* and the holotype of *Draconura catenata* and participated in the discovery that the alleged type of *A. opalinus* was not the true type but was *A. grahami* (Underwood and Williams, 1959). [Apparently, if any type of *A. opalinus* was ever sent to the British Museum, the confusion occurred early; possibly it was even Gosse's mistake. Cope, who in the next period visited the British Museum (1863b) and examined the so-called type of *A. opalinus*, described much later (1895) *Anolis flabellatus*, a synonym of that species. Boulenger in his Catalogue of the Specimens of Lizards in the Collection of the British Museum (1885, vol. 2) considered *A. opalinus* a synonym of *A. grahami* Cope and redescribed *opalinus* years later as *A. flabellatus* Cope, 1895. Everyone who has examined authentic or live *A. opalinus* has agreed that the species is valid. This "type" has only caused confusion.]

Gosse (1850) named *Sphaerodactylus argus* (thus ending the formal anonymity of Sloane's "*Salamandra minima*") and another *Sphaerodactylus*, *S. oxyrhinus*. Both are still valid today. His *A. iodurus* was a synonym of *A. grahami*. His *A. maculatus*, his Zebra Lizard, cited, "especially common around Kingston", (p. 491) is the equivalent of *A. lineatopus*. (Of the two synonymous Gray names Boulenger chose the latter.) His *Placops ocellata* is *A. valencienni* Duméril and Bibron, and his *D. catenata* is *A. sagrei* of the same authors. He is certainly not to be praised for his care in his anoline taxonomy, but he is

to be praised for his careful observations. His *A. opalinus* and two *Sphaerodactylus*, and also his one snake *Alsophis ater* and two frogs (*Osteopilus brunneus* and *Eleutherodactylus luteolus*) of his ventures into herpetological taxonomy alone bear his names. The *A. opalinus* survives, despite the absence of a verified type, because he was able to recognizably characterize the species in his description.

Gosse (1850) did not name either the giant anole of Jamaica, which he cites informally as the Venus lizard, insisting that this is (like, in fact, *Anolis*) an Indian name "used by the negroes", (p. 142) or the Yellow Boa. He called them by previously described names *A. edwardsii* Merrem, 1820 and *Chilabothrus inornatus* Reinhardt, 1843. Both are incorrect attributions to Jamaica of animals named from other islands, the anole from Nevis in the Lesser Antilles and the snake from Puerto Rico. In the case of the Yellow Boa, Gosse felt compelled on the inappropriateness of the species name:

The trivial name *inornatus*, which MM Duméril and Bibron have selected to designate the species, must be considered as comparative, for this boa, when seen alive, in its black and yellow livery, I think, is far from unadorned, the contrast of colours being fine, and the purple iridescent glow that is reflected in the playing light from the dark parts of its polished armor is very rich and brilliant.

Stejneger (late in the next period, 1899 and 1901) had to correct these errors.

Period IV

The fourth period is the latter half of the nineteenth century and the very beginning of the twentieth century and its primary exemplar was Edward Drinker Cope. There continued to be a tendency to separation between the "expert" in the museum and the collector in the field. Cope, while an enthusiastic collector regarding the United States and certain parts of Mexico (Orizaba, Lake Xochimilco, and Hidalgo; Osborn, 1931:489) is the absolute extreme of the museum describer regarding the West Indies. He never saw any West Indian island, but his names for West Indies species are almost competition for Schwartz. They far exceed all other authors except Schwartz. He was brilliant and so obviously so that he impressed every one he met. He soon had the collections of the Philadelphia Academy and the Smithsonian at his disposal and readily borrowed specimens from Louis Agassiz at the Museum of Comparative Zoology. When he visited the British Museum in 1863, he was given free access to the collections, and so it was in the European museums, which he visited the next

year. (When he described the British material in August 1864, he felt compelled to thank them for their courtesies: "Of the preceding species of *Anolis*, sixteen have been derived from the British Museum collections. My particular acknowledgments are due to Drs. Gray and Günther, the directors, for the ample facilities afforded me in the examination of these and other objects of interest under their care.")

Most of his herpetological collections are from his early years; actually, most of his new West Indian species are from the first years before he was preoccupied with paleontology. In the earliest years most of his specimens came from Charles Wright from Monte Verde in Cuba (these were deposited in the Philadelphia Academy or the Smithsonian) or from D. F. Weinland from near Jeremie in Haiti (these specimens were the property of the Museum of Comparative Zoology). He had to make his descriptions by external characters that were modified by preservation. His comment on Gosse's descriptions is characteristic of all workers who have not had the opportunity to observe the color of the animals in life: "they scarcely assist the student in separating the species when rendered unnatural by preservation in alcohol." Despite difficulties of this sort, Osborn (1931) quotes Theodore Gill (who knew Cope at the National Museum) as saying; "He found herpetology an art; he left it a science; he found it a device for the naming of specimens, he left it the expression of the coordination of all structural features." This is hyperbole, but there is also a measure of truth. It was in the broader aspects of herpetology that Cope excelled, in the delimitation of larger groups, and in the use of structures that had proved valuable for these larger groups, for example, the pectoral girdle of frogs and the lungs and hemipenes of snakes.

Albert Günther, a German (Gray's successor at the British Museum), described one Hispaniolan taxon as *Hypsirhynchus ferox* (explaining the trivial name of this rather small and inoffensive snake by the observation: "The ferocious physiognomy of the snake, produced by the peculiar form of the snout, in combination with the strong dentition, has caused the denomination.") He also provided the prior trivial name, *dorsalis*, of a taxon (also Hispaniolan) that Cope recognized as the genus *Ialtris*. These deserve mention as the only two generic names of West Indian colubrid snakes from this period that have survived unchanged since the generic revisions of Maglio (1970). However, there is general consensus that Günther's greatest contribution was his choice of a successor, a Belgian named Boulenger.

George Albert Boulenger, who in some sense popularized Cope's ideas by adopting most of them in his "Catalogues of the British Mu-

seum" (1882-1896) that are still useful today. They have survived not only because he lists, following the Gray and Günther precedents, all the species that the British Museum did not have, and thus provided a checklist of the world herpetofauna, but also because the useful characters for each group began to be known. Also, he used a standard format for each group that he described. (This was an element of description that Cope did not achieve consistently.) Boulenger's volumes are the superior successor reference volumes that definitively replace the *Erpétologie Générale*. The Catalogues, in contrast to Duméril and Bibron, do not discuss previous classifications. They do not report distributions except in the briefest of terms; they also do not give any natural history notes. Again they are unlikely to be duplicated, not by any single author or pair or trio of authors.

Boulenger never went into the field. He too never saw an Antillean island; in contradiction to Cope, he is not reported collecting salamanders or snakes. (When he retired, he did research on roses.) During all of his active career, he sat in London, very usefully describing the herpetological spoils of the British Empire. However, Boulenger's contributions to the description of the West Indian herpetofauna are meager, only *Chironius vincenti*, *Sphaerodactylus vincenti* and *Anolis watsi* were added, all species of the Lesser Antilles.

Samuel Garman was also a collector who was a describer. He had an extremely varied life before he became a museum man, associated with Louis and Alexander Agassiz and eventually the Curator of both Ichthyology and Herpetology at the Museum of Comparative Zoology. He took the opportunity of being Alexander Agassiz's assistant on the ship *Blake* to make the splendid collections that enriched our knowledge of the Lesser Antilles and other Caribbean islands as well.

It was these collections that were the occasion of serious disagreement between one museum expert and one man in the field. Boulenger, who was the Recorder for the Zoological Record in 1887, made the following comment in an entry on Garman's new names for anoles in the West Indies, primarily the Lesser Antilles:

Anolis, Daud. S. GARMAN, Bull. Essex Inst. xix, has greatly multiplied the binomials of the West Indian forms of this genus. The Recorder would refer the new species and varieties to the following more comprehensive terms: -- 1, *Anolis cristatellus*, D & B: *A. scriptus*, n. sp., p. 28, Silver and Lena Keys, Florida. 2, *A. gingivinus*, Cope: *A. virgatus*, n. sp., p. 41, St. Bart's. 3, *A. leachii*, D & B: *A. asper*, n. sp., p. 31, Marie Galante, *nubilus*, n. sp., p. 32, Redonda, *sabanus*, n. sp., p. 39, *speciosus*, n. sp., p. 42, Marie Galante, *lividus*, n. sp., Montserrat. 4, *A. alligator* D & B: *A. gentilis*, n. var., p. 34, Petit Martinique, *cinereus*, n. var., p. 35, Grenada, *extremus*, n. var., p. 35, Barbados, *luciae*, n. var., p. 44, St. Lucia, *vicentii*, n. var., p. 46, St. Vincent. 5, *A. richardii*, D & B: *griseus*, n. sp., St. Vincent, *A. trossulus*, n. sp., Grenada. Also copious notes on other known spe-

cies. *A. conspersus*, n. sp., P. Am. Phil. Soc., xxiv, p. 273 Grand Cayman, W. Indies [= *A. grahamii*], Gray. -- REC.

Boulenger was able to evaluate these species from preserved specimens (Boulenger, 1906). Garman had sent him examples of his recently described species.

Barbour (1914) and Lazell (1972) revalidated most of these species, and Lazell's is the revision of the Lesser Antillean anoles accepted today. The non-Lesser Antillean Garman species are also accepted as full species. Boulenger was, in my judgment, the best of the museum men, but he had not been able to see the animals alive. There is, in fact, no substitute for experience in the field, but there is also no substitute for a museum's opportunity for careful comparison. Boulenger did accurately identify the species groups to which all these species belong.

Stejneger, long Curator of Herpetology at the United States National Museum, by no means made as great a global herpetological contribution as Boulenger; he did not attempt any herpetological monographs or catalogs. (His original field was ornithology; he only took the job of Curator of Herpetology because it was available. The latter part of his long life was spent in administration.) He did have a legal background, having graduated in law from the University of Christiania in his native Norway, and so he was well prepared to be a member of the International Commission on Zoological Nomenclature in 1898.

He was not, however, merely legalistic; his strong point was accuracy. He was, therefore, bitterly opposed to the (in his view) casual behavior of Cope and Boulenger in employing series of syntypes instead of holotypes. He also instituted, for the same reason, possible confusion of two more species, the detailed description of a holotype, instead of a series.

It was undoubtedly as part of his search for accuracy that he provided names for the Great Crested Anole of Jamaica, his own English name (*Anolis garmani*, 1899), and for the Yellow Boa (*Epicrates subflavus*, 1901) of that island.

It is clear that there was never any justification for the use of *A. edwardsii* for the giant anole of Jamaica. Stejneger was able to demolish the use of this name. First, it was mentioned as "brought from the Nevis in the West Indies, by a young gentleman who came to London for education"; second, while the figure was life size, it is not nearly large enough to be the giant anole of Jamaica; third, it has no trace of

the saw tooth dorsal crest that distinguishes it from all other anoles on the island.

Epicrates subflavus presented more difficulty for Stejneger (1901). There is a difference in color (so vividly described by Gosse), unmentioned by Stejneger except in the (borrowed?) epithet, *subflavus*. Stejneger could find only one constant scale difference between *inornatus* and *subflavus*, the contact in *subflavus* of the prefrontals with the preocular rather than the separation by small scales.

Period V

The fifth period begins with Stejneger's "Herpetology of Porto Rico" (1904) and is the period typified by "island surveys." It is the first period in which there was a solid effort to collect the total herpetofauna of the individual large islands of the Antilles. Museum people frequently also collected. The museum man had "the big picture"; he was therefore anxious to fill in the gaps, geographic and taxonomic. He did so by collecting himself, fusing the roles of museum man and the field man.

The Greater Antilles were all surveyed. Puerto Rico was surveyed twice, by Schmidt (1928) and Stejneger (1904). Barbour and Ramsden (1919) did Cuba, Cochran (1941) did Hispaniola, and Lynn and Grant (1940) did Jamaica. The Lesser Antilles never had a genuine summary for all groups. Perhaps this is because the diversity is so special.

Of the successive surveys, Stejneger's is my favorite. He describes individual specimens in hand and when possible color in life. Stejneger (1904), on the first page of "The Herpetology of Porto Rico," he confesses that his own collecting experience was for a relatively frivolous reason:

The trip that I undertook in conjunction with Dr. C. W. Richmond, Assistant Curator of Birds, was primarily for the purpose of collecting specimens characteristic of the West Indian fauna for exhibition at the Pan-American Exposition in Buffalo. It was originally our intention to proceed from Porto Rico to the smaller on the east and south, but an attack of fever to which we both fell victims prevented us from carrying out our plans. The fever and the long convalescence also seriously interfered with our work in Porto Rico, so we lost nearly three weeks' collecting.

Barbour and Ramsden's (1919) Herpetology of Cuba is impeccable except that, as Schwartz and Cuban workers now active on the island have been able to show, it is woefully inadequate.

Schmidt's production of 1926 is mostly a copy of Stejneger, to the extent of quoting verbatim Stejneger's notes on color in life and

repeating his figures, but Schmidt does include "original descriptions." He cites even Cuvier's description of *Amphisbaena caeca*, which he describes as "evidently quite useless." He then presents, not Duméril and Bibron's redescription of the typical material, but Stejneger's account, which he declares "much the most useful."

Cochran's *The Herpetology of Hispaniola* (1941) is, in some ways, worse. She also fills up space with original descriptions, but she describes the hues and tints provided by specimens in alcohol at inordinate length. Her figures are acceptable, but her plates (those which show patterns a little obscurely) are photos of very obviously dead specimens that add no information at all.

However, she and Lynn and Grant (1940) deserve the faint praise of "useful." I have used them and have usually not been misled.

I am sure that Thomas Barbour was the most conspicuous West Indian herpetologist of the period. I use conspicuous in its invidious sense. I quote entire the paper by Anthony Curtis (1947:224) titled, "Prevalence of Snakes in Haiti":

Some authors have stated that snakes are practically unknown in Haiti. The late Dr. Barbour states that *Uromacer catesbyi* is widespread but rare. It is really very common, even in the gardens of this town, as many specimens to Dr. Cochran attest. Dr. Barbour stated that *Epicrates striatus* seems to be really uncommon. Every large dead tree which we have opened in the low lands, even in local gardens, contained one or more of these snakes. Many unopened trees are from time to time festooned with their shed skins. We find large boas stretched on branches after rainy nights or in our houses pursuing rats. I have found no boas in the mountains.

In some places the boa is respected as being the emblem of one of their jinn; hence Dr. Barbour may not have seen it. He used to send out blacks he met along the shore, retire to his boat, and buy what they brought in late that day. Many blacks here keep snakes in captivity, but seldom show them to strangers.

Barbour was a special mixture of the professional and the dilettante. He was indeed both a collector and a describer. Perhaps his early work, before the arrival of Arthur Loveridge, was his best. Loveridge was a specialist on Africa, particularly East Africa. He resented taking care of anything else. Loveridge trained Benjamin Shreve to take care of the non-African material. Barbour found this particularly useful. As Shreve complained to me, the several West Indian papers by Barbour and Shreve always took the same course: Shreve did the spade work, and Barbour did the florid introductions; Barbour was always the first author.

Barbour was independently wealthy, and his greatest service was to fund collectors, in particular Noble, Dunn, and Darlington, and also to buy collections. Loveridge pointed out to me that the Great Depression hit him hard, that after 1930 there were no acquisitions compa-

nable to those before that year and far fewer African trips for Loveridge.

Barbour composed three lists of Antillean reptiles and amphibians, in 1930, 1935, and 1937. All are limited to distributional comments of the sort that Anthony Curtis justly rebuked, and the third list must be stigmatized as erroneous because he used trinomials to show “races that are obviously closely related.” Unfortunately, in the group that I know most about, he got the “obvious relationships” wrong. *Anolis krugi* is not obviously close to *acutus*, *wattsii*, *forrestii*, nor *gingivinus*, all of which Barbour labeled as subspecies of *krugi*. Schwartz and Henderson (1991), following Lazell (1972), treat these “subspecies” as three full species (*krugi*, *acutus*, and *gingivinus*) and two subspecies (*wattsii watsii* and *wattsii forrestii*). *Anolis krugi* is more closely related to *pulchellus* and *poncensis*, as both Stejneger (1904) and Schmidt (1926) had already realized.

Period VI

The sixth period begins with Schwartz’s first trip to Cuba. Schwartz is the towering figure of this phase: His legendary NSF proposal (it was refused) to collect all the West Indies herpetofauna, like his near achievement of that goal, were the signal and significant events of this phase.

He did what he pleased and he did it well. He found the area inadequately collected, partly because there were fallacious predictions that there was little to collect. He had found in that first trip to Cuba that the West Indian fauna was very different from the mainland fauna: animals tended to be abundant there, not only the common widespread species but also local populations. He deliberately went off the beaten track, examined areas that had not been seen before, looked for at least local variation within species, found it, and described it. This self-imposed task avoided the vicious presumption that islands were devoid of interest because they were relatively depauperate; he demonstrated the old adage – not for the continent of Africa but for the West Indies – that from the islands, there comes always something new.

He could be critical of others. I experienced some of his criticism. He noticed that I had misspelled *bahorucoensis* throughout one paper and recorded that fact with a (justified) sic after each repetition of the misspelling *baharucoensis* in all the multiple editions of his coauthored checklists. He further complained that I mispronounced

cybotes with a hard c and took issue with my pronunciation of *Wetmor-ena* for that genus, pointing out that the genus *Wetmore-na* was supposed to honor Alexander Wetmore.

His mode of operation was intended to be professional in all that he did: i.e., (1) to be precise about localities, (2) to take detailed notes of color in life, (3) to collect large series whenever possible, (4) to select a single specimen as holotype, (5) to describe the scales and other external features of all specimens of the taxa carefully and in detail, and (6) to spell exactly, and to ensure that pronunciations honored whomsoever they were intended to honor. He succeeded.

He was not always the discoverer, but he was certainly the inspiration. Let's call the role of some startling surprises which the West Indies has provided: Schwartz himself described three anoles that can only be described as startling, the aquatic *eugeneagrahami*, the primitive *sheplani*, and the especially problematic *fowleri*. The ovoviviparous *Eleutherodactylus jasperi* is another species evoking incredulity. Astounding is the fact that it was not until 1966 that Richard Thomas described *Eleutherodactylus coqui*, the most common frog in Puerto Rico, the call of which gives it its name. Also amazing is the distribution of *Phyllodactylus* in the West Indies, Barbados (*pulcher*), both Haiti and the Dominican Republic, and Puerto Rico. [Schwartz (1979) described both of the disjunct Haitian (*sommeri*) and Dominican Republic populations (*hispaniolae*) as distinct species; the Puerto Rican population is the nominate one, *Phyllodactylus wirshingi* Kerster and Smith, 1955.]

Schwartz was primarily a collector and describer, not a jealous collector nor a jealous describer but rather he was genuinely interested in presenting what he believed to be true to the world at large.

He was a collector par excellence. He loved collection for its own sake, so he collected large series. There were larger samples for any problems that came to his notice. He was fortunate also in his choice of the West Indies; the West Indies had fewer taxa than the mainlands but usually greater abundances of the fewer taxa. For him, the West Indies was heuristic and enabling.

He published. Oh how, he published! He published alone or with (usually) one collaborator. He never published just for publication's sake; he was "getting the information out." He was fanatic about this. His ambitions were great, he wanted to collect every West Indian species, and it appears as though he nearly succeeded. Certainly the species he and his cohorts missed are very local (except in Cuba, for extraneous political reasons) and/or have a very special microhabitat.

As noted previously, he did not accomplish these feats alone. His collaborators comprise a formidable list. More than 50 are listed by name in Schwartz and Henderson (1991). The list almost included me, or perhaps it was Rodolfo Ruibal that was the attraction. When Ruibal and I were exploring the Cuban members of the *Anolis homolechis* group, Schwartz at one time orally suggested that he had some ideas and information about that and might join us. Almost immediately he withdrew, suggesting that it might prove complicated. (It is noticeable that he almost always collaborated with one author at a time.)

While Schwartz and I overlapped at times in the field and though, at one time we corresponded almost daily, we never did join forces. My primary concern with anoles was too limited for him.

His assistants and collaborators were diverse. He included some curators who were his students at first (e.g., Zug and Thomas). Zug was eventually distracted by the concerns of the National Museum; Thomas, who had teaching duties at the University of Puerto Rico, remained active in the West Indies, and alone or with Schwartz, has vied in publication numbers with his mentor. He also collaborated with McCoy at Carnegie (on subspecies of *Ameiva auberi*) and with Günther Peters of the Berlin Museum (on *Sphaerodactylus bromeliarum*; his only paper in German). His assistants in the field were mostly funded by himself. (He, like Barbour, was wealthy in his own right. Fortunately, and also fortunately for the West Indies, he was never a dilettante.)

If he specialized at all, he did so by genera and species complexes. Thus, from 1956 through 1967 he described (literally) an incredible and incredibly varied number of species and subspecies. He also reviewed and discussed alone or with one collaborator *Ameiva lineolata*, *A. taeniura*, and *A. chrysolema*, the Hispaniolan snakes then called *Dromicus*, the *pardalis-maculatus* complex of *Tropidophis*, *Leiocephalus* of the *carinatus*, *cubensis*, and *raviceps* groups, the *scaber* group of Cuban *Sphaerodactylus*; and the *vincenti* group of the Lesser Antilles; the sphaerodactyls of the Greater Puerto Rico region; the large toads of Cuba; and numerous other topics as well.

He never discussed the phylogeny of any group. The nearest that he came to doing so was his paper with Strahm on the characters of anguid osteoderms, and this for him was essentially a question of definition of genera – of what genus to which to assign a particular species. It was not a problem of relationship at all; it was solely a naming problem.

The other paper in which I remember a dendrogram (not a cladogram) is Schwartz and Thomas (1983) “The *difficilis* complex of

Sphaerodactylus (Sauria, Gekkonidae) of Hispaniola.” I am impressed by the fact that the paper is divided into two parts: Part I, authored solely by Schwartz on three species of the complex; and Part II authored by Thomas as first author and Schwartz as second author on five species of the complex and the conclusions, including the dendrogram. I assume that Schwartz was at least mildly uncomfortable with branching patterns. I point out that there are two unresolved terminal polytomies on the two branches on the dendrogram. Schwartz was clearly not one to regard precise branching patterns as essential.

He was also not primarily a biogeographer. He wrote only two specifically biogeographical papers: “Some Aspects of the Herpetogeography of the West Indies” in the 1975 Leidy Medal Symposium and “The Herpetogeography of Hispaniola, West Indies” (Schwartz, 1980b). In the latter he documented the concept of north and south island herpetofaunas relying not only on Mertens’ suggestion (1939) and Williams’ hypothesis (1961), but also on a geological reference (Khudoley and Meyerhoff, 1971) which asserts a fusion of two islands as a demonstrable fact.

He was fond of subspecies, inordinately some would say. He described many species, but the number of subspecies to species is about four to one. There are 40 described subspecies of *Ameiva auberi*, 29 of them Cuban (or the close satellite islands) and the rest Bahaman; and 17 subspecies of *Anolis distichus*, 12 of these Hispaniolan or on close satellite islands. Most of these were named by Schwartz alone or in collaboration.

There was always some flavor of reality to Schwartz’s subspecies. At the least, he described geographic variation within species elaborately and recognized the utility of naming “entities.” My current bias is that if there is not free exchange of genes, the populations should be called species, not subspecies (i.e., binomials and not trinomials). This might result in an enormous (excessive?) increase in the number of binomials, but I think that this complexity might reflect reality. I favor a case by case reexamination. If subspecies are not species, they are probably clines or morphs, not recognizable geographic units.

I have myself nibbled away at some of his taxa. With Susan Case, I have investigated certain of the distichoid anoles. Schwartz described approximately 20 subspecies in this complex. In one case – *distichus dominicensis* x *d. ignigularis* – there was a distinct hybrid zone with rare alleles and also relatively far away (near the north coast, in fact) there was an area of polymorphism in which dewlap color, the most conspicuous difference between the two “races,” ran the whole range between the two “defining” conditions. In the other case in the Bara-

hona region the absence of "interaction" between *distichus ravitergum* and *distichus favillarum* was accomplished by the intervention of a third member of the distichoid anoles, the indubitable species *brevirostris* between them. (This also occurs in another potential contact area. To the east, where *distichus ravitergum* should meet *distichus dominicensis*, there is a band of *brevirostris* between the two.) I do not present these cases as solved but as evidence of the presence of "complications."

The Seventh Period?

What comes after Schwartz? What will the seventh period in the history of the knowledge of the herpetofauna of the West Indies be like? It will concern itself more with history, and with cladogenesis and systematics. It will endeavor to date and document the events that produced the herpetofauna. My view is that this will be a more laborious process than the current enthusiasms may expect.

There will more species recognized in the future, but fewer will be discovered. Thus, the real legacy of Albert Schwartz is to leave the basic data complete enough to make unmistakable the incompleteness of our knowledge of the phylogeny and biogeography of the herpetofauna and raise the consciousness of the difficulties which we face in its analysis.

A Review of Cuban Herpetology

Alberto R. Estrada¹ and Rodolfo Ruibal²

¹*Department of Biology, University of Puerto Rico, P.O. Box 38030, San Juan, Puerto Rico 00937-1030*

²*Department of Biology, University of California, Riverside, California 92521-0427*

History

The first written account of Cuban herpetology, according to Poey (1865), was in the reports of Christopher Columbus in which he mentioned iguanas and crocodiles in the newly discovered islands. Gonzalo Oviedo in 1526 (1959) published his *Historia Natural de las Indias* and also wrote of crocodiles and iguanas, though it is unclear whether he was referring to Cuba or Hispaniola. Oviedo does mention Cuban sea turtles when describing their method of capture, by the natives, with remoras.

The formal taxonomy of Cuban herpetology starts with the description of *Typhlops lumbricalis* by Linnaeus in 1758. However, the type locality of the specimen was listed as "America." In 1807 Cuvier described *Crocodylus rhombifer*. The first major contribution to Cuban herpetology was in Ramon de la Sagra's *Historia Fisica, Política, y Natural de la Isla de Cuba*, published over a period of years (1838-1853). The herpetological section was written by Coccoateu and Bibron and included 34 species, of which 14 were new. Much of this publication was based on specimens brought to Paris by de la Sagra. Separate printings were made in French and Spanish and the actual dates of publication are ambiguous (see Smith and Grant, 1958). Gundlach (1880) specifically states that the herpetological part was first published in volume 6 in Spanish in 1843. Duméril and Bibron had also published Cuban species in their 1836-1841 *Erpétologie Générale*. Ramon de la Sagra was born in Spain but resided in Cuba for most of his life. He was director of the botanical gardens in Habana between 1824 and 1836 and was the founder of the first Cuban scientific publication; *Anales de Ciencias, Agricultura, Comercio y Artes de la Habana*, in 1827.

Alexander Humbolt traveled in Cuba during 1800 and 1801. In his *Island of Cuba* (1856) Gundlach discusses the differences between the "caiman" (*C. acutus*) and the "cocodrilo" (*C. rhombifer*).

One of the most prominent figures in Cuban natural history was Felipe Poey y Aloy. Between 1851 and 1858 Felipe Poey published in Cuba his *Memorias Sobre la Historia Natural de la Isla de Cuba*. This was the first Cuban publication devoted exclusively to zoological matters. The main herpetological component of this publication was a detailed description by Poey of the circulatory system of *C. acutus* and *C. rhombifer* and a discussion of the aortic circulation and the fact that the head of crocodiles receives a more oxygenated blood than the body. In the same article he also denies the existence of a "peritoneal respiration" that had been proposed by St. Hilaire to be present in crocodiles. Poey was the first director of the Museo de Historia Natural in Habana that was founded in 1838. These collections now reside in the Museo Felipe Poey de la Universidad de la Habana.

Charles Wright, a United States resident in Guantánamo, Cuba, during the middle of the nineteenth century was a collector of botanical and zoological specimens. He sent his herpetological material to the United States National Museum. E. D. Cope, between 1861 and 1864, described 6 new species of amphibians and 10 new species of lizards based on Wright's specimens and other specimens sent to Philadelphia by Poey.

During 1865-1868, Poey also published the *Repertorio Físico Natural de la Isla de Cuba*. It is in this publication that Juan Gundlach (1867) authored his first checklist of the Cuban herpetofauna. Gundlach was born and educated in Germany but came to Cuba as a young man (Adler, 1989). Gundlach and Poey collected and published on Cuban natural history, Poey concentrating on fishes and Gundlach on herpetology and other vertebrates. Gundlach's 1868 list contains 67 species of amphibians and reptiles. He provided distributional and ecological data for many of the species, having observed most of them in the field. He comments on the fact that by this date *Cyclura* had already become rare where once it was abundant along the coast and keys.

The checklist includes the common names for many of the species. He lists 16 species of *Anolis* and his comments on the ecology and color indicate that he had observed all of them in the field. Gundlach traveled extensively throughout Cuba. *Cricosaura typica*, which he and W. Peters described in 1863, was collected by Gundlach in Cabo Cruz, an isolated area in eastern Cuba. He apparently lived in Cabo Cruz with the turtle fishermen. He lists four sea turtle species as

known and observed by him or Felipe Poey: *Chelonia mydas*, *Eretmochelys fimbriata*, *Caretta caretta* and *Dermochelys coriacea*. The fifth species, *Chelonia virgata* (currently a synonym of *C. mydas*), he lists but states that he has never seen it.

In 1880 Gundlach publishes his book *Contribucion a la Erpetologia Cubana* in Habana with a total of 71 species of amphibians and reptiles. Barbour and Ramsden (1919) provides a detailed listing of the species in Gundlach. Gundlach's opus is more than a checklist and contains extensive information about the natural history of many of the species and detailed descriptions of the color patterns of species that he collected. Gundlach kept live specimens of *Epicrates angulifer*, *C. rhombifer*, and *C. acutus* and gave detailed information about their behavior in captivity. Barbour and Ramsden (1919) translate and quote a long section from Gundlach detailing the Cuban crocodiles.

After Gundlach's 1880 publication 39 years, lapsed before the Cuban herpetofauna was again reviewed. In 1919 Thomas Barbour and Charles T. Ramsden published *The Herpetology of Cuba* through Harvard's Museum of Comparative Zoology. Barbour had traveled and collected extensively in Cuba and Ramsden was also a collector and lived in Guantánamo. Barbour and Ramsden carefully reviewed Gundlach's work and the other publications referring to Cuban species since 1880. During this time the total number of species had changed only slightly: Gundlach listed 71 species, Barbour and Ramsden listed 73 (see Fig. 2.1). Barbour's contribution to Cuban herpetology included numerous other papers. The most important were the 1914 "A Contribution to the Zoogeography of the West Indies, with Special Reference to Amphibians and Reptiles" and the 1921 "*Sphaerodactylus*," both publications having a substantive Cuban component. In 1937 he published his "Third List of Antillean Reptiles and Amphibians" raising the number of Cuban species to 89, and introducing trinomials for a number of the species for the first time.

During the 1930s and 1940s a total of 11 new species were added to the Cuban herpetofauna. Barbour and Shreve were the major contributors with a total of 6 new species (4 *Eleutherodactylus* and 2 *Anolis*).

Albert Schwartz's contribution to West Indian herpetology started with his 1956 paper with L. H. Ogren that was based on a collection of amphibians and reptiles that he and Ogren collected in Cuba. For the next 35 years, Albert Schwartz was the major taxonomist of West Indian herpetology (see Bourgeois, 1995). In his papers on Cuban herpetology he added 23 new species to the fauna. Schwartz's papers

were on taxonomy and distribution and covered all groups of reptiles and amphibians. After the Cuban revolution of 1960 he collaborated with the Cuban herpetologist Orlando Garrido and between 1967 and 1985 they coauthored 14 papers.

Ernest Williams and Rodolfo Ruibal also contributed to Cuban herpetology during the 1960s with publications on the anoline lizards (see Rodriguez-Schettino, 1995, for these and other references).

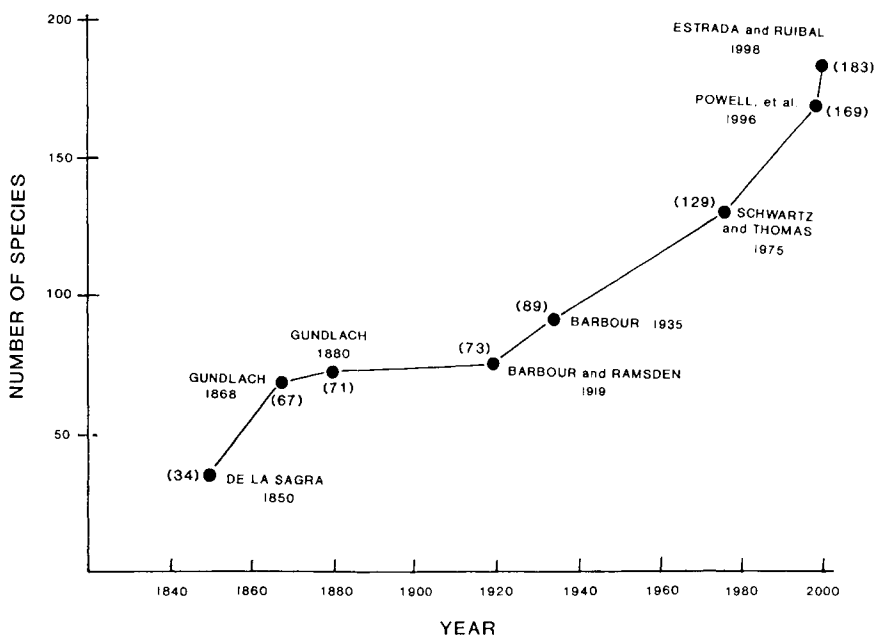


Figure 2.1. The number of recognized species in the Cuban herpetofauna. The totals include marine turtles.

The Cuban Revolution of 1959 ushered in a new era for Cuban science. The revolution restructured the academic and scientific institutions of Cuba (Margulis and Kunz, 1984). During the 1960s changes were initiated in the universities and all the other scientific entities (see Pruna Goodgall, 1994). The restructuring of the Academia de Ciencias de Cuba was accompanied by a surprising growth in scientific research and publication, including herpetology. Much of the research has been published in the new Cuban scientific journals published by the academy: *Ciencias Biológicas*, *Poeyana*, *Reporte de Investigacion del Instituto de Ecología y Sistemática*, and *Miscelania*

Zoológica. Publications of other Cuban institutions include *Revista Cubana de Biología*, *Garciana*, *Memorias de la Facultad de Ciencias* and *Torreia*. These publications, and others, contain much herpetological research carried out by the new and large cadre of Cuban biologists produced by the revolution.

The magnitude of the change in Cuban herpetology since 1960 can be gauged by tabulating the first authors of the articles cited in the iguanid bibliography of Rodríguez Schettino (1995). The bibliography lists 68 papers by non-Cuban first authors and 66 papers by Cuban first authors. With few exceptions the Cuban-authored articles are post-1960. Orlando Garrido and Alberto R. Estrada are major contributors, but many other Cuban herpetologists have made substantive contributions. Lourdes Rodríguez-Schettino has made numerous ecological contributions, and Octavio Pérez-Beato, Vincente Berovides, and Georgina Espinosa López have provided electrophoretic analyses of numerous species groups. Many of the Cuban-authored papers represent a new emphasis on ecology and the evolutionary relationship of the species rather than the description of new forms.

A further example of the increased participation of Cuban scientists can be seen by comparing the number of species in the 1996 checklist (Powell *et al.*, 1996a) described by Cubans (authored or coauthored) prior to 1970 with the number post 1969. A total of 8 species were by Cuban biologists prior to 1970, whereas 36 new species were by Cubans after 1969.

During the 1990s major contributions to the Cuban herpetofauna were made by S. Blair Hedges. In collaboration with Orlando Garrido and Alberto Estrada, he added more than 15 new species to the Cuban fauna.

The Species

Geographic Regions

With some simplifications of the map of Mateo and Acevedo (1989), Cuba can be divided into four geographic regions: Western, Central, Camagüey-Maniabón and Eastern (Fig. 2.2). In totality the regions contain more than 20 subregions.

The Western section is subdivided into seven subregions. The Plains of Guanahacabibes, which includes the peninsula with the same nature, and is a karst plains. The Cordillera de Guaniguanico, composed of two main ranges, the Organos and the Rosario ranges, has a maximum elevation of 699 m at the Pan de Gujaibón. The Sierra de los Organos is formed by a chain of large isolated blocks of limestone

called "mogotes," and it has numerous interior valleys and is bordered to the north and south by the clay hills the Alturas de Pizarras del Norte and Sur. The Sierra del Rosario occupies the eastern half of the Cordillera de Guaniguanico and is composed of conical hills. The cordillera is surrounded by the southern and northern plains of Pinar del Río. The eastern part of the region contains the subregions (north to south) of the Alturas de La Habana-Matanzas (up to 381m), the Alturas Bejucal-Coliseo, the Llanura de Antemisia-Colón, and the Llanura de Zapata. The first of these is a range of coastal hills, marine terraces and limestone sierras. The Alturas de Bejucal-Coliseo are formed by groups of isolated, and primarily limestone, elevations that extend from the center of the region to the east between the territories of La Habana and Matanzas, with a maximum elevation of 312 m. The Llanura de Antemisia-Colón flank this subregion on the south and is a continuation of the Llanura Sur of Pinar de Río. Currently 80% of this part of the Western region is devoted to agriculture. The Llanura de Zapata is formed by a karst plain that forms the Zapata Peninsula. This is covered by extensive forests, swamp vegetation, and mangrove and constitutes the major moist region of the archipelago and one of the major forest reserves of Cuba.

The Central region is the smallest and is divided into three subregions: The Llanura Corralillo-Yaguajay to the north, the Alturas y Llanos de Cubanacán in the central portion, and to the south the mountains of Guamuhaya, also known as the Escambray. This last subregion contains the Sierra de Trinidad, which occupies most of the western portion, and the Sierra de Sancti Spiritus, which occupies the most eastern portion. Both are separated by the basin of the Río Agabama-Manatí. The maximum elevation of this region is located in the Sierra de Trinidad at Pico San Juan (1140 m).

The Camagüey-Maniabón region has in its central portion a chain of low elevations and valleys called Alturas del Centro de Camagüey. The most prominent is the Sierra de Cubitas, a range of low (maximum elevation 330 m) limestone and serpentine hills, located in the northwest. The central southern area contains the Sierra de Najasa (301 m). In the extreme east is the Alturas de Maniabón-Banes (347 m). These hills are surrounded by the ample Llanura de Norte and Llanura de Sur, which meet in west to form the Llanura de Júcaro-Morón.

The Eastern region is the most mountainous in Cuba. Its western limit is the Llanura del Cauto, which is an alluvial zone with the largest river in Cuba, the Cauto. To the south and to the east along the length of the coast extends the Sierra Maestra, with the highest peak

in Cuba, Pico Turquino, at 1972 m. The Cuenca de Santiago is a depression that separates the Sierra Maestra from the Sierra de la Gran Piedra (1214 m) and continues to the east to the Cuenca de Guantánamo. A narrow valley to the north separates the eastern portion of the Sierra Maestra, the Cuenca de Santiago, and the Sierra de la Gran Piedra from the Nipe mountains (995 m) and the Sierra de Cristal (1231 m). Located on the eastern tip of the island are the Sagua-Baracoa mountains (1175 m), which contain the ridges of Moa, Toa, and Baracoa, and the fluvial basins of Jaguaní-Toa and Duaba.

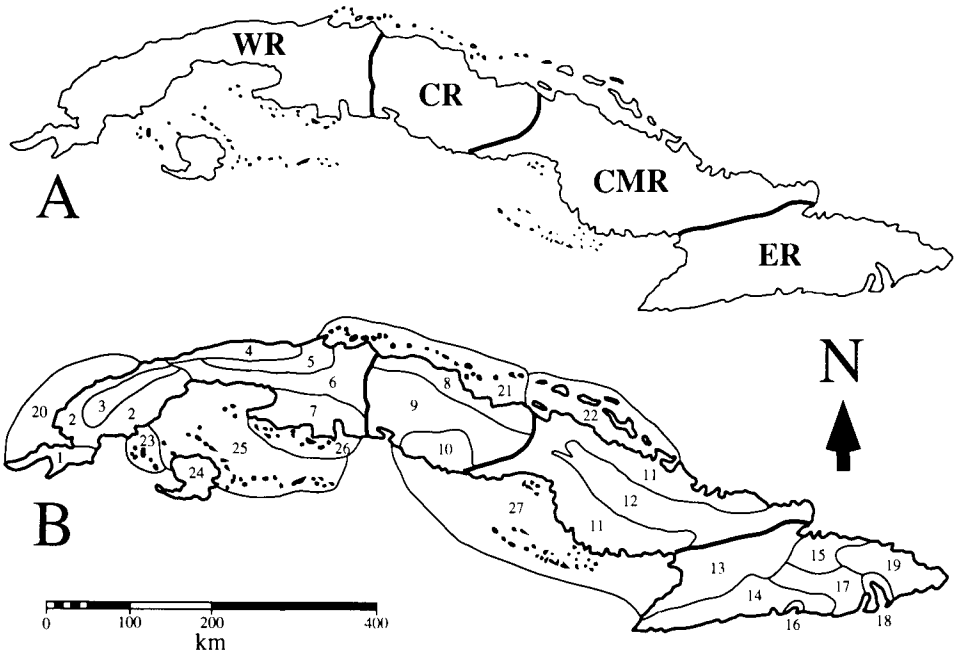


Figure 2.2. (A) Geographic regions of Cuba. WR, western region; CR, central region; CMR, Camagüey-Maniobón region; ER, eastern region. (B) Subregions: 1, Llanura y Península de Guanahacabibes; 2, Llanuras norte y sur de Pinar de Río; 3, Cordillera de Guaniguanico; 4, Alturas de La Habana-Matanzas; 5, Alturas de Bejucal-Coliseo; 6, Llanura Artemisa-Colón; 7, Llanura y Península de Zapata; 8, Llanura Corralillo-Yaguajay; 9, Alturas y Llanos de Cubancán; 10, Sierra de Guamuhaya; 11, Llanuras de norte y sur de Camagüey; 12, Alturas y Llanos de Camagüey-Maniobón; 13, Llanura de Cauto; 14, Sierra Maestra; 15, Alturas de Nipe-Cristal; 16, Cuenca de Santiago de Cuba; 17, Valle Central; 18, Cuenca de Guantánamo; 19, Montañas de Sagua-Baracoa; 20, Archipiélago de los Colorados; 21, Archipiélago de Sabana; 22, Archipiélago de Camagüey; 23, Cayos de San Felipe; 24, Isla de la Juventud; 25, Archipiélago de los Canarreos; 26, Cayos Blancos del Sur; 27, Archipiélago de Jardines de la Reina (Mateo and Acevedo, 1989).

The Eastern region also includes the marine terraces of the Meseta de Maisí (633 m), the Sierra de Purial (1059 m), Imías (1176 m) and the Meseta de Guaso (890 m). Other subregions are the Isla de la Juventud in the south and along the north coast the archipelagos of the Colorados, Sabana, and Camagüey. Along the south coast the abundant keys of the Canarreos and the Jardines de la Reina. Other keys are located to the northwest of the Isla de la Juventud (the Cayos de San Felipe), and to the south of the Zapata Peninsula (the Cayos Blancos de Sur).

Vegetation

The map of the natural vegetation of Cuba provided by Capote *et al.* (1989) recognizes 30 different botanical associations. In a simpler form the vegetation can be classified into four major groups: (i) broad-leaf tropical forests; (ii) tropical coniferous forests; (iii) tropical thickets; and (iv) herbaceous communities. The broad-leaf tropical forests contain two major divisions: (i) evergreen forests and (ii) semi-deciduous forests. The coniferous forests are characterized by four species of pines. In the west and Isla de la Juventud contain *Pinus caribea* and *P. tropicalis*, whereas the eastern portion of Cuba contain *P. cubensis* and *P. maestrensis*. The Central regions of Camagüey-Maniobón do not have natural pines. The thickets include the dry coastal associations, the vegetation on serpentine soils, and on the limestone mogotes. The herbaceous communities include the swamp vegetation formed by *Cladium jamaicense* and *Typha dominiguensis*.

During the nineteenth and twentieth centuries, Cuba was transformed from a predominantly forested island into arable agricultural land. Smith (1954) estimates that pre-Columbian Cuba was 60% forest and that only 15% of the forest survived. Similarly, Seifriz (1943) concluded that "little, if any, of Cuba's arable land has been left uncultivated". Since 1989 deforestation has increased due to the economic crisis and the lack of fossil fuels on the island. How this ecological transformation has affected the herpetofauna has gone unstudied. It is obvious that such extreme deforestation has modified the distribution of many species.

Amphibians

Of the 57 known species, 7 are bufonids and there is 1 hylid, 46 leptodactyls, and 1 introduced ranid (Table 2.1; Fig. 2.3). Two other species have been reported as introduced into Cuba, the bufonid *Bufo marinus* and the hylid *Pseudacris crucifer*.

Table 2.1. Cuban Amphibians^a

Species	Status	Distribution
CLASS AMPHIBIA, ORDER ANURA		
Family Bufonidae		
<i>Bufo cataulaciceps</i>	Endemic	WR, IJ
<i>Bufo empusa</i>	Endemic	IW, IJ
<i>Bufo fustiger</i>	Endemic	WR
<i>Bufo gundlachi</i>	Endemic	IW, IJ
<i>Bufo longinasus</i>	Endemic	
<i>B. l. longinasus</i>		WR
<i>B. l. cajalbanensis</i>		WR
<i>B. l. dunni</i>		CR
<i>B. l. ramsdeni</i>		ER
<i>Bufo peltocephalus</i>		CR-ER, IJ, ASC
<i>Bufo taladai</i>	Endemic	CR-ER
Family Leptodactylidae		
<i>Eleutherodactylus acmonis</i>	Endemic	ER
<i>Eleutherodactylus albipes</i>	Endemic	ER
<i>Eleutherodactylus atkinsi</i>	Endemic	
<i>E. a. atkinsi</i>		IW, IJ, ASC
<i>E. a. estradai</i>		ER
<i>Eleutherodactylus auriculatus</i>	Endemic	IW, IJ
<i>Eleutherodactylus bartonsmithi</i>	Endemic	ER
<i>Eleutherodactylus bresslerae</i>	Endemic	ER
<i>Eleutherodactylus casparii</i>	Endemic	CR
<i>Eleutherodactylus cubanus</i>	Endemic	ER
<i>Eleutherodactylus cuneatus</i>	Endemic	ER
<i>Eleutherodactylus dimidiatus</i>	Endemic	
<i>E. d. dimidiatus</i>		IW
<i>E. d. amelasma</i>		ER
<i>Eleutherodactylus eileenae</i>	Endemic	WR-CMR
<i>Eleutherodactylus emiliae</i>	Endemic	CR
<i>Eleutherodactylus etheridgei</i>	Endemic	ER
<i>Eleutherodactylus glamyrus</i>	Endemic	ER
<i>Eleutherodactylus goini</i>	Endemic	WR
<i>Eleutherodactylus greyi</i>	Endemic	CR
<i>Eleutherodactylus guanahacabibes</i>	Endemic	WR
<i>Eleutherodactylus Guantanamoera</i>	Endemic	ER
<i>Eleutherodactylus gundlachi</i>	Endemic	ER
<i>Eleutherodactylus iberia</i>	Endemic	ER
<i>Eleutherodactylus intermedius</i>	Endemic	ER

Table 2.1 cont.

<i>Eleutherodactylus ionthus</i>	Endemic	ER
<i>Eleutherodactylus jaumei</i>	Endemic	ER
<i>Eleutherodactylus klinikowskii</i>	Endemic	WR
<i>Eleutherodactylus leberi</i>	Endemic	ER
<i>Eleutherodactylus limbatus</i>	Endemic	IW
<i>Eleutherodactylus orientalis</i>	Endemic	ER
<i>Eleutherodactylus mariposa</i>	Endemic	ER
<i>Eleutherodactylus melacara</i>	Endemic	ER
<i>Eleutherodactylus pezopetrus</i>	Endemic	ER
<i>Eleutherodactylus pinarensis</i>	Endemic	WR, IJ
<i>Eleutherodactylus planirostris</i>	Cuba and Bahamas	
<i>E. p. planirostris</i>		IW, IJ, ASC
<i>Eleutherodactylus principalis</i>	Endemic	ER
<i>Eleutherodactylus ricordii</i>	Endemic	ER
<i>Eleutherodactylus sp. n.</i>	Endemic	WR-CMR, IJ
<i>Eleutherodactylus ronaldi</i>	Endemic	ER
<i>Eleutherodactylus symingtoni</i>	Endemic	WR
<i>Eleutherodactylus tetajulia</i>	Endemic	ER
<i>Eleutherodactylus thomasi</i>	Endemic	WR-CMR
<i>E. t. thomasi</i>		CMR
<i>E. t. trinidadensis</i>		CR
<i>E. t. zayasi</i>		WR
<i>Eleutherodactylus toa</i>	Endemic	ER
<i>Eleutherodactylus tonyi</i>	Endemic	ER
<i>Eleutherodactylus turquinensis</i>	Endemic	ER
<i>Eleutherodactylus varians</i>	Endemic	IW
<i>E. v. varians</i>		CR-ER
<i>E. v. olibrus</i>		WR
<i>E. v. staurometopon</i>		IJ
<i>Eleutherodactylus varleyi</i>	Endemic	IW
<i>Eleutherodactylus zeus</i>	Endemic	WR
<i>Eleutherodactylus zugi</i>	Endemic	WR
<i>E. z. zugi</i>		WR
<i>E. z. erythroproctus</i>		WR
Family Hylidae		
<i>Osteopilus septentrionalis</i>	Cuba and Bahamas	IW
Family Ranidae		
<i>Rana catesbeiana</i>	Introduced	IW

*The letters in the distribution column refer to the following geographic regions: WR, Western region; CR Central region; CMR, Camagüey-Maniabón región; ER Eastern region; ASC Archipié-

Iago de Sabana-Camagüey; IW, Islandwide and IJ, Isla de la Juventud (Mateo and Acevedo, 1989).

Bufo marinus was introduced in 1946 (Buide, 1967) and, unlike on the other Antillean islands, the introduction was unsuccessful in Cuba. Currently, there is no evidence of a resident population of *B. marinus* on Cuba. The existence of the populations of *P. crucifer* reported in Cuba by Schwartz and Thomas (1975) and by Schwartz and Henderson (1988, 1991) have never been confirmed. For more than 10 years attempts to locate the frogs near Canasí and Marianao have failed. Consequently, neither of these species have been included in the list of Cuban amphibians.

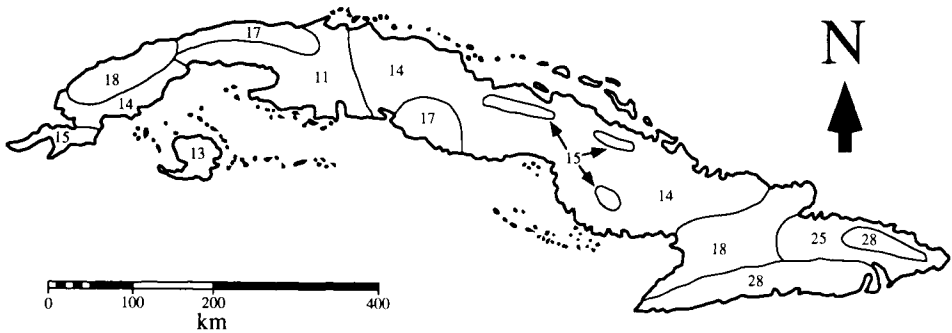


Figure 2.3. Estimated number of amphibian species in the different regions of Cuba. The numbers indicate the number of sympatric species and exclude species known only from a single locality.

The seven bufonid species are endemic to Cuba. *Bufo gundlachi* and *B. empusus* have a wide distribution in the lowlands and also on Isla de la Juventud and on the Coco, Romano, and Guajaba keys of the Sabana-Camagüey archipelago. *Bufo longinasus* is a polytypic species with four disjunct populations: two in the west, *B. l. longinasus* in the highlands of Pizarras del Sur, between the city of Pinar del Río and the Valle de Viñales, and *B. l. cajalbanensis* in the Cajalbana plateau (Valdés and Ruiz, 1980). *Bufo l. dunni* inhabits the mountains of

the Sierra de Guamuhaya in the areas of Trinidad as well as Sancti Spiritu (Valdés and Ruíz, 1980; Estrada, 1994). *Bufo l. ramsdeni* is the eastern-most form and inhabits the areas of Monte Verde and Monte Libano in the Guaso plateau (Valdés and Ruíz, 1980). All the populations of *B. longinasus* are associated with rivers and streams. *B. cataulaciceps* lives in the sandy plains in the west and south of the province of Pinar del Río and the northern part of the Isla de Juventud.

Bufo fustiger has a broad distribution in the lowlands and mountains of western Cuba, from the Guanahacabibes Peninsula to the borders of the Llanura de Zapata. *Bufo peltocephalus* resides in lowlands and moderate elevations from the Central region to extreme eastern Cuba, including the Santa María, Coco, Parédon Grande, Romano, Guajaba and Sabinal keys (Schwartz and Henderson, 1991). *Bufo taladai* is found in the lowlands and mountains of the Eastern region and some populations are known in the Central region in the Sierra Guamuhaya (Schwartz and Henderson, 1991).

The only members of the Hylidae is *Osteopilus septentrionalis*. Although it is not endemic to Cuba, it is widely distributed in Cuba and is found in lowlands, mountains, forests, savannas, mangroves, and agricultural areas. It is present on the Isla de la Juventud and the keys of Sabana-Camagüey and Canarreos. Outside of Cuba it is indigenous to the Bahamas and Cayman Islands and has been introduced to Puerto Rico, the Virgin Islands, and the keys and mainland of Florida.

The bulk of the amphibian species are leptodactylids in the genus *Eleutherodactylus*. Dunn (1926a) divided the Cuban *Eleutherodactylus* into four groups: *auriculatus*, *dimidiatus*, *ricordi*, and *varleyi*. Schwartz (1957) added a fifth group, *symingtoni*. Hedges (1989) performed a phylogenetic synthesis of Antillean *Eleutherodactylus* and proposed three subgeneric categories: *Euhyas*, *Pelorius*, and *Eleutherodactylus*. The Cuban species fall within two subgenera: 12 species in *Eleutherodactylus* and 28 species in *Euhyas*. The species show four patterns of distribution within Cuba: islandwide (7 species), western (7 species), central (3 species), from the Western region to Camagüey-Maniabón (3 species), and the Eastern region (26 species).

Some of the western species are characterized by having local narrow distributions. *Eleutherodactylus guanahacabibes* is endemic to the Guanahacabibes Peninsula (Estrada and Novo, 1985b); *E. zeus* is only found in the Sierra de los Organos (Estrada *et al.*, 1989); *E. klinikowski* is known only from the Sierra de los Organos and a few localities in the Sierra del Rosario (Schwartz and Henderson, 1991); and *E. symingtoni* is only known from five localities from the Sierra de

los Organos to the Alturas La Habana-Matanzas (Estrada *et al.*, 1989). *Eleutherodactylus zugi* has two disjunct subspecies (Schwartz, 1960) with populations in the Cordillera de Guaniguanico (*E. z. zugi*) and in the Alturas La Habana-Matanzas (*E. z. erythroproctus*). *Eleutherodactylus pinarensis* exhibits an interesting pattern of distribution: Llanura de Guanahacabibes, Isla de la Juventud, and the rest of the Western region excluding the Llanura de Pinar del Río and the Cordillera de Guaniguanico (Estrada and Novo, 1985a). *E. goini*, a species once considered a subspecies of *E. planirostris* (Estrada and Hedges, 1997b) is found in the forest spurs of the Cordillera de Guaniguanico.

The three species with an exclusively Central region distribution are: *E. caspari*, *E. emiliae* (both endemic to the Sierra de Guamuhaia), and *E. greyi*, which also has some populations in Camagüey-Maniabón region in the Sierra de Cubitas and Sierra de Najasa (Schwartz and Henderson, 1991). From the Western region to Camagüey-Maniabón are found *E. eileenae*, *E. sp. n.* and *E. thomasi*. *Eleutherodactylus sp. n.* was long confused with *E. cuneatus*, a species with similar riparian habits (Estrada and Hedges, 1998). *Eleutherodactylus thomasi* has three subspecies: *E. t. thomasi* in the Cubitas and Najasa mountains of Camagüey-Maniabón, *E. t. trinidadensis* on the south slopes of the Sierra de Guamuhaia, and *E. t. zayasi* from the Pan de Matanzas in the Alturas de La Habana-Matanzas. Additionally, some populations that have not been subspecifically identified are known from north of the Central region and from Cayo Coco (Schwartz and Henderson, 1991; Estrada, 1993b).

Most species of *Eleutherodactylus* have an Eastern region distribution and 18 of these (*E. albipes*, *E. bartonsmithii*, *E. bresslerae*, *E. cubanus*, *E. etheridgei*, *E. glamyrus*, *E. iberia*, *E. intermedius*, *E. jaumei*, *E. leberi*, *E. orientalis*, *E. mariposa*, *E. melacara*, *E. pezopetrus*, *E. principalis*, *E. tetajulia*, *E. tonyi*, and *E. turquinensis*) are known from a single locality or from a very limited area (Estrada and Alonso, 1997; Estrada and Hedges, 1996a, 1997a; Hedges *et al.*, 1992a, 1995; Schwartz and Henderson, 1991). The other 8 Eastern region species have a broader distribution in the region (*E. acmonis*, *E. cuneatus*, *E. guantanamera*, *E. gundlachi*, *E. ionthus*, *E. ricordii*, *E. ronaldi*, and *E. toa*).

Seven species have a pan-Cuban distribution: *E. atkinsi*, *E. auriculatus*, *E. dimidiatus*, *E. limbatus*, *E. planirostris*, *E. varians*, and *E. varleyi* (Schwartz and Henderson, 1991). There are two subspecies of *E. atkinsi*, one having a distribution throughout Cuba, Isla de la Juventud, and the keys of Sabana-Camagüey (*E. a. atkinsi*) and the

other (*E. a. estradai*) located in the Sagua-Baracoa mountains in the Eastern region (Schwartz and Henderson, 1991; Lynch, 1991). Three subspecies are recognized for *E. varians*; *E. v. varians* from the eastern portion of the Llanura de Zapata to the Camagüey-Maniabón region, *E. v. olibrus* from Cordillera de Guaniguanico, and *E. v. staurometopon* from the Isla de la Juventud. The populations of *E. varians* from the Llanura de Guanahacabibes and the Alturas de La Habana-Matanzas have not been assigned to any of the described subspecies (Schwartz and Henderson, 1991).

The only ranid species, *Rana catesbeiana* was introduced at the beginning of the nineteenth century and currently is widely distributed throughout Cuba and the Isla de la Juventud (Schwartz and Henderson, 1991).

Reptiles

There are 122 recognized species of reptiles in Cuba (Table 2.2; Fig. 2.4). There is a single pond turtle, *Trachemys decussata*, that is widely distributed in the swamps, rivers, ponds and dams of the entire archipelago, including most of the keys.

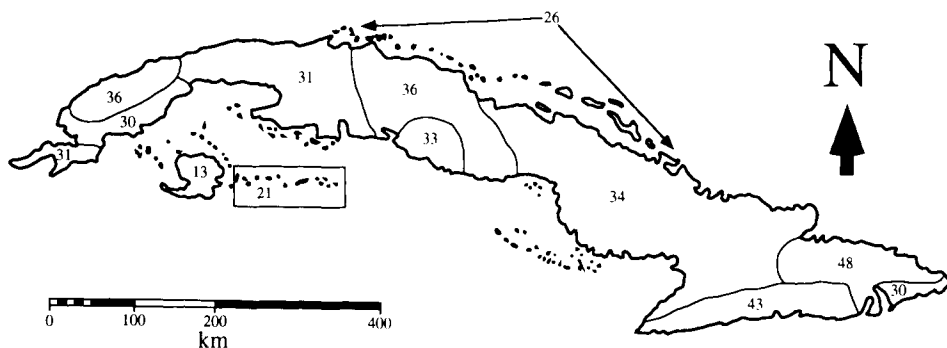


Figure 2.4. Estimated number of reptilian species in the different regions of Cuba. The numbers indicate the number of sympatric species and exclude species known only from a single locality.

Table 2.2. Cuban Reptiles^a

Species	Status	Distribution
CLASS REPTILIA, ORDER TESTUDINES		
Family Emydidae		
<i>Trachemys decussata</i>	Cuba, Cayman Is.	
<i>T. d. decussata</i>	Endemic	WR-ER
<i>T. d. angusta</i>		WR, IJ
ORDER SQUAMATA, SUBORDER AMPHISBAENIA		
Family Amphisbaenidae		
<i>Amphisbaena blanoides</i>	Endemic	ER
<i>Amphisbaena cubana</i>	Endemic	ER
<i>A. c. cubana</i>		CR-ER, IJ
<i>A. c. barbouri</i>		
<i>Amphisbaena palirostrata</i>	Endemic	IJ
SUBORDER LACERTILIA		
Family Anguidae		
<i>Diploglossus delasagra</i>	Endemic	IW, IJ
<i>Diploglossus nigropunctatus</i>	Endemic	ER
<i>Diploglossus garridoi</i>	Endemic	ER
Family Gekkonidae		
<i>Gonatodes albogularis</i>	Cuba, Jamaica, Cayman Is., and Hispaniola	
<i>G. a. fuscus</i>		IW
<i>Hemidactylus brooki</i>	Cuba, Hispaniola and Puerto Rico	
<i>H. b. brooki</i>		IW
<i>Hemidactylus maboui</i>	Cuba, Hispaniola and Puerto Rico	WR, ER
<i>Hemidactylus turcicus</i>	Introduced	
<i>H. t. turcicus</i>		WR, CR, ER
<i>Sphaerodactylus armasi</i>	Endemic	ER
<i>Sphaerodactylus argus</i>	Cuba and Jamaica	
<i>S. a. argus</i>		WR-CMR, AJR
<i>Sphaerodactylus bromeliarum</i>	Endemic	ER
<i>Sphaerodactylus celicara</i>	Endemic	ER
<i>Sphaerodactylus cricoderus</i>	Endemic	ER
<i>Sphaerodactylus docimus</i>	Endemic	ER
<i>Sphaerodactylus elegans</i>	Endemic	IW, IJ
<i>Sphaerodactylus intermedius</i>	Endemic	WR
<i>Sphaerodactylus nigropunctatus</i>	Cuba and Bahama Is.	
<i>S. n. alayoi</i>		ER

Table 2.2 cont.

<i>S. n. granti</i>		ASC
<i>S. n. lissodesmus</i>		CMR
<i>S. n. ocujal</i>		ER
<i>S. n. strategus</i>		ER
<i>Sphaerodactylus notatus</i>	Cuba and Bahama Is.	
<i>S. n. atactus</i>		IW, IJ
<i>Sphaerodactylus oliveri</i>	Endemic	CR
<i>Sphaerodactylus ramsdeni</i>	Endemic	ER
<i>Sphaerodactylus richardi</i>	Endemic	WR
<i>Sphaerodactylus ruibali</i>	Endemic	ER
<i>Sphaerodactylus scaber</i>	Endemic	CR-CMR, AS
<i>Sphaerodactylus storeyae</i>	Endemic	IJ, ACA
<i>Sphaerodactylus schwartzi</i>	Endemic	ER
<i>Sphaerodactylus torrei</i>	Endemic	
<i>S. t. torrei</i>		ER
<i>S. t. spielmani</i>		ER
<i>Tarentola americana</i>	Cuba and Bahama Is.	
<i>T. a. americana</i>		IW, ASC
Family Iguanidae		
<i>Cyclura nubila</i>	Cuba and Cayman Is.	
<i>C. n. nubila</i>	Endemic	IW, IJ
Family Polychrotidae		
<i>Anolis ahli</i>	Endemic	CR
<i>Anolis alayoni</i>	Endemic	ER
<i>Anolis alfaroi</i>	Endemic	ER
<i>Anolis allisoni</i>	Cuba and	IW
<i>Anolis allogus</i>	Endemic	IW
<i>Anolis alutaceus</i>	Endemic	IW, IJ
<i>Anolis anfiloquioid</i>	Endemic	ER
<i>Anolis angusticeps</i>	Cuba and Bahama Is.	
<i>A. a. angusticeps</i>		IW, IJ
<i>Anolis argenteolus</i>	Endemic	CMR-ER
<i>Anolis argillaceus</i>	Endemic	ER
<i>Anolis baracoae</i>	Endemic	ER
<i>Anolis barbatus</i>	Endemic	WR
<i>Anolis bartchi</i>	Endemic	WR
<i>Anolis birama</i>	Endemic	ER
<i>Anolis bremeri</i>	Endemic	
<i>A. b. bremeri</i>		WR

Table 2.2 cont.

<i>A. b. insulsepinorum</i>		IJ
<i>Anolis centralis</i>	Endemic	
<i>A. c. centralis</i>		CR-CMR
<i>A. c. litoralis</i>		ER
<i>Anolis chamaelionides</i>	Endemic	IW
<i>Anolis clivicola</i>	Endemic	ER
<i>Anolis confusus</i>	Endemic	ER
<i>Anolis cupeyalensis</i>	Endemic	ER
<i>Anolis cyanopleurus</i>	Endemic	
<i>A. c. cyanopleurus</i>		ER
<i>A. c. orientalis</i>		ER
<i>Anolis delafuentei</i>	Endemic	CR
<i>Anolis equestris</i>	Endemic	
<i>A. e. equestris</i>		WR
<i>A. e. buidei</i>		WR
<i>A. e. cincoleguas</i>		AS
<i>A. e. juraguensis</i>		CR
<i>A. e. persparsus</i>		CR
<i>A. e. potior</i>		AC
<i>A. e. thomasi</i>		CMR
<i>A. e. verreonensis</i>		ER
<i>Anolis fugitivus</i>	Endemic	ER
<i>Anolis garridoi</i>	Endemic	CR
<i>Anolis guafe</i>	Endemic	ER
<i>Anolis guamuhaya</i>	Endemic	CR
<i>Anolis guazuma</i>	Endemic	ER
<i>Anolis homolechis</i>	Endemic	
<i>A. h. homolechis</i>		IW, IJ
<i>A. h. turquinensis</i>		ER
<i>Anolis imias</i>	Endemic	ER
<i>Anolis inexpectatus</i>	Endemic	ER
<i>Anolis isolepis</i>	Endemic	
<i>A. i. isolepis</i>		CR-ER
<i>A. I. altitudinalis</i>		ER
<i>Anolis juangundlachi</i>	Endemic	WR
<i>Anolis jubar</i>	Endemic	
<i>A. j. jubar</i>		CMR
<i>A. j. albertschwartzi</i>		ER
<i>A. j. balaenarum</i>		AC
<i>A. j. cocoensis</i>		AC

Table 2.2 cont.

<i>A. j. cuneus</i>		CMR
<i>A. j. gibarensis</i>		CMR-ER
<i>A. j. maisensi</i>		ER
<i>A. j. oriens</i>		ER
<i>A. j. santamariae</i>		AS
<i>A. j. yaguajayensis</i>		CR
<i>Anolis loysianus</i>	Endemic	IW
<i>Anolis lucius</i>		IW
<i>Anolis luteogularis</i>	Endemic	
<i>A. l. luteogularis</i>		WR
<i>A. l. calceus</i>		WR
<i>A. l. coctilis</i>		ACA
<i>A. l. delacruzii</i>		IJ
<i>A. l. hassleri</i>		IJ
<i>A. l. jaumei</i>		WR
<i>A. l. nivevultus</i>		WR
<i>A. l. sanfelipensis</i>		CSF
<i>A. l. sectilis</i>		IJ
<i>Anolis macilentus</i>	Endemic	ER
<i>Anolis mestrei</i>	Endemic	ER
<i>Anolis mimus</i>	Endemic	ER
<i>Anolis noblei</i>	Endemic	
<i>A. n. noblei</i>		ER
<i>A. n. galeifer</i>		ER
<i>Anolis ophiolepis</i>	Endemic	IW
<i>Anolis paternus</i>	Endemic	
<i>A. p. paternus</i>		IJ
<i>A. p. pinarensis</i>		WR
<i>Anolis pigmaequestris</i>	Endemic	AS
<i>Anolis porcatus</i>	Endemic	IW, IJ
<i>Anolis porcus</i>	Endemic	IW
<i>Anolis pumilus</i>	Endemic	WR-CR-AS-AC
<i>Anolis quadriocellifer</i>	Endemic	WR
<i>Anolis rubribarbus</i>	Endemic	ER
<i>Anolis sagrei</i>	Cuba, Jamaica, Bahama Is., NA	
<i>A. s. sagrei</i>		IW
<i>A. s. greyi</i>		CMR
<i>Anolis smallwoodi</i>	Endemic	ER
<i>A. s. smallwoodi</i>		

Table 2.2 cont.

<i>A. s. palardis</i>		
<i>A. s. saxuliceps</i>		
<i>Anolis spectrum</i>	Endemic	WR
<i>Anolis vanidicus</i>	Endemic	
<i>A. v. vanidicus</i>		CR
<i>A. v. rejectus</i>		ER
<i>Anolis vermiculatus</i>	Endemic	WR
<i>Anolis vescus</i>	Endemic	ER
Family Teiidae		
<i>Ameiva auberi</i>	Cuba and Bahama Is.	
<i>A. a. auberi</i>	Endemic	WR
<i>A. a. abducta</i>	Endemic	WR
<i>A. a. atrothorax</i>	Endemic	CR
<i>A. a. cacuminus</i>	Endemic	WR
<i>A. a. citra</i>	Endemic	CMR
<i>A. a. denticola</i>	Endemic	WR
<i>A. a. extorris</i>	Endemic	ASC
<i>A. a. extraria</i>	Endemic	ASC
<i>A. a. galbiceps</i>	Endemic	AJR
<i>A. a. garridoi</i>	Endemic	CR
<i>A. a. gemmea</i>	Endemic	CR
<i>A. a. granti</i>	Endemic	ER
<i>A. a. hardyi</i>	Endemic	ER
<i>A. a. llanensis</i>	Endemic	CMR
<i>A. a. marcida</i>	Endemic	IJ
<i>A. a. nigriventris</i>	Endemic	IJ
<i>A. a. orlandoi</i>	Endemic	ASC
<i>A. a. paulsoni</i>	Endemic	WR
<i>A. a. peradusta</i>	Endemic	CR
<i>A. a. procer</i>	Endemic	WR
<i>A. a. pullata</i>	Endemic	WR
<i>A. a. sabulicolor</i>	Endemic	ER
<i>A. a. sanfelipensis</i>	Endemic	CSF
<i>A. a. schwartzi</i>	Endemic	WR
<i>A. a. secta</i>	Endemic	IJ
<i>A. a. sublesta</i>	Endemic	ASC
<i>A. a. ustulata</i>	Endemic	ER
<i>A. a. zugi</i>	Endemic	ACA, WR

Table 2.2 cont.

Family Tropiduridae		
<i>Leiocephalus carinatus</i>	Cuba, Cayman and Bahama Is.	
<i>L. c. carinatus</i>	Endemic	WR
<i>L. c. acuarius</i>	Endemic	ER
<i>L. c. cayensis</i>	Endemic	AJR
<i>L. c. labrossytus</i>	Endemic	WR-CR
<i>L. c. microcyon</i>	Endemic	IJ
<i>L. c. mogotensis</i>	Endemic	WR
<i>L. c. zayasi</i>	Endemic	WR
<i>Leiocephalus cubensis</i>	Endemic	
<i>L. c. cubensis</i>		IW
<i>L. c. gigas</i>		IJ
<i>L. c. minor</i>		CSF
<i>L. c. pambasileus</i>		ACA
<i>L. c. paraphrus</i>		AJR
<i>Leiocephalus macropus</i>	Endemic	
<i>L. m. macropus</i>		ER
<i>L. m. aegialis</i>		CMR
<i>L. m. asbolomus</i>		ER
<i>L. m. felinoi</i>		WR
<i>L. m. hoplites</i>		CR-CMR
<i>L. m. hyacinthurus</i>		CR
<i>L. m. immaculatus</i>		ER
<i>L. m. koopmani</i>		WR
<i>L. m. lenticulatus</i>		CMR
<i>L. m. phylax</i>		ER
<i>L. m. torrei</i>		WR
<i>Leiocephalus onaneyi</i>	Endemic	ER
<i>Leiocephalus raviceps</i>	Endemic	
<i>L. r. raviceps</i>		ER
<i>L. r. delavarai</i>		ER
<i>L. r. jaumei</i>		WR
<i>L. r. klinikowskii</i>		WR
<i>L. r. uzzelli</i>		ER
<i>Leiocephalus stictigaster</i>	Endemic	
<i>L. s. stictigaster</i>		WR
<i>L. s. astictus</i>		IJ
<i>L. s. celeustes</i>		ER
<i>L. s. exothetus</i>		IJ

Table 2.2 cont.

<i>L. s. gibarensis</i>		CMR-ER
<i>L. s. lipomator</i>		CR
<i>L. s. Lucianus</i>		CMR
<i>L. s. naranjoi</i>		CR
<i>L. s. ophioplacodes</i>		CMR
<i>L. s. parasphex</i>		AC
<i>L. s. septentrionalis</i>		ASC
<i>L. s. sierrae</i>		WR
Family Xantusiidae		
<i>Cricosaura typica</i>	Endemic	ER
ORDEN SERPENTES		
Family Boidae		
<i>Epicrates angulifer</i>	Endemic	IW
Family Colubridae		
<i>Alsophis cantherigerus</i>	Cuba and Cayman Is.	
<i>A. c. cantherigerus</i>	Endemic	WR-CR, IJ, ACA
<i>A. c. adspersus</i>	Endemic	ER
<i>A. c. pepeii</i>	Endemic	ER
<i>A. c. schwartzi</i>	Endemic	CR, CMR, ER AC
<i>Antillophis andreaei</i>	Endemic	
<i>A. a. andreaei</i>		WR-CMR
<i>A. a. melopyrrha</i>		ACA
<i>A. a. morenoi</i>		ASC
<i>A. a. nebulatus</i>		IJ
<i>A. a. orientalis</i>		CMR-ER
<i>A. a. peninsulae</i>		WR
<i>Arrhyton ainictum</i>	Endemic	ER
<i>Arrhyton dolichura</i>	Endemic	WR
<i>Arrhyton landoi</i>	Endemic	ER
<i>Arrhyton procerum</i>	Endemic	WR
<i>Arrhyton supernum</i>	Endemic	ER
<i>Arrhyton taeniatum</i>	Endemic	WR, CR, ER, IJ
<i>Arrhyton tanyplectum</i>	Endemic	WR
<i>Arrhyton vittatum</i>	Endemic	IW, IJ
<i>Nerodia clarki</i>	Cuba	
<i>N. c. compressicauda</i>	Gulf Coast, Cuba and Florida Keys	WR-CR
<i>Tretanorhinus variabilis</i>	Cuba and Cayman Is.	

Table 2.2 cont.

<i>T. v. variabilis</i>	Endemic	WR, CR, ER
<i>T. v. binghami</i>	Endemic	ER
<i>T. v. insulaepinorum</i>	Endemic	IJ
<i>T. v. wagleri</i>	Endemic	WR
Family Tropidophidae		
<i>Tropidophis feicki</i>	Endemic	WR
<i>Tropidophis fuscus</i>	Endemic	ER
<i>Tropidophis haetianus</i>	Cuba and Haiti	
<i>T. h. haetianus</i>		ER
<i>Tropidophis maculatus</i>	Endemic	WR
<i>Tropidophis melanurus</i>	Cuba and Cayman Is.	
<i>T. m. melanurus</i>	Endemic	IW
<i>T. m. dysodes</i>	Endemic	WR
<i>T. m. ericksoni</i>	Endemic	IJ
<i>Tropidophis nigriventris</i>	Endemic	
<i>T. n. nigriventris</i>		CMR
<i>T. n. hardyi</i>		CR
<i>Tropidophis pardalis</i>	Endemic	IW, IJ
<i>Tropidophis pilsbryi</i>	Endemic	
<i>T. p. pilsbryi</i>		ER
<i>T. p. galacelidus</i>		CR
<i>Tropidophis semicinctus</i>	Endemic	WR
<i>Tropidophis wrighti</i>	Endemic	CR-ER
Family Typhlopidae		
<i>Typhlops biminiensis</i>	Cuba and Bahama Is.	
<i>T. b. biminiensis</i>		WR, CR, ER
<i>Typhlops lumbricalis</i>	Cuba and Bahama Is.	IW, IJ
ORDER CROCODYLIA		
Family Alligatoridae		
<i>Caiman crocodilus</i>	Introduced	IJ
Family Crocodylidae		
<i>Crocodylus acutus</i>		IW, IJ
<i>Crocodylus rhombifer</i>	Endemic	WR

*The letters in the Distribution columns refer to the following geographic regions; WR, Western region; CR, Central region; CMR, Camagüey-Maniabón region; ER, Eastern region; AS, Archipiélago de Sabana; AC, Archipiélago de Camagüey; ASC, Archipiélago de Sabana-Camagüey; CSF, Cayos de San Felipe; ACA, Archipiélago de los Canarreos; AJR, Archipiélago de Jardines de la Reina; IW, Islandwide; IJ, Isla de la Juventud (Mataeo and Acevedo, 1989).

There are two subspecies: *T. d. decussata*, extending from the east end to the western end of Cuba, including the keys of Sabana-Camagüey, and *T. d. angusta* is found in the extreme east end of Cuba, on the Isla de la Juventud, and also on the Cayman Islands (Seidel, 1988a,b, 1990).

Of the remaining 121 reptiles, 93 are saurians. There are three species of *Amphisbaena*. *Amphisbaena cubana* has two subspecies: *A. c. cubana* found from the Ciénaga de Zapata to the Eastern region and Isla de la Juventud and *A. c. barbouri* distributed in the Western region (excluding the western Ciénaga de Zapata). *Amphisbaena blainoides* is common in the Cordillera de Guaniguanico and *A. palirostrata* is only known from the northern portion of the Isla de la Juventud. These last two species were previously included in the genus *Cadea*, however, Hedges (1996b) considered *Cadea* synonymous with *Amphisbaena*.

Three species of anguids are present and all belong to the genus *Diploglossus*. The distribution of *D. delasagra* includes the Western and Central regions, the Alturas del Centro de Camagüey, and the southern part of the Eastern region. *Diploglossus nigropunctatus* is found in the northern part of the Camagüey-Maniabón region and the mountains of Sagua-Baracoa. The third species, *D. garridoi* is from the northern slopes of the Sierra Maestra (Hedges and Thomas, 1998).

The Gekkonidae is represented by four genera: *Gonatodes*, *Hemidactylus*, *Sphaerodactylus*, and *Tarentola*. There is a single species of *Gonatodes*: *G. albogularis fuscus* which is found in some urban areas in Cuba and is a species widely distributed in Central and South America. *Hemidactylus brooki haitianus* is found throughout Cuba, including the keys of Sabana-Camagüey, and also occurs in Hispaniola and Puerto Rico (Schwartz and Henderson, 1991). Another member of this genus, *H. mabouia*, is known from Guantánamo and the city of La Habana, and is also found in Hispaniola, Bahamas, Virgin Islands, Puerto Rico, Mona, Culebra, and Vieques (Schwartz and Henderson, 1991). The third species, *H. turcicus turcicus*, presumably introduced into Cuba, is found in the cities of La Habana, Matanzas, Santa Clara, Cienfuegos, Santiago de Cuba, and Guantánamo.

The genus *Sphaerodactylus* is represented by 18 species, but 4 of these are not endemic to Cuba. The Cuban *Sphaerodactylus* have been classified into seven morphological groups (Thomas *et al.*, 1992; Hedges and Garrido, 1993): (1) the *nigropunctatus* complex contains *S. nigropunctatus* and *S. torrei*. Both are large species with granular scales and a pattern of bands and are sexually dimorphic in color; (2)

S. elegans is similar to the prior complex, but does not have color differences between the sexes; (3) the *intermedius* complex has four species, *S. intermedius*, *S. armasi*, *S. docimus*, and *S. ruibali*, all characterized by imbricate scales and a less prominent pattern of bands; (4) the *scaber* complex has four species, *S. scaber*, *S. oliveri*, *S. richardi*, and *S. storeyae*, all are medium sized, with bulky dorsal scales and a middorsal zone of small granular scales, and all are sexually dichromatic; (5) the *notatus* complex has three species, *S. notatus*, *S. bromeliarum*, and *S. celicara*, are all medium-size lizards with flat imbricate dorsal scales (there are various species of this complex in Hispaniola); (6) the *ramsdeni* complex has three species, *S. ramsdeni*, *S. cricoderus*, and *S. schwartzi*, and all have dark dorsal scales that do not overlap and more than one internasal scale; and (7) *S. argus* is a species with small imbricate dorsals and an ocellated color pattern.

Sphaerodactylus argus argus is known from south coastal localities of the Central, Camagüey-Maniabón, and Eastern regions, including the keys of the Jardines de la Reina. This subspecies is also found in the northern Bahamas and is broadly distributed on Jamaica. *Sphaerodactylus a. andresensis* is found on the island of San Andrés in the Gulf of Honduras (Schwartz and Henderson, 1991). *Sphaerodactylus elegans elegans* has a wide distribution on Cuba, the keys of Sabana-Camagüey, Canarreos, and Jardines de la Reina, as well as on Isla de la Juventud. There are also presumably introduced populations on the Florida Keys (Schwartz and Henderson, 1991). Another subspecies, *S. e. punctatissimus* inhabits the western part of Hispaniola and the island of Gonave. *Sphaerodactylus nigropunctatus* is a polytypic species with at least five subspecies in Cuba and another five in the Bahamas. The Cuban subspecies are: *S. n. alayoi* is found on the edge of the Guaso plateau and the eastern slopes of the Cuenca de Guantánamo; *S. n. granti* is found in the Sabana-Camagüey keys, the Alturas de Maniabón-Banes, the mountains of Nipe-Cristal, and the northeast of the Cauto Valley; *S. n. lissodesmus* is known only from the Sierra de Cubitas in Camagüey; *S. n. ocujal* is found on the southern slopes of the Sierra Maestra and the Meseta de Cabo Cruz; and *S. n. strategus* is found on the coastal slopes of the Guantánamo valley (Schwartz and Henderson, 1991). Of the four subspecies recognized for *S. notatus*, only *S. n. atactus* is found at isolated localities in the Western, Central, Camagüey-Maniabón, and Eastern regions. It is most common in the Eastern region. The populations in the San Felipe keys, Isla de la Juventud, Canarreos keys, and various localities in the Eastern region show marked differences from the *atactus* morphology. *Sphaerodactylus intermedius* is distributed across the Alturas

de La Habana-Matanzas; *S. richardi* is found along the length of the coastal forests between the bays of Cochinos and Cienfuegos (Hedges and Garrido, 1993). *Sphaerodactylus oliveri* is located on the spurs of the Sierra de Trinidad. *Sphaerodactylus storeyae* was described by Grant (1944a) but Schwartz (1961), Schwartz and Garrido (1981a), and Schwartz and Henderson (1991) treated it as a subspecies of *S. oliveri*. Hedges and Garrido (1993) proposed considering it a separate species. It is restricted to the Isla de la Juventud and the Canarreos keys. *Sphaerodactylus scaber* is distributed through the Llanura Corralillo-Yaguajay and the Llanura Sur de Camagüey. The rest of the species are found in the Eastern region: *S. armasi*, *S. bromeliarum*, and *S. celicara* in the mountains of Sagua-Baracoa; *S. cricoderus* on the north and south slope of the Sierra Maestra; and *S. docimus* on the South slopes of the meseta de Cabo Cruz and the eastern end of the Sierra Maestra (Schwartz and Henderson, 1991; Thomas *et al.*, 1992). *Sphaerodactylus schwartzi* and *S. ruibali* are distributed to the west and east of the Bahía de Guantánamo. *Sphaerodactylus ramsdeni* has two populations, one in the Sierra de Gran Piedra and another more than 40 km to the northeast at the Meseta de Guaso (Schwartz and Garrido, 1985; Thomas *et al.*, 1992). *Sphaerodactylus torrei* has two subspecies: *S. t. torrei* in the vicinity of Santiago de Cuba and *S. t. spielmani* on the west of Guantánamo bay (Schwartz and Garrido, 1985).

Cyclura nubila has a wide distribution throughout Cuba along the coastal regions and on the keys, including the Isla de la Juventud. The populations of *Cyclura* have decreased due to development and construction along the coast. There are some interior populations in the Sierra de los Organos and the Alturas de La Habana-Matanzas. Three subspecies are recognized: *C. n. nubila* on Cuba and *C. n. caymanensis* and *C. n. lewisi* on the Cayman Islands (Schwartz and Carey, 1977; Schwartz and Henderson, 1991).

The Polychrotidae is the most numerous and best represented family in Cuba with 55 anoline species, including the 4 species of *Chamaeleolis* included in the synonymy of *Anolis* by Hass *et al.* (1993). There are four patterns of distribution: Species that are exclusively western, central or eastern and species that are broadly distributed throughout the island. The broadly distributed species are: *Anolis alutaceus*, *A. allisoni*, *A. angusticeps*, *A. chamaeleonides*, *A. equestris*, *A. homolechis*, *A. loysiana*, *A. lucius*, *A. ophiolepis*, *A. porcatus* and *A. sagrei* (Schwartz and Henderson, 1991). The distribution of *A. allisoni* extends from the eastern edge of the Western region to the rest of the island. Outside of Cuba it is known from populations

on the Islas de la Bahía off the north coast of Honduras and Half Moon Cay off the coast of Belize (Ruibal and Williams, 1961). *Anolis allogus* is not present in the eastern part of the Western region and is absent in the Central region (Schwartz and Henderson, 1991). *Anolis angusticeps* is represented in Cuba by the nominative subspecies *A. a. angusticeps*, whereas *A. a. ologapsi* is restricted to the Bahamas (Garrido, 1975d, Hardy, 1967). Eight subspecies have been described for *A. equestris*, but some populations in the Cordillera de Guaniguanico in western Cuba have not been identified subspecifically (Schwartz and Garrido, 1972; Garrido, 1981; Schwartz and Henderson, 1991). Garrido (1973a) described *A. homolechis turquinensis* as an altitudinal subspecies from the Pico Turquino. With respect to *A. lucius*, populations have been reported in the Cordillera de Guaniguanico, but they require confirmation. *Anolis lucius* has also been accidentally introduced to Cayo Largo del Sur (Garrido and Jaume, 1984). *Anolis sagrei* is the most common of the Cuban anoles, is found throughout the island and keys, and has a single subspecies, *A. s. greyi*, from the Camagüey-Maniabón region (Schwartz and Henderson, 1991).

Eleven species have a distribution in the Western region: *Anolis barbatus* is in the Sierra del Rosario (Garrido, 1982a), and *A. mestrei* and *A. bartschi* are in the Cordillera de Guaniguanico (Schwartz and Henderson, 1991). *Anolis bremeri* has two subspecies, *A. b. bremeri* in the Llanura Sur of Pinar del Río and *A. b. insulaepinarum* on the Isla de la Juventud (Garrido, 1972). *Anolis juangundlachi* is known only from the vicinity of Carlos Rojas in the Llanura Artemisa-Colón (Garrido, 1975c). *Anolis luteocularis* has nine subspecies in the region, including the Isla de la Juventud and the Archipelago de los Canarreos (Schwartz and Garrido, 1972; Schwartz and Henderson, 1991). *Anolis paternus* has two subspecies, one on the Isla de la Juventud (*A. p. paternus*) and the second, *A. p. pinarensis*, from the Llanura Sur of Pinar del Río (Garrido, 1975a). The distribution of *A. pumilus* extends into some localities in the Central region and to the keys of Sabana and Camagüey on the north and also the Isla de la Juventud (Garrido, 1988). *Anolis quardiocellifer* is restricted to the Llanura de Guanahacabibes (Garrido, 1973b). *Anolis spectrum* is known from various localities in the Sierra de los Organos, a single locality near La Habana, and also in the vicinity of Carlos Rojas in the eastern part of the Llanura Artemisa-Colón (Garrido and Schwartz, 1972). *Anolis vermiculatus* is found in the vegetation along the banks of rivers and streams that arise in the Cordillera de Guaniguanico (Schwartz and Henderson, 1991).

The species restricted to the Central region are all endemics of the Sierra de Guamuhaya: *Anolis ahli*, *A. delafuentei*, *A. garridoi*, and *A. guamuhaya* (Garrido *et al.*, 1991; Diaz *et al.*, 1996).

Most species of Cuban anoles have an eastern distribution: *Anolis alayoni*, *A. alfaroi*, *A. anfiloquioi*, *A. argenteolus*, *A. argillaceus*, *A. baracoae*, *A. birama*, *A. clivicola*, *A. cupeyalensis*, *A. fugitivus*, *A. guafe*, *A. guazuma*, *A. imias*, *A. inexpectatus*, *A. macilentus*, *A. mimius*, *A. noblei*, *A. porcus*, *A. rubribarbus*, *A. smallwoodi*, and *A. vescus*. Some of these species stand out because of their restricted distribution. For example, *A. alfaroi* inhabits the grasses and ferns below the pines of La Munición in the Cuchillas de Toa, in the mountains of Sagua-Baracoa (Garrido and Hedges, 1992). *Anolis birama* is restricted to the environs of the Ciénaga de Birama to the northwest of the mouth of the Río Cauto (Garrido, 1990). *Anolis clivicola* is found in the Sierra Maestra above 1000 m in elevation (Schwartz and Henderson, 1991). *Anolis fugitivus* is known from the northeastern slopes of Monte Iberia in the mountains of Sagua-Baracoa (Garrido, 1975c). Garrido (1983) described *A. guazuma* from Pico Turquino and the species is found on the north and south slopes of the Sierra Maestra. *Anolis macilentus* is known only from the vicinity of the Río Pai in the Maseta de Guaso (Garrido and Hedges, 1992). The extreme eastern end of the Sierra Maestra is the locality for *A. mimus* (Garrido, 1975c; Schwartz and Thomas, 1975), whereas *A. vescus* has a similar narrow distribution in the vicinity of Los Calderos in the Sierra de Imías (Garrido and Hedges, 1992). In the Meseta of Cabo Cruz there are two species found exclusively in the dry forests of the zone: *A. guafe* and *A. confusus* (Estrada and Garrido, 1991). Some of the species with an eastern distribution contain various subspecies in different subregions. For example, in the case of *A. cyanopleurus*, one of the subspecies (*A. c. cyanopleurus*) is found in the northwestern part of the Sagua-Baracoa subregion while the other subspecies (*A. c. orientalis*) is restricted to the extreme southeastern portion of the subregion in the Meseta de Maisí (Garrido, 1975c). *Anolis noblei* has two subspecies, *A. n. noblei* in the Sierra Maestra and *A. n. galeifer* in the Sierra Nipe-Cristal (Schwartz and Garrido, 1972).

Other species with patterns of distribution different from those mentioned are *A. centralis*, *A. isolepis*, and *A. jubar*. In the case of *A. centralis* it has a subspecies in the Camagüey-Maniabón region, *A. c. centralis*, and another, *A. c. litoralis* ranging from the southern part of the Sierra Maestra, Cuenca de Santiago de Cuba, Valle Central, Cuenca de Guantánamo, to the southern part of Sagua-Baracoa (Garrido, 1975b). *Anolis isolepis* has two subspecies: *A. i. isolepis* is found

in the Guamuhaya mountains and various localities in the Eastern region, and no differences have been documented distinguishing these separate populations. *Anolis i. altitudinalis* is on Pico Turquino in the Sierra Maestra (Garrido, 1985). Of the nine subspecies of *Anolis jubar*, one (*A. j. yaguajayensis*) is from the Central region in the Llanura Corralillo-Yaguajay; two subspecies are found in the keys off the north coast, *A. j. cocoensis* on the Coco, Paredón Grande and Romano keys, and *A. j. santamariae* on Cayo Santa María; the nominative subspecies, *A. j. jubar*, is found in part of the Llanura Norte de Camagüey; *A. j. balaenarum* is found on the Los Ballenatos keys in the Bahía de Nuevitas; *A. j. cuneus* is in the eastern half of the Llanura Norte of Camagüey; and in the same region *A. j. gibarense* is found in the Alturas de Maniobón and Gibara extending to the east of Bahía de Nipe (Schwartz and Henderson, 1991; Estrada and Garrido, 1990). The remaining three subspecies are distributed along the coastal zone from Cabo Cruz to the Punta de Maisí; *A. j. oriens* from Cabo Cruz to Santiago de Cuba, *A. j. albertschwartzi* from the Cuenca de Guantánamo to near the Meseta de Maisí, and *A. j. maisensis* on the terraces of Punta de Maisí, *Anolis pigmaequestrus* is only known from the Francés and Santa María keys on the north coast (Garrido, 1975a).

The family Tropiduridae has 6 species and, with the exception of *Leiocephalus carinatus*, all are endemic to Cuba. *Leiocephalus carinatus* has 13 subspecies, 6 of which are not Cuban but are found in the Bahamas, Cayman Islands and Swan Island. The seven Cuban subspecies are: *L. c. carinatus* along the north coast in the subregion of Alturas de La Habana-Matanzas; *L. c. aquarius* along the entire south coast of the Eastern region, from Cabo Cruz to Maisí; *L. c. cayensis* is in the keys of the Jardines de la Reina; *L. c. labrossytus* is located near Bahía de Cochinos along the coast and the south slopes of the Sierra de Trinidad; *L. c. microcyon* on the Isla de la Juventud; *L. c. mogotensis* in the Sierra de los Organos; and *L. c. zayasi* in the Llanura de Guanahacabibes (Schwartz and Henderson, 1991). Other populations, not identified subspecifically, are known from Cayo Coco, Villa Clara, Nuevitas, Holguín, Inglés and Rosario keys in the Archipiélago de los Canarreos, and the Cayos Blancos south of the Zapata Peninsula (Estrada, 1992, 1993a,b).

Leiocephalus cubensis has 5 described subspecies: *L. c. cubensis* is widely distributed on Cuba, though the records from Guanahacabibes and the Cordillera Guaniguanico are questionable (Schwartz and Henderson, 1991); *L. c. gigas* is exclusively on the Isla de la Juventud; *L. c. minor* is only found on the San Felipe keys to the northwest of

Isla de la Juventud; *L. c. pambasilous* is found on the Hicacos and Campo keys at the extreme western end of the Archipiélago de los Canarreos; and *L. c. paraphrus* is restricted to the keys of the Jardines de la Reina. One population not assigned subspecies status is on Cayo Coco on the north coast in the Sabana-Camagüey keys (Estrada, 1993b). The third species of *Leiocephalus* is *L. macropus*, which has 11 subspecies, of which six are restricted to the subregions of the Eastern region: *L. m. macropus*, *L. m. asbolomus*, *L. m. immaculatus*, *L. m. lenticulatus*, and *L. m. phylax*. One subspecies, *L. m. aegialus*, is restricted to Playa Santa Lucía on the north coast of Camagüey. Other subspecies with narrow local distributions are: *L. m. hoplites* in the northern part of the Llanura Júcaro-Morón, *L. m. hyacinthurus* in the mountains of Guamuhaya, *L. m. felinoi* in the Alturas de La Habana-Matanzas, *L. m. koopmani* in the Llanura de Guanahacabibes, and *L. m. torrei* from San Miguel de los Baños in the Alturas Bejucal-Coliseo (Schwartz and Henderson, 1991). There are also other populations not identified subspecifically in the Sierra de Rosario, Isla de la Juventud, Archipiélago de Sabana, and others in the Eastern region (Schwartz and Henderson, 1991).

Leiocephalus onaneyi is a monotypic species from the summit of the Loma de Macambo in a coastal dry forest to the east of the Cuenca de Guantánamo (Garrido, 1973a). *Leiocephalus raviceps* has five subspecies *L. r. raviceps* along the coast of the Cuenca de Guantánamo; *L. r. delavarai* in the vicinity of Gibara in the Alturas de Mamiabón; *L. r. jaumei* on the western end of the Llanura Sur de Pinar del Río; *L. r. klinikowskii* on the Hicacos Peninsula, and *L. r. uzzelli* along the coastal area between Santiago de Cuba and Guantánamo (Schwartz and Henderson, 1991).

Leiocephalus stictigaster is one of the most diverse species of the Cuban tropidurids with twelve described subspecies. Some of the subspecies are restricted to dry coastal areas: *L. s. stictigaster* on the Llanura de Guanahacabibes, *L. s. astictus* on the southern coast of Isla de la Juventud, *L. s. naranjoi* on the coastal southern slopes of the Sierra de Guamuhaya, *L. s. parasphex* and *L. s. septentrionalis* on the Archipiélago de Sabana-Camagüey, *L. s. lucianus* in the Playa Santa Lucía on the north coast of Camagüey, and *L. s. gibarensis* of the coastal zone of the Alturas de Maniabón (Schwartz and Henderson, 1991). Other subspecies are located in interior areas such as *L. s. sier-rae* in the Alturas de Pizarras surrounding the Sierra de los Organos, *L. s. lipomator* in the Alturas de Cubanacán near Santa Clara, *L. s. ophiplacodes* in the western part of the Llanura Norte de Camagüey, *L. s. exothetus* in the northern area of Isla de la Juventud, and *L. s.*

celeustes on the border between the Llanura de Cauto and the northern slopes of the Sierra Maestra (Schwartz and Henderson, 1991). There are also other populations of this species without subspecific determination along the southeast coast of the Península de Zapata, on the Cayos Blancos south of the peninsula, and in the Sierra de Gran Piedra (Schwartz and Henderson, 1991).

The family Teiidae has only one species in Cuba, *Ameiva auberi*, and it is probably one of the most diverse species of the genus, judging by the fact that 40 subspecies have been described, of which 28 are restricted to Cuba (Schwartz and Henderson, 1991). The distribution of these subspecies covers the entire island and the surrounding keys, ranging from coastal zone to mountains and in different types of forests.

The only member of the Xantusiidae in the Caribbean in *Cricosaura typica*, a species that for many years was only known from a few localities in the plateau near Cabo Cruz in eastern Cuba. Recently the distribution has been expanded and the species is known from populations in the dry forest of the Meseta de Cabo Cruz and the coastal zone along the south slope of the Sierra Maestra extending to the vicinity of Uvero (Estrada and de Armas, 1998). The origin of this species has been the subject of considerable debate, which is ongoing (Crother *et al.*, 1986; Crother and Guyer, 1996; Crother and Presch, 1992, 1994; Hedges *et al.*, 1991; Hedges and Bezy, 1993, 1994; Hedges, 1996b).

The 25 Cuban species of the Serpentes include the families Boidae, Colubridae, Tropidophidae, and Typhlopidae. The Boidae contains an endemic species, *Epicrates angulifer*, known throughout Cuba and the keys of Sabana-Camagüey, Canarreos, Cayos Blanco del Sur, and Isla de la Juventud. The colubrids are represented by 10 species and the genera *Alsophis*, *Antillophis*, *Arrhyton*, *Nerodia*, and *Tretanorhinus*.

The only species of *Alsophis* is *A. cantherigerus* with eight subspecies, of which three are restricted to the Caiman Islands and another to Swan Island (Schwartz and Henderson, 1991). Of the five Cuban endemic subspecies the one with the most extensive distribution is *A. c. cantherigerus* found in all of the Western region including the Canarreos keys and Isla de la Juventud, and extends to the northwestern part of the Central region including the Sabana keys (Schwartz and Henderson, 1991). *Alsophis c. schwartzi* is found in the rest of the Central region, the Camagüey-Maniobón region, including the Archipiélago de Camagüey, the southwestern portion of the Llanura del Cauto, the Meseta de Cabo Cruz, Sierra Maestra, Valle Central, the ba-

sins of Santiago de Cuba and Guantánamo, and part of the Meseta de Guaso (Schwartz and Henderson, 1991). The subspecies *A. c. adspersus* and *A. c. pepei* extend along the northeast and southeast of the Eastern region. The populations in the extreme eastern end of Cuba are considered intermediates between the *adspersus* and *pepei* subspecies, and those in the southwestern Sierra Maestra are considered intermediates between *pepei* and *schwartzi* (Schwartz and Henderson, 1991).

Antillophis contains one species *A. andreaei* with six subspecies. *Antillophis a. andreaei* is found throughout the west (except for the Llanura de Guanahacabibes) to the western half of the Central region; *A. a. orientalis* is distributed from the eastern half of Camagüey-Maniabón to the entire Eastern region. There are at least three subspecies with localized distribution: *A. a. peninsulae* in Guanahacabibes, *A. a. nebulatus* on the Isla de la Juventud, *A. a. melopyrrha* on Cantiles key in the Canarreos and *A. a. morenoi* on the keys of Sabana-Camagüey. The populations between the eastern part of the Central region and the western half of Camagüey-Maniabón show intergradation between *andreaei* and *orientalis* (Schwartz and Henderson, 1991).

The genus *Arrhyton* has six species in Cuba, *A. ainictum* is known only from the type locality at Cueva del 18 in the extreme east of the Llanura Sur of Camagüey; *A. dolichura* is another species with a poorly documented distribution and is known only from the vicinity of the city of La Habana. *Arrhyton landoi* is restricted to the southern part of the Eastern region. Two species, *A. taeniatum* and *A. vittatum*, have been reported from different localities of the Western, Central, and Eastern regions and Isla de la Juventud (Schwartz and Henderson, 1991). Another species *A. tanyplectum*, is known from a few localities in the Sierra de los Organos. Recently two new species have been described, *A. supernum* in the mountains of Sagua-Baracoa in eastern Cuba, and *A. procerum* from the vicinity of Playa Girón in the Llanura de Zapata (Hedges and Garrido, 1992a).

The two remaining species of colubrids, include *Nerodia clarki compressicauda*, a water snake of swamps, estuaries and bays of marine or brackish waters, and found at various coastal sites in the Western and Central regions and the keys of Sabana-Camagüey. *Tretanorhinus variabilis*, in contrast, has five subspecies, four endemic to Cuba and the fifth to the Cayman Islands. *Tretanorhinus v. variabilis* is present throughout most of Cuba, except for the Isla de la Juventud where *T. v. insulaepinorum* is found, and in the western half of the Western region where *T. v. wagleri* is present, and the Cabo Cruz plateau in the western Sierra Maestra where *T. v. binghami* is resident.

The populations to the west of La Habana and those from the Canarreos keys have not been identified subspecifically (Schwartz and Henderson, 1991).

The family Tropidophidae is well represented on Cuba with 11 species of *Tropidophis*, of which 9 are endemic. *Tropidophis melanurus* has four subspecies of which only one is found outside of Cuba on Navassa Island. *Tropidophis m. melanurus* is found throughout Cuba, while *T. m. dysodes* is known from a small area in the center of the Llanura Sur de Pinar del Río and *T. m. ericksoni* from the Isla de la Juventud. This species has populations on the Canarreos, Sabana-Camagüey, and San Felipe keys that have no subspecific designation (Schwartz and Henderson, 1991).

Three of the *Tropidophis* species have a western distribution: *T. feicki*, *T. maculatus* and *T. semicinctus*. Another species, *T. nigriventris*, has two disjunct populations, considered subspecies: *T. n. nigriventris* from the Alturas de Camagüey and *T. n. hardyi* in the southern part of the Sierra de Guamuha. *Tropidophis pilsbryi* is another species with disjunct populations; *T. p. pilsbryi* in the Eastern region and *T. p. galacelidus* in the Sierra de Guamuha (Schwartz and Henderson, 1991). *Tropidophis pardalis* is a common species in western Cuba, but isolated populations are found throughout the main island and Isla de la Juventud and the keys of Sabana-Camagüey. *Tropidophis wrighti* has a distribution from the Central to the Eastern region. *Tropidophis haetianus haetianus* is not endemic to Cuba and has been reported from the northern part of the Eastern region (Schwartz and Henderson, 1991). *Tropidophis fuscus* is known from two localities in the Sagua-Baracoa mountains (Hedges and Garrido, 1992b).

The Family Typhlopidae has two species of *Typhlops* in Cuba: *T. biminiensis*, a Bahaman species that has been reported from some localities in the Western, Central and Eastern regions of Cuba and *T. lumbricalis* widely distributed on Cuba, Isla de la Juventud, and in the southern and northern Bahamas (Schwartz and Henderson, 1991).

The order Crocodylia has three species on Cuba, one an introduced member of the Alligatoridae, and two native species of Crocodylidae: *Caiman crocodylus* was introduced in 1959 in the Ciénaga de Lanier, on Isla de la Juventud, and today the species is found throughout the wet areas of the island (Varona, 1976; Schwartz and Henderson, 1991); *Crocodylus acutus* is widely distributed in all the coastal and swampy areas including the keys and *C. rhombifer*, an endemic species, today found only in the western part of the Ciénaga de Zapata.

Jamaica

Ronald I. Crombie

Division of Amphibians and Reptiles, National Museum of Natural History, Smithsonian Institution, Washington, D.C. 20560

Introduction

Jamaica is the second smallest of the major Greater Antillean islands, a bit more than 11,000 km² in area, with a maximum elevation of 2256 m. It lies on its own bank, separated by very deep water from both the mainland and the other Antilles, with few, mostly close in-shore surrounding islets (Steers 1940a,b; Steers *et al.*, 1940; Crombie *et al.*, 1984). Consequently it could be considered the smallest major island because the Greater Puerto Rican Bank is slightly larger. Despite its modest size, Jamaica has a diverse (65+ species) and distinctive herpetofauna (Table 3.1). The fauna in general is striking as much by the elements that are absent as by the significant radiations of the taxa that are present, reflecting the island's isolation from its Antillean neighbors and (perhaps) by its geological history relative to the Middle American mainland (Buskirk, 1985; Hedges, 1989a,b). The list of Jamaican species is by no means complete, however, and the rugged limestone terrain continues to hide surprises and make a mockery of "definitive" statements on the rarity or even extinction of some species.

Because this book is a well-deserved tribute to Albert Schwartz, it is interesting to note that Jamaica was not one of his favorite places in the West Indies. For example, compared to the multitude of his papers on most of the rest of the Antilles, Schwartz published only two (Schwartz, 1971a; Schwartz and Fowler, 1973) exclusively on Jamaica and one other (Thomas and Schwartz, 1974) partly on a Jamaican species. Shortly after my first trip to the island in 1970 I visited with Schwartz in Miami, where he encouraged my interest in the Jamaican fauna, candidly admitting that he did not relish working there much more but recognizing that extensive investigations were needed.

Over a beverage or two, we discussed what seemed to me a peculiar prejudice, because I found the place utterly entrancing. As usual, Schwartz had a perfectly pragmatic and understandable basis for his mild aversion to Jamaica. His field trips were strongly automobile oriented and at that time Jamaica had a road system that was generally passable between the major population centers and the tourist resorts of the north coast. The small rural roads that provided access to the "good places" on many other islands were relatively uncommon on Jamaica and important collecting sites could often only be reached by hard overland trek through unforgiving terrain, decidedly not Schwartz's style at that point of his career. He also objected to the "touristy" aspect of Jamaica, which did not require many out-of-the-way pensiones or small eateries. The "tourist prices" for everything may have struck Schwartz, ever thrifty with field money, the hardest and he hated to pay full price for anything. However, Jamaicans rarely bargained, extrapolating the frequent tourist dim-wittedness to all foreigners. Schwartz believed that even the people well out of the tourist mainstream lacked the warmth and friendliness he valued so much on other islands. I did not entirely agree and being young and stupid (a trait some would deny I ever outgrew), I continued my Jamaican field work for the next fifteen years.

As I gained more experience on other Antillean islands, I could understand Schwartz's points far more clearly, and Jamaica was indeed never an "easy" place to work. The proud, aloof, almost arrogant Maroons never totally warmed to my presence in their forests but we shared many trials together and their unparalleled bush knowledge was enormously helpful to my projects. The activities of the U.S. Drug Enforcement Agency in attempting to stem the flow of drugs from Jamaica to the United States often made innocent foreigners suspect and unwelcome in isolated areas. However, the kindness, generosity, and inherent charm of the Jamaican people more than overwhelmed any negative encounters. The culture, the fauna, and all things Jamaican are the product of an environment that can oscillate from idyllic to fierce and hostile, seemingly without effort. The result is a unique kaleidoscopic effect that can seem far more foreign than Jamaica's proximity to the rest of the Antilles would warrant.

Table 3.1. Checklist of the Jamaican Herpetofauna

Anura:Bufonidae

Bufo marinus (Linnaeus, 1758) — Introduced

Anura: Hylidae

“Calyptahyla” crucialis (Harlan, 1826)*Hyla marianae* Dunn, 1926*Hyla wilderi* Dunn, 1925*Osteopilus brunneus* (Gosse, 1851)*Osteopilus* sp. nov.

Anura:Leptodactylidae

Eleutherodactylus alticola Lynn 1937*Eleutherodactylus andrewsi* Lynn 1937*Eleutherodactylus cavernicola* Lynn, 1954*Eleutherodactylus cundalli* Dunn, 1926*Eleutherodactylus fuscus* Lynn and Dent, 1943*Eleutherodactylus glaucoreius* Schwartz and Fowler, 1973*Eleutherodactylus g. gossei* Dunn, 1926*Eleutherodactylus gossei oligaulax* Schwartz and Fowler, 1973*Eleutherodactylus grabhami* Dunn, 1926*Eleutherodactylus griphus* Crombie, 1986*Eleutherodactylus jamaicensis* Barbour, 1910*Eleutherodactylus johnstonei* Barbour, 1914 – Introduced*Eleutherodactylus junori* Dunn, 1926*Eleutherodactylus luteolus* (Gosse, 1851)*Eleutherodactylus nubicola* Dunn, 1926*Eleutherodactylus orcutti* Dunn, 1928*Eleutherodactylus p. pantoni* Dunn, 1926*Eleutherodactylus pantoni amiantus* Schwartz and Fowler, 1973*Eleutherodactylus pentasyringos* Schwartz and Fowler, 1973*Eleutherodactylus p. planirostris* Cope, 1863 - Introduced*Eleutherodactylus sisypodemus* Crombie, 1977

Anura: Ranidae

Rana catesbeiana Shaw, 1802 - Introduced

Sauria: Diploglossidae

Celestus barbouri Grant, 1940*Celestus c. cruscus* Garman, 1888

Table 3.1 cont.

Celestus cruscus cundalli Grant, 1940
Celestus cruscus molesworthi Grant, 1940
Celestus duquesneyi Grant, 1940
Celestus fowleri Schwartz, 1971
Celestus hewardi Gray, 1845
Celestus microblepharis Underwood, 1959
Celestus occiduus Shaw, 1802

Sauria: Gekkonidae

Aristelliger praesignis Hallowell, 1857
Gonatodes albogularis notatus Reinhardt and Luetken, 1863
Sphaerodactylus a. argus Gosse, 1850
Sphaerodactylus dacnicolor Barbour, 1910
Sphaerodactylus gilvitorques Cope, 1862
Sphaerodactylus goniorhynchus Cope, 1895
Sphaerodactylus oxyrhinus Gosse, 1850
Sphaerodactylus parkeri Grant, 1939
Sphaerodactylus r. richardsoni Gray, 1845
Sphaerodactylus richardsoni gossei Grant, 1939
Sphaerodactylus semasiops Thomas, 1975
Sphaerodactylus sp.

Sauria: Iguanidae

Cyclura collei Gray, 1845

Sauria: Polychrotidae

Anolis garmani Stejneger, 1899
Anolis g. grahamii Gray, 1845
Anolis grahamii aquarum Underwood and Williams, 1959
Anolis l. lineatopus Gray, 1840
Anolis lineatopus ahenobarbus Underwood and Williams, 1959
Anolis lineatopus merope Underwood and Williams, 1959
Anolis lineatopus neckeri Underwood and Williams, 1959
Anolis opalinus Gosse, 1850
Anolis reconditus Underwood and Williams, 1959
Anolis s. sagrei Dumeril and Bibron, 1837 – Introduced ?
Anolis valencienni Dumeril and Bibron, 1837

Sauria: Scincidae

Mabuva sp.

Table 3.1 cont.

Sauria: Teiidae

Ameiva dorsalis Gray, 1838

Serpentes: Boidae

Epicrates subflavus Stejneger, 1901

Serpentes: Colubridae

Alsophis ater (Gosse, 1851)*Arrhyton callilaemum* (Gosse, 1851)*Arrhyton funereum* (Cope, 1862)*Arrhyton polylepis* (Buden, 1966)

Serpentes: Tropidophiidae

Tropidophis jamaicensis Stull, 1928*Tropidophis stejnegeri* Grant, 1940*Tropidophis stullae* Grant, 1940

Serpentes: Typhlopidae

Typhlops jamaicensis (Shaw, 1802)

Testudines: Emydidae

Trachemys terrapen (Lacepede, 1788)

Crocodilia

Crocodylus acutus Cuvier, 1807

Location - Topography - Geology

Jamaica is located about 200 km south of the eastern end of Cuba and about the same distance west of the tip of the Tiburon Peninsula of Haiti. The island is approximately 230 km long and 80 km wide at its broadest (central) point, tapering at both ends, with a total area of 11,500+ km² (Fig. 3.1). Its topography is very simple compared to the larger Greater Antillean islands, consisting primarily of a large, heterogeneous limestone block atop an igneous and metamorphic core, sloping slightly upward from south to north. The southern alluvial plain is fairly extensive and includes several ridges or uplifts (Portland Point and Hellshire Hills) that were probably islands during

periods of elevated sea levels. The dominant feature of the limestone block is the extensive flat or rolling central plateau, mostly 2000-2500 ft in elevation, with a variety of associated ridges (Don Figuero, Dry Harbour, and Santa Cruz Mountains) and karst areas, including the extremes of the Cockpit Country and the more weathered limestones of the western Round Hill District. Unlike the other Greater Antilles, Jamaica has a paucity of offshore islands, most of them associated with the south coast peninsulas of Portland Point and the Palisadoes (Steers, 1940a,b; Steers *et al.*, 1940; Lazell, 1996). Cabarita Island, off Port Maria, St. Mary, is one of the few north coast islets (Crombie *et al.*, 1984). The more distant Pedro (65 km south) and Morant Cays (95 km southeast) are not on the Jamaican Bank but are currently claimed by Jamaica.

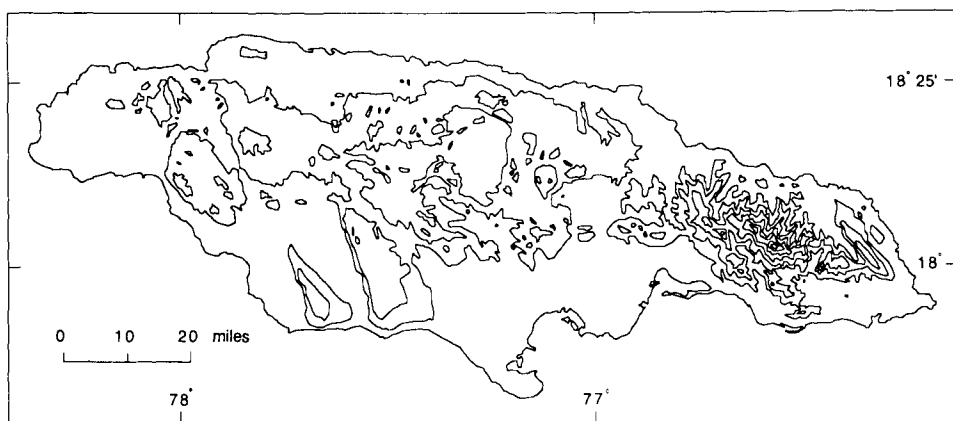


Figure 3.1. Topographic map of Jamaica. The contours mark 1000 ft. Most of the island's elevational features are associated with the limestone block which sits on top of the island. The greatest elevational changes occur on the eastern end as the Blue and John Crow Mountains.

The eastern tip of the island is a largely granitic-shale conglomerate that rudely contacted the limestone block by movement along a fault. The resulting compression and slip faulting caused the uplift of the eastern mountains (Blue and John Crow ranges), which reach an elevation of more than 2200 m. This uplift was a relatively recent event, generally thought to be Pliocene (Buskirk, 1985).

The geological history of Jamaica, including its tectonic movements, submergence, and reemergence, was thoroughly reviewed by Buskirk (1985) and Hedges (1989a,b). Various aspects of Caribbean geology (particularly regarding Jamaica) remain controversial, as does the debate on the role of vicariance versus dispersal in the herpetological colonization of the islands. I believe that more data are required before I would speculate on the "definitive" geological history of Jamaica since I do not agree totally with any of the proposals presented thus far. Rather than greatly expanding the literature cited herein by citing the same literature already summarized in several recent reviews, I suggest interested readers make their own conclusions from these more detailed discussions: Crother and Guyer (1996), Guyer and Crother (1996), and Hedges (1996a,b,c).

Rivers

Despite the porous limestone base of most of the island, Jamaica has a multitude of rivers draining to both coasts but with more emptying to the north. The Negril River and associated Morass is the major stream at the western end of the island and (moving eastward) other important waterways of the northern part of the limestone block include the Lucea, Great, Montego, Martha Brae, Rio Bueno, Roaring, White, Rio Nuevo, and the Wag Water. The southwest coast, considerably lusher than the south-central area, has numerous smaller streams and the Cabarita River system, but the single most extensive wetlands area in Jamaica is the Black River and its tributaries. With its large estuary and shallow bay, in addition to broad inland freshwater swamps, this area remains very poorly collected and barely explored. East of the Black River many river tracks are dry at least part of the year and the Milk River system is the primary drainage on the south-central coast. Other rivers meander through the central plateau and either vanish underground or join other rivers, usually flowing south.

The eastern mountains are also well supplied with rivers. The main systems draining north are the Buff Bay, Spanish, Rio Grande,

and Drivers. South of the mountains are the Plantain Garden, Negro-Morant, Yallahs, and Cane-Mammee.

This abundance of surface water is misleading since there is considerable evidence that suggests Jamaica was considerably drier and perhaps even devoid of surface water for long periods after its emergence, possibly until very recently in geological time. Most compelling is the absence of native frogs with larvae that develop in ponds or streams. All other Greater Antillean islands have xeric-adapted bufonids that can breed in ephemeral pools. Another Greater Antillean species, *Leptodactylus albilabris*, copes with xeric habitats by producing a foam nest which can provide food, insulate against extreme temperatures, and deter desiccation for the larvae. Although this species is ubiquitous on the Puerto Rican Bank, including some harshly dry areas, it is restricted to a small mesic enclave on extreme eastern Hispaniola and is absent from all islands to the west. Since Heyer (1978, personal communication, 1996) suggests South American rather than Middle American relationships for *albilabris*, it seems most likely that it is a recent adventive to Hispaniola and never reached the more western islands (Cuba and Jamaica). Jamaica lacks native bufonids and *Leptodactylus* but has 17 endemic *Eleutherodactylus* (direct development) and five hylids, even greater than the hylid radiation (four) on much larger Hispaniola. Whereas most other Antillean hylids are either very generalized pond breeders, including ephemeral pools (*Hyla pulchrrilineata*, *Osteopilus dominicensis*, and *O. septentrionalis*) or stream-adapted (*Hyla heilprini*, and *H. vasta*), all the Jamaican species have evolved highly specialized phytotelmatous larvae (Dunn, 1926a; Lannoo *et al.*, 1987; Thompson 1996), that usually develop in bromeliad axils but may use tree holes or other small cavities. The larvae of the undescribed *Osteopilus* are not definitely known but variation in the few known samples of supposed "*Calypthahyla*" *crucialis* larvae suggests that both species may be represented therein. Although there is some dispute whether the Jamaican hylids represent a single radiation (Hedges, 1996b) or convergence in larval morphology (Lannoo *et al.*, 1987; Anderson 1996), it is clear that they do not (and probably cannot) use ponds or streams for reproduction. In addition, some Jamaican *Eleutherodactylus* occasionally or routinely (*E. jamaicensis*) use bromeliads and other cavities as egg deposition sites, also suggesting an extended drier regime on the forest floor. Some genera of normally terrestrial reptiles such as anguid lizards and *Sphaerodactylus* have evolved obligate bromeliad species on Jamaica (*Celestus fowleri*, *Sphaerodactylus oxyrhinus*, and *S. semasiops*), as have a variety of invertebrates (carabid beetles,

crabs, scorpions, and worms). In addition, Pregill *et al.* (1992) found Holocene remains of xeric or at least open-formation birds and lizards (*Ameiva dorsalis*, *Leiocephalus cf. jamaicensis*) in a cave deep within the mesic Cockpit Country. While hardly conclusive, it is hard to imagine why such extensive adaptation for life in the mesic enclaves of bromeliads would have taken place if moist terrestrial refuges and more typical reproductive sites were available (Fig. 3.2).

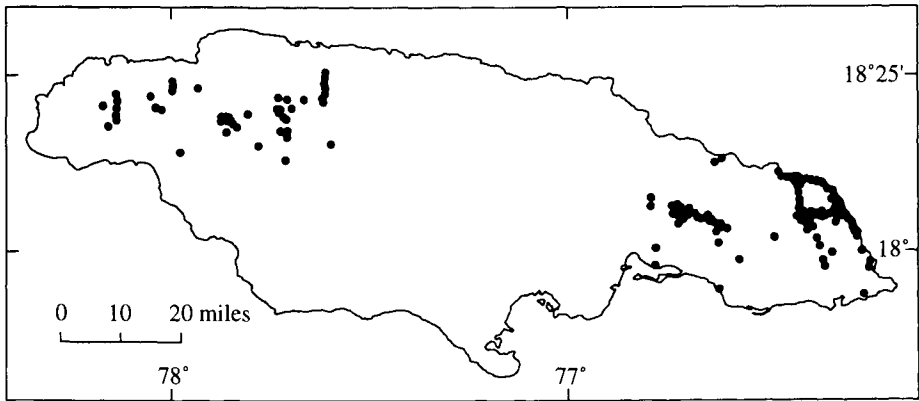


Figure 3.2. Combined superimposed distributions of 15 taxa which exhibit geographic ranges associated with the two major areas of endemism. The western distributions are represented by *Eleutherodactylus fuscus*, *E. griphus*, *E. sisypodemus*, *Celestus fowleri*, *Sphaerodactylus oxyrhinus*, and *S. semasiops*. The eastern distributions are represented by *E. alticola*, *E. andrewsi*, *E. glaucoreius*, *E. nubicola*, *E. orcutti*, *E. pentasyringos*, *Anolis reconditus*, *Arrhyton polylepis*, and *S. dacnicolor*.

Climate and Vegetation

Jamaica has a seasonal tropical maritime climate, dominated by north or northeast trade winds which bring substantial rainfall to most of the northern part of the island. The northern and eastern slopes of the Blue and John Crow Mountains can receive more than 7 m of rain annually and the Cockpit Country only slightly less. Most of the south coast is considerably more arid and the limited rainfall rapidly percolates away in the limestone areas. A 91-year survey of rainfall in the *Handbook of Jamaica* for 1963 reported a mean rainfall for the combined 14 parishes of 84.83 in., with a rainy season from May to October (6.92-12.15 in.) and a slightly drier one from November to April (2.91-7.74 in.). The mean maximum-minimum temperatures at sea level are 87.5 and 70.8°F. On the central plateau at around 2000

ft this drops to 78.6 and 66.1°F. The highest elevation for which climatological data are available is Hill Gardens, Cinchona, St. Thomas, at about 4400 ft, where mean highs are in the high 60s and overnight lows in the mid-50s. Higher in the Blue Mountains, overnight temperatures near 40°F are not uncommon.

A discussion of Jamaican floristics is almost a moot point since estimates of natural vegetation loss range from 75+% (Downer and Sutton, 1990) to more than 90% (Hedges, manuscript in preparation). Furthermore, most of this habitat destruction (at least 50%) took place during the early period of colonization, when forest was extensively cleared for agricultural and grazing land. Although this habitat alteration continues today, primarily in the higher elevations which were not suitable for early exploitation, it is relatively minor compared to the early clearing. Asprey and Robbins (1953: 361, Fig. 3) mapped their major vegetational types and even at that time the Wet Limestone Forest was centered in the Cockpit Country with disjunct patches in the western Parishes (Hanover and Westmoreland) and additional outliers as far east as St. Ann-St. Catherine. Many of these isolated patches were greatly reduced or nonexistent by the 1970s (R. Crombie, personal observation).

Similarly, Dry Limestone Forest was concentrated along the south coast, particularly the Hellshire Hills-Portland Point area, with isolates in the western parishes, east of Kingston in the rain shadow of the Blue Mountains, and a few small patches along the north coast. All of these have been severely reduced by development and overgrazing since Asprey and Robbins' report.

Montane Forest is restricted to the higher elevations of the Blue and John Crow Mountains. The Lower Montane Rain Forest and Montane Sclerophyll Forest have been severely depleted by development, as Kingston residents seek to escape to the cooler temperatures of the mountains. Still more restricted in area are the Mist Forest and Elfin Woodland, progressively higher on the peaks. However, it should be noted that these "upper" montane communities can extend to surprisingly low elevations in some areas (e.g., Elfin Woodland at 2500 ft in the John Crow Mountains, Asprey and Robbins, 1953: 402). For more detailed descriptions of these communities, the reader is referred to Asprey and Robbins, whom devote more than 50 pages and 20 figures to it.

The obvious question is, "What effect has this extensive deforestation had on the herpetofauna?" Since the most significant clearing of midelevation forest took place 200 years ago when most of the herpetofauna had not even been described, we have no "before" to

compare with the “after.” Forest-associated species such as many of the current Cockpit endemics may well have been widespread in forest throughout the Central Plateau and the current patterns of distribution are probably totally artifacts of early deforestation. Certainly most of the herpetofauna is ecologically very versatile, occurring in disturbed as well as pristine habitats, but distributional data on the Jamaican herpetofauna are so vestigial that even current range maps are only an approximation of the actual distribution for most species. Looking for historical changes with such paltry data would be pure speculation.

Historical Summary

I have divided this section into two parts: the historical literature, up to 1959, and the recent literature, 1960 to the present. Although this is admittedly arbitrary, there is a rationale behind it. Rather than follow a strictly chronological summary of the literature, it is more cohesive and space efficient to discuss the contributions of a given institution or individual as “eras” with a general chronological arrangement. In some cases (e.g., the Harvard era) the contributions span nearly a century and rather than subsume the more recent contributions with those from the late 1800s, I chose the division of “older” versus “recent” because it approximately corresponds to the time when Schwartz became involved in Jamaican herpetology; it also coincides with the departure of the most famous herpetologist resident in Jamaica, Garth Underwood. The contributions of some individuals (e.g., Coleman J. Goin) slightly span this arbitrary point, in which case I have included those individuals in the period that their publications began. In other cases (e.g., the Harvard era), the publications in both the older and more recent literature are so voluminous that I have separated them. This summary of the literature is not intended to be a bibliography of Jamaican herpetology, which would greatly exceed the space constraints of this chapter, but it is more a “highlights reel” of significant advances.

Historical Literature

Specimens from the New World colonies probably began arriving in Europe soon after trade ships began plying the West Indian waters. Precise locality data rarely were associated with the early specimens, at best only the island where the material allegedly originated, and even these minimal data were unreliable. For example, Gray’s (1839)

Tiliqua jamaicensis (type locality "Jamaica") is a synonym of *Diploglossus monotropis*, a species found in lower Middle and northwest South America. The type likely was not even collected in a British colony, evidence that the colonial powers did not respect each others territorial claims in the New World. Jamaican material even made its way to Paris, devoid of any locality (which might have been incriminating) and was studied by Daudin, Duméril and Bibron (e.g. *Anolis valencienni* Duméril and Bibron, 1837). The American colonies were not left out and Jamaican specimens in Philadelphia and Washington were described by Harlan, Hallowell, and Cope.

The earliest records of Jamaican reptiles consisted of anecdotal notes in voyage reports or travelogs, usually by surgeons who served as physician to the colonial governor. Grant (1940a: 146-148) adequately summarized most of them and pointed out the fanciful nature of the information therein. The most notable (or at least most extensive) of the early reports was the two-volume epic of Sir Hans Sloane (1725) on his extensive Antillean travels, including a 15 month stay in Jamaica in 1688 and 1689. Ahrenfeldt (1953, 1954) discussed Sloane's herpetological contributions (all appearing in the 1725 volume) in some detail, but Sloane's descriptions were all nonbinomial and not nomenclaturally available. Although some of his natural history information is historically significant, I agree with Gosse's (1848c: 61) comments on Sloane's observations: "like most of his zoological notes, ... full of confusion and error." Although Sloane apparently made a significant collection of plants (Ahrenfeldt, 1954), the status of his herpetological specimens is unclear. Gray (1845) did not list Sloane as a donor of material to the British Museum, the most logical depository. However, the French, whose relations with Britain were strained by conflicts in the New World, were uncharacteristically lavish in their praise of Sloane, even dedicating several species to this "learned traveler and friend of the Sciences" (Duméril and Bibron 1839: 107, as translated by Ahrenfeldt, 1954: 639). Perhaps Sloane's specimens were indeed deposited in Paris but without even general locality.

Following the Sloane tradition, Patrick Browne included some natural history in his three-part treatise on Jamaica (mostly in Part II), but his Latin descriptions were also nonbinomial, and some of his observations appear to be taken almost verbatim (including errors in spelling and logic) from Sloane. There has never been any suggestion that Browne actually collected any herpetological specimens, but his comments and descriptions were used by Lacépède, Shaw, and other later authors.

There is a tendency that the largest, most obvious, and edible species are usually the first taxa described from new territory, with the most unusual ones a close second. Following this trend, the first species described validly and expressly from Jamaica was the fresh-water turtle, *Trachemys terrapen* (Lacépède, 1789), although the description is marginal and the locality not as clear-cut as subsequent treatment would indicate. The page generally cited for the species (p. 129) does indeed contain a description for La Terrapen or Terrapene, nowhere therein are the names latinized or used in binomial form. Lacépède's name *Testudo terrapen* is nomenclaturally validated, somewhat debatably, only by the fold-out table that provides a latinized binomial (see discussion in Barbour and Carr, 1940:393). Barbour and Carr (1940) also point out that the species was described from "aux Antilles" and "particulièrement a la Jamaïque." However, since Lacépède clearly used Browne's (1789) comments on the species in his description, restriction of the locality to Jamaica is reasonable.

Also drawing from the works of Sloane (1725) and Browne (1789), Shaw (1802) described the largest Jamaican diploglossid and the endemic *Typhlops*.

Aside from these descriptions, most of the early Jamaican material was studied by John Edward Gray at the British Museum. Gray's descriptions were often peremptory, and the specimens poorly preserved, which compounded the taxonomic problems due to unreliable locality data. Still, Gray is responsible for six currently valid Jamaican species, including two of the most common and widespread anoles (*Anolis grahamii*, 1845, and *A. lineatopus*, 1840), a galliwasp (*Celestus hewardii*, 1845), the ground lizard (*Ameiva dorsalis*, 1838), the iguana (*Cyclura collei*, 1845), and the largest species of *Sphaerodactylus* (*richardsoni*, 1845). He also provided a significantly larger number of synonyms and species incorrectly attributed to Jamaica.

One of the American contributions preceding Gray's pioneering studies was Harlan's (1826b) description of the largest Jamaican tree-frog. Although *Hyla crucialis* was well and accurately described and the type specimen remains in the Academy of Natural Sciences (ANSP 2180, Crombie, 1973), the description was long overlooked, probably due to it being obscurely attached as a separately titled note to Harlan's larger paper (1826a).

All of the earlier work notwithstanding, the true father of Jamaican herpetology (and many other aspects of Jamaican biology) was Philip Henry Gosse, who lived on the island for 18 months in 1844 and 1845. Gosse was a careful observer and a talented writer who

traveled extensively throughout Jamaica. His "A Naturalist's Sojourn in Jamaica" (1851) remains an indispensable classic for anyone interested in the natural history of the island. Unlike many early visitors, who were confined to Kingston and areas easily accessible from it, Gosse was based in the Bluefields/Content region, now in Westmoreland Parish, in the far western limestone districts. Clearing of forests for plantations and lumber was common even at this early stage, but Gosse had easy access to the wet limestone forest that was less suitable for early development. Many significant localities such as the Black River Morass were not far away, but Gosse traveled even to the distant Blue Mountains and was probably the first European to see the full diversity of Jamaican habitats. He certainly was the first to describe it in print. As a result of his travels and meticulous observations, Gosse (1850, 1851) described 13 new species, seven of which are currently recognized as valid species or subspecies. Of his six synonymous names, *Placopsis ocellata* was described by Duméril and Bibron (1839) as *Anolis* (not *Xiphosurus*, as stated in Schwartz and Henderson, 1988) *valencienni* with no locality data, *Trachycephalus lichenatus* is Harlan's (1826b) *Hyla crucialis* (but see Problems section), and *Draconura catenata* is the Cuban species *Anolis sagrei*, obviously a very early introduction or natural dispersant. Gosse's descriptions of *Trachycephalus anochloros* (currently a synonym of *Calyptahyla crucialis*) and *Natrix capistrata* (= *Alsophis ater*) are so distinctive that I am not convinced the species are invalid, as discussed in the Problems section. Gosse also drew extensively from the notes of Dr. Anthony Robinson, a surgeon and botanist who never published his Jamaican observations. However, a copy of his manuscript was preserved in the Institute of Jamaica Library and many of the herpetological notes have since been published (Cockerell, 1894). Richard Hill of Spanish Town also contributed greatly to Gosse's work, particularly "The Birds of Jamaica," and Hill is often credited as being the first permanently resident naturalist of Jamaica.

An interesting adjunct of Gosse's work was the discovery of an unpublished color plate of his Jamaican amphibians and reptiles nearly 100 years after the publication of his "Sojourn" (Underwood, 1949). Chapman Grant disagreed with some of Underwood's identifications (Anonymous, 1950) and a revised identification key appeared in Lewis (1950).

After Harlan's (1826b) early note, the American contributions continued with Hallowell's (1857) description of *Aristelliger praesignis* and Cope's (1862a,b, 1895) additional new species (*Sphaerodactylus gilvitorques* and *goniorhynchus* and *Arrhyton funereus*). Al-

though most new species to this time had only "Jamaica" or an inferred more specific type locality, the two Jamaican species described in Cope (1895), based on material from the University of Pennsylvania West Indian Expedition of 1890-1891, actually had specific localities: *S. goniorhynchus* from Port Antonio (Portland Parish) and *Anolis flabellatus* from Port Morant (St. Thomas) and Port Lucea (Hanover), at opposite ends of the island. Cope included data on coloration, ecology, and behavior from the notes of the expedition. The two *Eleutherodactylus* species Cope reported from the expedition (*Hylodes martinicensis* from Port Lucea and Blue Peak; *Lithodytes lentus* from Port Lucea) are not identifiable (but clearly not the Lesser Antillean *martinicensis* or the St. Croix endemic *lentus*), but the description (handsome black and orange tree-toad ... clinging to a shrub overhanging a stream) of the latter could be *E. jamaicensis*. Although *A. flabellatus* is a synonym of *Anolis opalinus*, the description warrants additional discussion because it contains much of the confusing and contradictory information that typifies so many Cope descriptions. Cope clearly had more than one specimen of *flabellatus*, but his description and measurements were from one individual and variation in the other(s) was briefly noted. The material from this expedition was deposited at the Academy of Natural Sciences, Philadelphia (ANSP) but Malnate (1971) did not list any types of *flabellatus* in the collection. By way of (brief) explanation, Cope traveled frequently between Philadelphia and Washington, usually carrying specimens from both institutions to facilitate study at whichever museum was his current destination. Although perhaps authorized by the authorities of United States National Museum (USNM) and ANSP, these transfers were NOT invoiced or documented in any way and seemed entirely at Cope's discretion. Unfortunately, his organization of research materials was negligible and after his death utter chaos reigned in attempting to return specimens to their proper institutions, which was never really achieved. As a result, many ANSP specimens (including some types) were catalogued at USNM and vice versa, obviously an attempt to simplify what must have been a monumental task. Cope also removed tags when illustrating specimens (or for some other unfathomable reason) and after his death USNM received a sizable batch of these tagless specimens, which were recatalogued as "returned by Cope's estate." They were usually completely dataless, but occasionally a label associated with a specimen (or specimens) provided a shred of locality information. Because of these informal "exchanges" between ANSP and USNM, researchers should be aware that Cope's types should be sought at both institutions, regardless of

any specific comments on their deposition in Cope's descriptions. Additional missing types may well be included (but unrecognized) in the now dataless "returned by Cope's estate" material in USNM. The types of *A. flabellatus*, listed as "unlocated" by Schwartz and Henderson (1988: 158), are a good example of the "unintentional exchange" due to Cope's transport of specimens between the two institutions. Although not listed in Cochran's (1961) USNM type catalog, two specimens of Cope's *flabellatus* remained on a "problem shelf" in USNM until April 1968, when they were finally recognized as missing syntypes and cataloged as USNM 164931 (from Port Morant, ledger remarks "type") and 164932 (Port Lucea, "type of drawing").

The end of the nineteenth century also marked the beginning of the "Harvard era" in West Indian herpetology, a notable period that continues to the present day. Samuel Garman made collections for the Museum of Comparative Zoology (MCZ) throughout the Antilles, including a brief visit to Jamaica. Although most of the other islands he visited yielded a treasure trove of novelties, Garman (1887a) described only a single currently valid new species from Jamaica, the most common small diploglossid, *Celestus crusculus*. Garman's activities appear to have been limited to the Kingston, Spanish Town, and Moneague area, but his successor, Thomas Barbour, was far more successful in reaching more distant parts of the island through his contacts with major plantation owners. Barbour (1910) added two more valid taxa to the faunal list, including the first indication that Jamaica was inhabited by more than one widespread, exceedingly variable species of *Eleutherodactylus*. With the exception of Harlan (1826b) and Gosse (1851), Jamaican amphibians were neglected by the early writers and Gosse's *Litoria luteola* was the only species of *Eleutherodactylus* recognized on the island. Barbour's (1910) description of *Eleutherodactylus jamaicensis*, an arboreal, bromeliad dwelling species with expanded digital tips and sticky bluish skin secretions, was particularly noteworthy and the species so distinctive that some subsequent authors (Goin, 1954; Crombie, 1977; Flores, 1984) regarded it as unique among Jamaican and even West Indian *Eleutherodactylus*. Barbour's other new species was *Sphaerodactylus dacnicolor*, also an arboreal bromeliad dweller. Barbour (1922) later produced the first list of the Jamaican herpetofauna and discussed Jamaican species in his West Indian checklists and taxonomic reviews. However, his later reviews of Jamaican species (particularly *Anolis* and *Sphaerodactylus*) lacked the initial clarity of his 1910 paper.

Although he taught at Haverford College, Emmett Reid Dunn could be included in the Harvard era due to his strong institutional af-

filiation with MCZ and his long friendship with Barbour. Dunn single-handedly revamped the Jamaican amphibian fauna in just 10 days of fieldwork in August and September of 1925. He worked primarily at Spaldings, on the Clarendon-Manchester Parish border, near where the weathered central plateau meets the karst of the Cockpits and Dry Harbour Mountains. Dunn (1925) had previously described *Hyla wilderi*, named after and based on specimens collected at Moneague, St. Ann, by Harris Hawthorne Wilder and Inez Whipple Wilder, colleagues from Smith College. The Wilder's interesting discovery may have prompted Dunn to stop in Jamaica on his return from Cuban fieldwork in 1925, but whatever his motivation, the 10 days of frog-oriented fieldwork yielded all seven species previously known from the island and seven new species. Except for *Eleutherodactylus nubicola*, based on older material from two localities in the Blue Mountains (which Dunn did not visit), the new *Hyla (marianae)* and five new *Eleutherodactylus* were all from Dunn's Spaldings collections. In more than tripling the number of endemic *Eleutherodactylus*, Dunn conclusively demonstrated that Jamaica did indeed have a significant radiation of the genus, like all other Greater Antillean islands. He later (1928) described the Blue Mountain aquatic frog from Arntully, St. Thomas as *Eleutherodactylus orcutti* and *cunctator*, the latter a minor variant and synonym of the former. The material Dunn (1928) reported as *Eleutherodactylus grabhami* from Arntully (not cited by number in the text but USNM 73862-64) was later reidentified as *E. pantoni* by Lynn (probably = *E. pentasyringos*). Other contributions included an update and key for Jamaican frogs (Dunn 1927) and valuable natural history notes on what is now known as *Calyptohyla crucialis* (*Hyla lichenata*, 1929). Dunn's patronyms for some of his new species acknowledged several prominent Jamaican naturalists (E. Stuart Panton of Mandeville, Frank Cundall of the Institute of Jamaica, and Philip Henry Gosse) and his Jamaican hosts (Percy Junor of Spaldings and the unidentified Mr. Grabham). Dunn's etymology (or lack thereof) for *Hyla marianae*, a female patronym for "Marian", presented an interesting sidelight of mystery. The enigmatic "Marian" is neither mentioned nor identified in ANY of Dunn's works, and queries to Dunn's wife and friends (including Roger Conant and H. G. Dowling) yielded no clue to the identity of the "mystery woman" commemorated in the description of this very distinctive and attractive little treefrog.

Following a short hiatus, the MCZ contributions to West Indian herpetology continued with the work of Ernest E. Williams and his multitudes of students, primarily on *Anolis*. Their taxonomic contri-

butions on Jamaica included the descriptions of four new subspecies of *Anolis grahamii* and *lineatopus* and the Blue Mountain endemic, *Anolis reconditus* (Underwood and Williams, 1959). Williams also maintained close contact with colleagues at the American Museum of Natural History (AMNH), Max Hecht and Karl Koopman, often traveling in the field with them. Hecht's (1952) work on *Aristelliger* contained information on the Jamaican *A. praesignis* and the description of a giant fossil species, *A. titan*. Etheridge (1965) synonymized *A. titan* with the Hispaniolan *A. lar*, but I disagree with this allocation.

The USNM Era

Although Leonhard Stejneger and Doris Cochran were doing extensive work elsewhere in the West Indies, their contributions to Jamaican herpetology were minimal. Stejneger's impeccable nomenclatorial and historical background compelled him to correct the common usage of names for the largest Jamaican anole (1899) and the Jamaican boa (1901), the latter a clear acknowledgment of Sloane's "Serpens major subflavus". Although USNM collectors (including E. A. Chapin, H. L. Clark, W. Harris, W. R. Maxon, G. S. Miller, Jr. and C. R. Orcutt) and collections provided resources for other researchers on Jamaica, the USNM staff added nothing further to the literature until my field work began in 1970.

Lynn and Grant Period

The fieldwork and collections of W. Gardner Lynn and Chapman Grant produced another spate of taxonomic advances and a great increase in our knowledge of the basic biology of the Jamaican herpetofauna. Lynn's extensive travels in the summers of 1932, 1936, and 1941 yielded two new Blue Mountain endemics (*Eleutherodactylus alticola* and *andrewsi*, Lynn 1937), the Portland Ridge cave frog (*E. cavernicola* Lynn, 1954), and the cryptic western limestone species *E. fuscus* (Lynn and Dent, 1943). Lynn and Dent (1942) also described a very distinctive little frog from western Jamaica as *Eleutherodactylus lewisi*, in honor of C. B. Lewis of the Institute of Jamaica. At this time, Gosse's name *luteolus* was being misapplied to the most common and widespread Jamaican *Eleutherodactylus*, but Lynn and Dent (1942) actually redescribed real *luteolus*, as pointed out by Goin (1953). Lynn (1937) also noted the introduction of the Cuban species *Eleutherodactylus ricordi* based on some specimens from Montego Bay. Schwartz (1965c) later demonstrated that *ricordi* and *planirostris* were distinct species and that *planirostris* was the common and widespread taxon that had been introduced in Jamaica

(1973c). In addition to his taxonomic contributions, Lynn (1940) provided substantial ecological information on Jamaican amphibians and a useful historical summary of the literature. Both he and his students used Jamaican material in embryological studies (Lynn, 1936, 1942, 1944; Lynn and Peadon, 1955; Schreckenberg, 1956) and papers on the descriptive morphology of the thyroid (e.g., Lynn and Walsh, 1957). The embryological studies were important additions to our understanding of clutch size and other life history parameters. Lynn (1936) also pointed out that the Sampson's (1904) paper was based on two and possibly three Jamaican species, none of which was actually *Hylodes* (= *Eleutherodactylus*) *martinicensis*. Lynn and all other authors believed that *E. martinicensis* had been introduced and established in Jamaica from the Lesser Antilles, however. Schwartz (1967c) documented that *martinicensis* and *johnstonei* were valid Lesser Antillean species and that the latter had been widely introduced elsewhere.

Grant's Jamaican fieldwork was performed in March-May 1937, March 1938, and April 1946. In searching for reptiles, he visited more xeric areas than Lynn, and their collections complemented each other nicely. Grant named 12 new taxa (three species and nine subspecies) that have received varying degrees of acceptance. In reviewing the large-scaled *Sphaerodactylus*, Grant (1939b) noted the striking differences between the north and south coast populations, corrected Barbour's (1921) confused treatment of them, and proposed *S. parkeri* for the south coast species. He also described a subspecies of the north coast *S. richardsoni* from the mouth of the Roaring River, St. Ann, but variation in *richardsoni* has never been carefully analyzed (Schwartz and Henderson, 1988: 190) and the status of *S. r. gossei* remains questionable.

To his credit, Grant was the first to recognize that Jamaica had a radiation of diploglossine lizards second only to Hispaniola and he did much to clarify the many synonyms and species erroneously reported from Jamaica. He first (1940a) described *Celestus duquesneyi* from the xeric Portland Ridge, Clarendon, a *hewardii*-like species notable not only for its blue-banded tail but also for its habitat (all other larger Jamaican *Celestus* are decided mesophiles). The much more widely distributed *Celestus barbouri* (Grant, 1940b) probably escaped detection and description because earlier authors often regarded the smaller species as juveniles or subadults of *occiduus* or *hewardii*; for those who recognized that these "juveniles" were adults, Cope's *Celestus impressus* (1868) was a convenient catch-all name for moderate sized species. Grant (1940b) discovered that *impressus* was actually based

on one specimen of *hewardii* and one of *crusculus*, clearly not applicable to the intermediate-sized *barbouri*. Schwartz (1964: 56) later resolved the problem by designating the larger syntype (ANSP 9225, clearly the specimen Cope described) as the lectotype of *impressus*, firmly placing it in the synonymy of *hewardii*. Had the other syntype (ANSP 9226) been a justifiable lectotype selection, the name *impressus* would have antedated and replaced *crusculus* Garman (1887a).

Other aspects of Grant's diploglossine arrangement were far less rational. He regarded *Celestus occiduus* and *hewardii* as subspecifically related, which Cousens (1956) dismissed as ludicrous. His two new subspecies of *C. crusculus* were also rejected by Cousens, but she admitted that *molesworthi* was possibly worthy of recognition. Schwartz (1964: 55) disagreed and felt that there was definite subspecific differentiation of *crusculus* on Jamaica and that, "likely some or all of the races recognized by Grant are valid." He later (Schwartz and Henderson 1988: 97) concluded that "geographic variation in this species is more complex than considered by Grant." However, in the absence of more detailed analysis, Grant's arrangement still stands (see Problems).

Continuing his preoccupation with subspecies, Grant described two new dwarf boas, *Tropidophis maculatus stulli* and *T. pardalis stejnegeri*, and recognized Stull's *T. m. jamaicensis* as yet another Jamaican subspecies. Schwartz and Marsh (1960) transferred all three as subspecies of the Hispaniola *T. haetianus*, concluding that, "certainly there is no question of the distinctness of the three Jamaican forms from one another." S. Blair Hedges (personal communication, 1996) informs me that all three taxa are valid species, more closely allied to the Cuban radiation, thus confirming Grant's suspicions on their relationships if not his subspecific arrangement. Grant's other subspecies (three of *Anolis lineatopus*, one of *Sphaerodactylus argus*) were all very weakly defined and not placed in the perspective of variation throughout the species distribution. Thomas (1975) synonymized *S. argus henriquesi* with the nominate subspecies. Underwood and Williams (1959) retained *A. lineatopus neckeri* but synonymized *coxi* and *lynni* with the nominate subspecies.

It should be noted that Grant's Jamaican descriptions were published in such an idiosyncratic and inconsistent way that they had potential nomenclatural consequences, which fortunately never arose. The first and simplest problem involves the date of publication of the Institute of Jamaica *Science Series Bulletins* on the herpetology of Jamaica and the Cayman Islands (Grant, 1940b, 1940c). Grant con-

sistently listed the date of both as 1941 in his personally issued bibliographies, although he cited the dates as 1940 in some of his later publications (e.g., Grant, 1951). Lynn's bibliography (USNM reprint files) lists *The Herpetology of Jamaica* as published in 1940. Grant greatly respected Leonhard Stejneger and maintained close contact with both him and Doris Cochran throughout his career, so it is likely that Grant sent Stejneger copies of his papers as soon as possible. The USNM copy of the Jamaican monograph is marked (in Stejneger's hand) "received 30 Aug 1941," and the *Natural History Notes of the Natural History of Jamaica* for April 1941 (Vol. 1, No. 1) announces that the publication was "now on sale at the Institute bookshop." I was unable to resolve the date problem at the library of the Institute of Jamaica in the 1970s, but things were somewhat in disarray at that time, so I continue to use the "traditional" date of 1940 for the only species actually described in the 1940b monograph, *Celestus barbouri*. It is clear that the "preliminary descriptions" in *Jamaica Today* (Grant, 1940a) appeared first since Grant cited it in 1940b issue (including pagination) as the original description for most of the new taxa. However, both *Tropidophis maculatus stulli* and *T. pardalis stejnegeri* were listed as "subsp. nov." in the 1940b issue, with no mention of their previous description in the 1940a issue. It could be argued that in the *Jamaica Today* article's the descriptions were not nomenclaturally valid descriptions, published in a journal that was not "generally available," but all new taxa described therein were accompanied by type localities, numbers for the holotypes, and diagnostic characters, albeit brief. It is unclear why Grant considered seven of the nine taxa as validly described in the 1940a issue but specifically proposed the two *Tropidophis* as "new" in the 1940b issue, but the original description of all must stand as in the 1940a issue. It should also be noted that although he clearly dedicated the taxon to "Mrs. Olive Stull Davis" (1940a,b) he used a masculine ending for his patronym. Schwartz and Fowler (1973: 131) properly corrected it to *stullae*, but without comment.

Goin/University of Florida

Coleman Jett Goin made his primary contribution in describing and quantifying the pattern of polymorphism in Jamaican *Eleutherodactylus*, with preliminary data on the genetic basis of this variation (Goin, 1950, 1954, 1958, 1960). His summer fieldwork in 1948, 1950, 1952, and 1957 resulted in sizable herpetological collections that formed the basis of his genetic studies and were also used in his work on the maxillary dentition of frogs (Goin, 1959a). However,

taxonomic progress was (and still is) the inevitable result of fresh field collections and Goin's were no exception. Until Goin's work, few researchers had attached much importance to the lack of agreement between Gosse's (1851) description of *Litoria luteola* and the common *Eleutherodactylus* found throughout Jamaica to which the name had been applied. Dunn perhaps had an inkling when he described *E. gossei* (1926a) but he still mistakenly referred other *gossei* to *luteolus*. Lynn (1940) synonymized *gossei* with *luteolus* to return to the status quo. Goin collected specimens that matched Gosse's description and convincingly demonstrated that *E. luteolus* was a western Jamaican species with a fairly restricted distribution in that limestone region (1953). Although *Eleutherodactylus lewisi* (Lynn and Dent, 1942) was clearly a synonym of *luteolus*, Goin never suggested it and the synonymy first appeared in Cochran (1961: 45), possibly on the advice of Dr. Lynn. Appropriately, the most widespread and abundant Jamaican frog was now named *Eleutherodactylus gossei*, a fitting tribute to Gosse's pioneering contributions. Goin also made the first attempt at a phylogeny of Jamaican frogs with his comments on the relationships of the *gossei* group.

Goin described only one new species, *Eleutherodactylus lynni* (Goin and Cooper, 1950), which Schwartz and Fowler (1973) synonymized with *E. cundalli* (but see Problems). Goin also noticed that Taylor (1952) described a Dunn specimen of *Hyla wilderi* (erroneously cataloged with Panamanian locality data) as a new species, *Hyla shrevei*. Ironically, Taylor (1952: 1) stated of his supposed new species, "It is presumed that it is a rivulet species, living in the neighborhood of small streams, rather than a bromeliad species." Not only was Taylor's locality information for this species incorrect, but also his assumptions about its ecology were also 100% wrong, since *Hyla wilderi* is a bromeliad obligate. Although the University of Florida was (and continues to be) very involved in various aspects of Jamaican biology, the only other herpetologically notable paper of this period was Laessle (1961), which contains considerable information on hylid frog larvae and has import for other Jamaican bromeliadicoles.

The Underwood Era

After his Army service in India during World War II, Garth Underwood accepted a position at the University of the West Indies, teaching at both the Jamaican and Trinidadian campuses. His remarkably diverse reptilian interests utilized a variety of Jamaican species for broader evolutionary studies, including, for example, papers on the visual system (1951), digital structure (1954a), systematics

(1954b), the circulatory system (Adams *et al.*, 1957), and syntheses of evolutionary processes (1977). Underwood was active in the Natural History Society of Jamaica, becoming president in 1953, and he published several popular and educational papers in their *Natural History Notes* (1949, 1950, 1954c,d,e), including his multipart "Introduction to the Study of Jamaican Reptiles" (1951-52). His taxonomic contributions include the classic study of Jamaican anoles with Ernest Williams (1959), which resolved the status of older names by examination of type specimens, synonymized most of the subspecies Grant (1940b) recognized, and reanalyzed variation of each species throughout Jamaica, resulting in a redefined *Anolis grahamii* (with two subspecies) and *lineatopus* (with four subspecies). They also described the Blue Mountain endemic, *Anolis reconditus*. Later, Underwood (1959a) described *Diploglossus microblepharis*, which remains known only from the type, and reraised the question of whether the diploglossine genera *Celestus* and *Diploglossus* are indeed distinct, a debate that continues unresolved (see Problems). Underwood's departure from the West Indies in 1960 marked the end of a productive and important period of contributions by a resident biologist. However, it is noted with pleasure that Dr. Underwood has recently (1993) continued his Antillean studies with a paper on the *Clelia* of the Lesser Antilles, long an area of special import to him.

In addition to Underwood, other members of the Natural History Society also published herpetological information in the *Natural History Notes* (nearly 50 titles by my count). Some of these were trivial, but others contained valuable ecological (e.g., Lewis, 1941, 1943; Panton, 1952), behavioral, or distributional data, including information on the spread of *Eleutherodactylus martinicensis* (= *johnstonei*) from the area of original introduction in Kingston (Perkins, 1942; Anonymous, 1943; Jeffrey-Smith, 1946).

Modern Era

It is only appropriate that the discussion of the "modern literature" should begin with the Harvard contributions. Under the tutelage of Ernest E. Williams, Harvard students and postdocs produced an enormous number of papers on virtually all aspects of anole biology, many of them involving Jamaican species only obliquely if at all. A representative (but by no means complete) selection of those most important for Jamaica follows, arranged alphabetically by author: Blake (1986), Floyd and Jenssen (1983), Gorman and Atkins (1968a), Hicks (1973), Jenssen (1973, 1977, 1979), Jenssen and Andrews (1984), Jenssen and Nunez (1994), Lazell (1966), Licht and Gorman

(1970), Rand (1964a, 1967a,b), Schoener and Schoener (1971a), Taylor and Gorman (1975), Trivers (1976), Wyles and Gorman (1980a). Although most of the data for the papers were collected on short visits to Jamaica, both A. S. Rand (August 1961 - June 1962 in Kingston) and Thomas A. Jenssen (early 1970s in Mandeville) were resident while conducting their ecological studies. Certainly the major thrust of the Harvard studies was *Anolis*, but other Jamaican contributions included an important paper on xenodontine snakes (Maglio, 1970) and an insightful thesis on relationships of *Eleutherodactylus* (Flores, 1984).

Harvard has not had a monopoly on *Anolis*, however, and several recent authors discussed Jamaican species, e.g., Landwer *et al.* (1995), Losos (1990a,b,c, 1992), Losos and Sinervo (1989), Powell and Russell (1992). Peter Vogel at the University of the West Indies, Mona, has studied *Anolis lineatopus* in some detail (Vogel, 1983, 1984; Vogel and Brockhusen-Holzer, 1984; Vogel and Curio, 1983; Vogel *et al.*, 1986). He also published an interesting paper on *Cyclura* (Vogel *et al.*, 1996), adding substantially to Woodley's (1971, 1980) "rediscovery" of the species.

Frogs were much less popular than anoles and only a few papers on ecology and morphology appeared during this period: Garrick *et al.* (1985), Lannoo *et al.* (1987), Pough *et al.* (1977), Stewart (1979), Stewart and Martin (1980), and Trueb (1970). Mahon and Aiken (1977) in the "first" report of *Rana catesbeiana* from Jamaica noted that 22 pairs were intentionally introduced in the Black River in March 1967 and were well established by the time of their paper. Unfortunately, as early as April 1946 Chapman Grant reported sight records of ranids (which he suspected were *R. catesbeiana*) from the "Baldwin, Main, or Parnassus Rivers" (Grant 1946). Taxonomic papers included some by Crombie, Hedges, and Schwartz, Trueb (1972), and Trueb and Tyler (1974). Trueb (1972) unsuccessfully attempted to have Harlan's *Hyla crucialis* suppressed for nomenclatural purposes. Trueb and Tyler (1974) ambitiously attempted to resolve higher systematics of Antillean hylids but suffered from a lack of field experience in the area. The genus *Osteopilus* was resurrected for most of the casque-headed species (including the Jamaican *Hyla brunnea*), but *lichenata* (= *crucialis*) was placed in a monotypic genus, *Calypthahyla* (see Problems).

Mike Seidel included the Jamaican pond-turtle in his revisions of Antillean emydids (1988a, 1996; Seidel and Atkins, 1987).

In terms of cumulative field time, comprehensiveness of collections, and geographic scope, three groups are notable in the "Modern"

period: Al Schwartz, Blair Hedges, Ron Crombie and their respective associates. Each of these will be discussed in greater detail.

Albert Schwartz

As noted previously, Albert Schwartz did not publish extensively on exclusively Jamaican topics but this is somewhat misleading in a number of ways. Schwartz was a master of including nuggets of information (often substantial ones) on topics quite unrelated to the title of the paper. For example, Schwartz (1964b) included very significant taxonomic information on Jamaican "*Diploglossus*" (including designation of a lectotype for *D. impressus*) in a paper on the Hispaniolan *costatus* group. I learned very early that each and every Schwartz paper should be read carefully and thoroughly.

Schwartz's Jamaican fieldwork began in 1961 (June-August) with additional collections in 1967 (June-September), 1969, and August 1970 (as summarized in Schwartz and Fowler, 1973). The earliest collection was the basis for Buden's (1966) revision of Jamaican *Dromicus* (now *Arrhyton*, sensu Maglio, 1970), including the description of *D. polylepis* from the northeast coast. With the exception of Thomas's (1975) *Sphaerodactylus argus* group review, the rest of the reptile collections were never summarized. However, Schwartz and Fowler (1973) used the frog collections to produce the humbly titled "Progress Report," with the stated purpose of bringing the, "knowledge of Jamaican frogs closer to that of the frogs of other Greater Antillean islands." It succeeded admirably. With his characteristic thoroughness, Schwartz (and his associates) collected fresh material of every Jamaican species except *Eleutherodactylus alticola*, and the wealth of associated ecological data presented in the 1973 paper far surpassed everything accumulated by others to that point. The keen Schwartzian eye for variation resulted in the description of four new subspecies (one each of *Eleutherodactylus cundalli* and *gossei*, and two of *E. pantoni*), two of which were later elevated to full species. Schwartz and Fowler (1973) also synonymized *E. lynni* (Goin and Cooper, 1950) with *E. c. cundalli*, but I remain unconvinced that this was justified (see Problems).

Other contributions included description of the bromeliad-dwelling *Diploglossus fowleri* (Schwartz, 1971a), a thorough summary of information on the Jamaican iguana in a review of the genus *Cyclura* (Schwartz and Carey, 1977), and a re-evaluation of the distinctive and unique holotype of *Sphaerodactylus gilvitorques* (Thomas and Schwartz, 1974). Thomas's (1975) review of the *Sphaerodactylus argus* group included the description of the striking, ocellate

Sphaerodactylus semasiops. (I've tried hard to forgive Richard for getting to this amazing beast first.) Thomas also considered *Sphaerodactylus dacnicolor* and *oxyrhinus* to be conspecific (but recognizable subspecies), despite their thoroughly different habitat (terrestrial vs bromeliads) and broadly allopatric distributions at opposite ends of the island. They are once again considered full species.

S. Blair Hedges

Blair Hedges first visited Jamaica in 1986 with one of my field teams. With National Science Foundation support during his tenure at the University of Maryland and later at Penn State University he and his associates have made numerous trips to the island to continue their pursuit of tissue for molecular analysis. An impressive number and variety of papers have emerged from the Jamaican side of Blair's Antillean-wide project, some of which I even agree with. Among the topics addressed are *Anolis* relationships and biogeography (Burnell and Hedges, 1990; Hass *et al.*, 1993; Hedges and Burnell, 1990); *Eleutherodactylus* relationships (Bogart and Hedges, 1995; Hedges, 1989a,b); natural history data (Hedges, 1987, Hedges *et al.*, 1994); protein variation in *Typhlops jamaicensis* (Hedges, 1989c); and general Caribbean biogeography (Hedges, 1982, 1996 a,b,c; Hedges *et al.*, 1994). Hedges' primary interest was relationships of species throughout the Antilles and his interpretations of Jamaican species have been controversial. His Jamaican *Eleutherodactylus* phylogeny is so totally opposed to earlier gut-feeling concepts (Goin, 1954; Schwartz and Fowler, 1973; Crombie, 1977, 1985) and more carefully tested arrangements (Flores, 1984; Joglar, 1989) that it requires additional analysis. The few alpha-taxonomic conclusions (*Eleutherodactylus glaucoreius* as a valid species and confirmation of *pentasyngos* as distinct, Hedges, 1989a) are valid.

Ronald I. Crombie

My Jamaican fieldwork began with a 6-week trip in the summer of 1970 and continued mostly in the summer months in increments of 1-3 months, until 1985, with more than 15 months total field time. Collections were made in every month of the year except November and December. Virtually every stretch of passable road was traveled and overland hikes were made across every area of significant topography, including transects of the Cockpit Country from south to north (twice), west to east, and east to west. Collections were made from every parish and 95% of the herpetofauna was sampled. Taxonomic advances included description of the two minute Cockpit en-

demic *Eleutherodactylus* (Crombie, 1977, 1985) and elevation of *pentasyringos* to a full species (1985). General vertebrate surveys of Cabarita Island (Crombie *et al.*, 1984) and the Cockpit Country (Pregill *et al.*, 1992) were also published. Additional descriptive work on larval morphology of hylids and acoustic analyses of frog calls is planned, in addition to description of the new *Osteopilus*.

Problems

Throughout this chapter I have mentioned “problems” involving the taxonomic status or other aspects of some species; I will discuss them in taxonomic order. However, I should preface this section by stating that in general, data on the distribution, systematics, and natural history of Jamaican amphibians and reptiles remain so basic that very few “definitive” statements can be made. The following assessment is very much my personal opinion, but it is based on a substantial amount of data and experience. Some may disagree (and I know some do), and others may fault my “objectivity”. I tend to expect an author’s opinions on controversial subjects, not an egalitarian discourse.

Hylidae

The Jamaican hylid frogs remain a problem on several levels. The undescribed *Osteopilus* is represented by only four specimens from three localities, despite its broad distribution and apparent abundance (based on call surveys). Schwartz and Fowler’s (1972) record of “*Calyptahyla*” *crucialis* from eight miles south Seaman’s Valley, Portland is probably this species. *Osteopilus* sp. nov. is about the same size as *O. brunneus*, with the cranial ornamentation of *crucialis*, and its call is somewhat intermediate between the two. It is clearly not a hybrid (confirmed by molecular data, S. B. Hedges, personal communication), but it does negate the morphological distinction between the genera *Osteopilus* and *Calyptahyla*, hence my use of “*Calyptahyla*” *crucialis* in this chapter. My analysis of the osteology of *crucialis* based on two additional specimens (MCZ 11251 dry and USNM 252374, cleared and stained) indicates that most of Trueb and Tyler’s osteological characters distinguishing *Osteopilus* and *Calyptahyla* are either equivocal (characters 1, 2, 19, 20) or aberrant (6) in the one skeleton they used. Both of my skeletons have a normal (= *Osteopilus*-like) cultriform process of the parasphenoid, very unlike that illustrated for BMNH 52.12.1 by Trueb and Tyler. The only dif-

ferences between the skulls of *crucialis* and *brunneus* are minor details of decoration seen within other genera of casque-headed hylids (e.g. *Trachycephalus*). When a skeleton of *Osteopilus* sp. nov. becomes available I anticipate that the osteological distinctions between the two genera will be even more negligible. Morphologically, the major difference is the lack of nuptial pads in males of *Calyptahyla* and their presence in the four *Osteopilus* (including sp. nov.). Hedges (1996c) suggested that *Calyptahyla* was a synonym of *Osteopilus* based on molecular data, and I agree completely.

However, Hedges (1996c) also concluded that the two smaller Jamaican hylids (as well as the Hispaniolan *Hyla pulchrrilineata* and *vasta*) were also referable to *Osteopilus*. This is not a novel idea, since Dunn (1926a) first suggested that *wilderi* and *marianae* might be "neotenic" forms of *brunneus* and Pregill (1981a) described a similar form of paedomorphosis in the Antillean bufonids. Although the skulls of *wilderi* and *marianae* do resemble juvenile *Osteopilus* in some respects, because of major differences in larval morphology (i.e., Lannoo *et al.*, 1987) I am skeptical of Hedges' arrangement. Clearly the generic level relationships of Jamaican (and other Antillean) hylids needs careful analysis.

Leptodactylidae

The roster of Jamaican *Eleutherodactylus* may not be complete. Zoogeography would argue for a high-elevation endemic in the John Crow Mountains and I have heard unfamiliar calls there. Call variation in *E. jamaicensis* suggests that east-west speciation may have occurred in this species. *Eleutherodactylus cundalli* also has unprecedented variation in morphology, call, and ecology, particularly in the startling reproductive behavior of the Windsor Cave "cundalli" reported by Diesel *et al.* (1995). Early in my fieldwork I noted differences in size, skin fragility, limb proportions, and calls of cave-associated versus forest *cundalli* in the western limestone regions, similar to the characters Goin and Cooper (1950) used to diagnose *E. lynni*. Schwartz and Fowler (1973) discounted this as intraspecific variation but I remain unconvinced.

Diploglossidae

Much remains to be done on the relationships and biology of the Jamaican galliwasp. As noted above, the subspecies of *Celestus crucculus* remain controversial. Grant's subspecies *molesworthi*, from the highly endemic northeast coast, is very distinctive and almost surely a valid species, but *cundalli* from the central plateau is poorly defined

from the nominate, xerophytic subspecies. *Celestus barbouri*, *duquesneyi*, *fowleri*, and *microblepharis* are generally very poorly represented in collections, with two of them (*fowleri* and *microblepharis*) known only from the types. The validity of these species is not in question but their relationships and distributions would be greatly clarified by additional material. However, Dips (even large ones) can be extremely difficult to see, much less catch on a limestone substrate. I have visited all the type localities at least twice (and spent considerable time at or near that of *fowleri*), with very little success. Blair Hedges and Richard Thomas have had similar experiences with these taxa.

I agree with Cousens (1956) that Grant's subspecific arrangement of *hewardi* and *occiduus* is unrealistic. Certainly the giant *occiduus* is in serious decline from observations by Gosse (1851) and particularly from the astounding abundance in the recent Cockpit Cave remains reported by Pregill *et al.* (1992), but I think declarations that the species is extinct are premature. As noted above, dips in general and giant species in particular can be most elusive. Schwartz's discoveries of the huge *Diploglossus anelpistus* (Schwartz *et al.*, 1978) and *carraui* (Inchaústegui *et al.*, 1985) in "well-collected" areas of the Dominican Republic are but two examples. Part of the problem may be that, other than size, the characters used to distinguish *occiduus* from *hewardi* are unreliable and nobody has yet critically examined the available material of the two taxa. Variation in *C. hewardi* has also never been analyzed and there are a few specimens (USNM and CM) from an uncharacteristically high elevation (Clydesdale) in the Blue Mountains that may be a distinct species.

Gekkonidae

Although I list *Aristelliger praesignis* as a monotypic species, there is a subspecies from the Swan Islands (*A. p. nelsoni*) that has never been formally synonymized (Bauer and Russell, 1993). The genus is remarkably conservative and characters to distinguish between unquestionably distinct species are few, so analysis of intraspecific variation is meaningless using traditional meristic and morphological characters. Jamaican, Cayman, and Swan Islands *praesignis* "look" different, and molecular data might reveal them to be distinct taxa. As noted previously, Etheridge (1965) synonymized the fossil species *Aristelliger titan* (Hecht, 1951) with the Hispaniolan *A. lar*. Although we have not found any additional fossils of giant *Aristelliger* in Jamaica, the presence of an Hispaniolan species in the Pleistocene of Jamaica would be a zoogeographic anomaly. Etheridge took a com-

mendably conservative approach, but we have since prepared considerably more skeletal material of extant *Aristelliger* than was available to Etheridge and we find that other than size, there are few osteological characters that will distinguish between obviously distinct species either. Consequently, I prefer to recognize *titan* as a valid species on purely zoogeographic grounds.

The insular subspecies *Gonatodes albogularis notatus* is also problematical. Its distribution around major centers of human habitation in both Jamaica and Hispaniola smacks of an introduction, but the status of *albogularis* in northern South America and its supposed Middle American subspecies *fuscus* are also chaotic.

Jamaican *Sphaerodactylus* are in reasonably good shape, thanks to Thomas' (1975) careful review of the *argus* group. However, the very distinctive *S. gilvitorques* has never been rediscovered in Jamaica or elsewhere (Thomas and Schwartz, 1974), but there is no reason to doubt that the type actually came from Jamaica. As noted previously, Grant's (1940b) study of variation in *S. richardsoni* is suspect and the status of *S. r. gossei* needs verification. *Sphaerodactylus* sp. in my list is a diminutive taxon related to *goniorhynchus*, under study by Richard Thomas. It is a xeric, coastal species (as opposed to the mesophilic *goniorhynchus*) but with some seemingly "intermediate" populations near Kingston.

The Hispaniolan Herpetofauna: Diversity, Endemism, and Historical Perspectives, with Comments on Navassa Island

*Robert Powell*¹, *José A. Ottenwalder*,² and *Sixto J. Incháustegui*³

¹ *Department of Natural Sciences, Avila College, 11901 Wornall Road, Kansas City, Missouri 64145*

² *Conservation and Management of Biodiversity Project, United Nations Development Programme, P.O. Box 1424, Santo Domingo, República Dominicana*

³ *Grupo Jaragua, El Vergel 33, El Vergel, Santo Domingo, República Dominicana*

Introduction

Once, in a casual conversation with Albert Schwartz, he spoke of Hispaniola as an “island of islands” in reference to the myriad pockets of dramatically different habitats often separated by only a few kilometers. The discussion had focused on the tremendous diversity of the Hispaniolan herpetofauna and the difficulty experienced by those who had never been there in accepting as valid the surprisingly vast number of species and subspecies that had been described. However, the diversity is real, amazing for an island so relatively small in area (ca. 76,500 km²), and a visiting herpetologist quickly appreciates the distinctiveness of geographically proximate forms separated solely by habitat.

Geography and Physiography

The variety of amphibians and reptiles may be attributed largely to three factors: (1) the rugged and mountainous terrain of which the dominant relief features are parallel ranges that run primarily from the northwest and west in a generally easterly direction (Weil *et al.*, 1982; Lewis

and Draper, 1990), (2) a variety of satellite islands of various sizes and with exceedingly different topographies and habitats, and (3) the peculiar geological history of the island, which is unique among the Greater Antilles.

The mountainous terrain of parallel ranges (Fig. 4.1), along with an extended complex of satellite ranges, results in an inordinately diverse topography. Extensive valleys lie between the primary ranges, which are often impressive in their elevation. Satellite ranges create narrow and often broken coastal lowlands, and a lowland plain covers much of the eastern end of the island. This structural complexity, in combination with the very high elevations and resultant rainshadows, produces a juxtaposition of, among other habitat types, harsh deserts, dry scrub forests, rainforests, and high-elevation pine savannas.

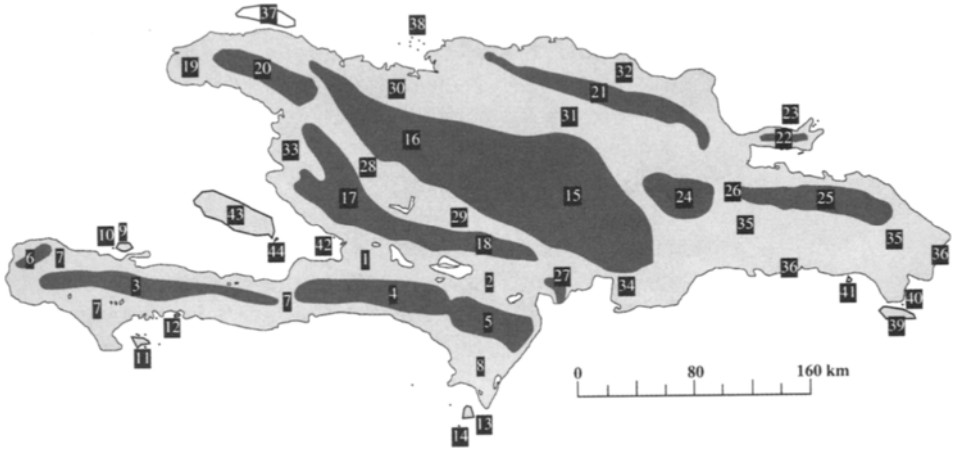


Figure 4.1. Biogeographic and physiographic features of Hispaniola (modified from Schwartz, 1980b; Schwartz and Henderson, 1984; Hedges, 1998). Darker shading indicates the approximate extent of major mountain ranges. **Key:** 1, Plaine du Cul de Sac; 2, Valle de Neiba; **South Island:** 3, Massif de la Hotte; 4, Massif de la Selle; 5, Sierra de Baoruco; 6, Monts Cartaches; 7, Peninsula de la Tiburon; 8, Peninsula de Barahona; 9, Île Grande Cayemite; 10, Île Petite Cayemite; 11, Île à Vache; 12, Île Grosse Caye; 13, Isla Beata; 14, Isla Alto Velo; **North Island:** 15, Cordillera Central; 16, Massif du Nord; 17, Montagnes du Trou-d'Eau and Montagnes Noires; 18, Sierra de Neiba; 19, Pointe de Nord Ouest; 20, Montagnes du Nord Ouest; 21, Cordillera Septentrional; 22, Sierra de Samaná; 23, Peninsula de Samaná; 24, Sierra de Yamasá; 25, Cordillera Oriental; 26, Los Haitises; 27, Sierra de Martín García; 28, Plateau Central; 29, Valle de San Juan; 30, Plaine du Nord; 31, Valle de Cibao; 32, northern coastal plain; 33, Vallée de l'Artibonite; 34, Llanos de Azua; 35, Llanura del Este; 36, Costera del Caribe; 37, Île de la Tortue; 38, Cayos Siete Hermanos; 39, Isla Saona; 40, Isla Catalinita; 41, Isla Cabrit; **Île de la Gonâve:** 43, Île de la Gonâve; 44, Île Petite Gonâve.

Important satellite islands (Fig. 4.1), many with endemic taxa, range in size from Île de la Gonâve, with an area of approximately 650 km² and a maximum elevation of over 700 m near its southeastermost point (Schwartz, 1980b), to individual cays in the Cayos Siete Hermanos, the smallest of which (Cayo Muertos) has an area measuring less than 0.02 km² and none are more than a few meters above sea level (Burns *et al.*, 1992). Other satellites of herpetological significance include Île de la Tortue, Grande Cayemite, Île à Vache, and Île Grosse Caye on the Haitian side of the main island, whereas Isla Saona, Isla Catalina, Isla Beata, and Isla Alto Velo lie on the Dominican side. In addition, Navassa Island, situated between Hispaniola and Jamaica, has a unique and apparently disproportionately large herpetofauna (Thomas, 1966a). With an area of < 4 km² (Proctor, 1959), this island supported, at least historically, eight species of reptiles, most of which demonstrate Hispaniolan affinities.

Although "Considerable controversy exists over the interpretation of the early tectonic evolution of Hispaniola" (Lewis and Draper, 1990), the peculiar history of the island lies largely in its origin as two paleoislands (Schwartz, 1978a, 1980b), although as many as four elements may have combined to form the present island (Lewis and Draper, 1990). The two major elements (the north and south paleoislands) ultimately were joined via the lateral translocation of southern Hispaniola; in effect, the south paleoisland "caught" the north island after the latter collided with the Bahama Platform (Lewis and Draper, 1990). The juncture of the paleoislands most likely occurred during the Miocene (Huebeck and Mann, 1991) and maybe as early as the Eocene (Khudoley and Meyerhoff, 1971); however, (correlated primarily with rising or falling sea levels) partial or complete separation may have occurred at intervals throughout the Pleistocene. Reef limestones deposited during the Pleistocene are currently exposed in coastal areas and in the former marine channel (Lewis and Draper, 1990). In the herpetological literature (e.g., Williams, 1961), the paleoislands have become known as the North and South islands, respectively (Fig. 4.1; subsequently used parenthetical numbers in the text correspond to those used in the figure to designate biogeographic and/or physiographic entities). The former marine strait is now a barren valley known as the Plaine de Cul-de-Sac in Haiti (1) and the Valle de Neiba in the Dominican Republic (2). Much of this valley still lies below sea level and it is characterized by four remaining large

lakes, two of which are saline: Etang Saumâtre in Haiti and nearby Lago Enriquillo in the Dominican Republic.

Further complicating the historical biogeography of Hispaniola is the sketchy pattern of emergence and submergence affecting both paleoislands (Hedges, 1996b). The present Cordillera Central, Cordillera Oriental, and possibly a portion of the La Selle – Baoruco range were emergent during the mid-Tertiary (Bowin, 1975; Lewis, 1980; Maurrassee, 1982; McLaughlin *et al.*, 1991). The remainder of the South Island probably began to emerge during the Miocene, in correlation with the collision between the paleoislands, and the entire island took shape during the Pliocene and Pleistocene (Lewis and Draper, 1990). The Sierra de Neiba and the Sierra de Martín García, along the southern shore of the North Island, apparently emerged in the late Miocene or Pliocene (McLaughlin *et al.*, 1991).

The South Island is the smaller of the two paleoislands, with an area of over 9500 km², and is much less complex than the North Island. The principal relief features constitute a series of three major mountain ranges (3-5), which run from west to east, and a smaller, independent range, the Monts Cartaches (6), lies near the northwesternmost tip of the Tiburon Peninsula (7). The westernmost of the three major ranges is the Massif de la Hotte (3), the highest peak of which (Pic Macaya) has an elevation of 2347 m and lies near the western end of the range. The central range is the Massif de la Selle (4), with several peaks exceeding 2100 m. The easternmost of these ranges is the Sierra de Baoruco (5), with a maximum elevation of 2367 m (Loma del Toro). The first two of these ranges form the backbone of the narrow Tiburon Peninsula, the width of which ranges from 30 to 68 km. Extending south of the Sierra de Baoruco for approximately 85 km is the Peninsula de Barahona (8). The Loma Gran Sabana, a limestone ridge with a maximum elevation of nearly 1100 m, vegetated by dry forest, and running essentially from north to south, effectively divides the peninsula into an extremely xeric western region and a somewhat less xeric eastern plain. Important satellite islands associated with the South Island are Île Grand Cayemite (9) and Île Petite Cayemite (10) off the northern shore of the Tiburon Peninsula, Île-à-Vache (11) and Île Grosse Caye (12) off the southern shore of the peninsula, and Isla Beata (13) and Isla Alto Velo (14) off the southern tip of the Peninsula de Barahona.

The structurally much more complex North Island has an area of about 67,700 km². The most dominant relief feature is the Cordillera

Central (15), which extends to the northwest as the Massif du Nord (16). Its highest peak, Pico Duarte, with an elevation of 3087 m, is the highest point in the entire West Indies. Essentially paralleling the southern shore of the North Island and to the south of the Cordillera Central lie the Montagnes du Trou-d'Eau and Montagnes Noires (17), which extend to the east as the Sierra de Neiba (18). The highest peak, Pico Neiba, has an elevation of 2279 m. The northwesternmost part of the island, Pointe de Nord Ouest (19), is generally dry. Uplands in that area form the Montagnes du Nord Ouest (20). To the east and along the northern coast, parallel to the Cordillera Central is the Cordillera Septentrional (21), the greatest elevation of which (Pico Diego de Ocampo) is 1250 m. Smaller, more or less isolated ranges largely to the east include the Sierra de Samaná (22) (highest point, Monte Mesa, 606 m) on the Península de Samaná (23) and the Sierra de Yamasá (24), essentially an eastern isolate of the Cordillera Central which blends into the Cordillera Oriental (25), situated to the south of the Bahía de Samaná. The maximum elevation in these ranges is 856 m at the Picos Siete Cabezas. The western extent of the Cordillera Oriental extends into the karst landscape of the Los Haitises region (26), which is sometimes considered separately as the Sierra de El Seibo. The Sierra de Martín García (27) is an eastern isolate of the Sierra de Neiba. Its highest peak is Loma del Curro, with an elevation of 1343 m. This small range is literally a mesic island surrounded by desert.

Important lowlands of the North Island include the Plateau Central (28) – Valle de San Juan (29), which lies between the Cordillera Central – Massif du Nord to the north and the Montagnes du Trou-d'Eau – Montagnes Noires – Sierra de Neiba to the South. This valley is locally mesic, but largely xeric as a result of rainshadows from the northeasterly trade winds. The Plaine du Nord (30), north of the Cordillera Central – Massif du Nord, is largely mesic near the western coast, but becomes increasingly xeric as it extends to the east and becomes the Valle de Cibao (31). Farther to the east, the area becomes gradually more mesic until the easternmost portions are among the most mesic lowlands on the Dominican side of the island. North of the Cordillera Septentrional lies the northern coastal plain (32). South of the Massif du Nord is the Vallée de l'Artibonite (33), which is mesic along the coast and increasingly more xeric in inland areas. The arid Llanos de Azua (34) lie to the south and in the rainshadow of the Cordillera Central. Much of the eastern portion of the island is flat to somewhat hilly. This Llanura del Este (35) is

moderately mesic. Craggy dogtooth limestone and pocket beaches constitute most of the *Costera del Caribe* (36).

Important satellite islands include Île de la Tortue (37) in the northwest. The Cayos Siete Hermanos (38) form an archipelago of small islets off the north central coast. Isla Saona (39) and the much smaller Isla Catalinita (40) are situated off the southeastern tip of the island, Isla Catalina (41) lies a few kilometers to the west off the southern coast, and tiny Île à Cabrit (42) is situated off the southwestern coast of the North Island.

Île de la Gonâve (43) and its minute satellite, Île Petite Gonâve (44), lie slightly closer to the North than to the South Island, but a currently submerged bank extending to the southeast provided a historical connection with both paleoislands (Schwartz, 1980b). In addition to several insular endemics, its herpetofauna is composed of a combination of both North and South island species.

Navassa Island (not shown in Fig. 4.1) is somewhat of an anomaly. Although more proximate to Hispaniola than Jamaica (approximately 65 km west of Cap de Irois, Haiti), it cannot be considered a true satellite, because of distance and faunal affinities (Thomas, 1966a). The island itself is a small limestone mass characterized by steep cliffs, about 10-15 m in height, outlining a central plateau with a maximum elevation of nearly 70 m (Proctor, 1959; Turner, 1960).

Vegetation

Because of its very diverse physiography, Hispaniola supports a diversity of floral communities. Although difficult to summarize, Hedges (1999), in describing West Indian vegetation patterns, provided a useful overview (paraphrased herein). Characterization of forest types is largely from SEA/DVS (1990), Hager and Zanoni (1993), and Hedges (1999).

Prevailing winds from the northeast carry moisture which typically precipitates on northern and eastern regions. Southern areas usually are dry. Vegetation patterns generally follow rainfall patterns. Well-developed moist forests occur on northern and eastern slopes and dry, xerophytic vegetation is found in southern regions or in rainshadows.

Before human alterations, Hispaniola was largely forested (Hedges, 1999). Lowland rainforests, with high canopies and tall buttressed trees, graded into montane rainforests on lower slopes. These forests have

high, dense canopies and occur on slopes up to about 1500 m elevation receiving more than 2 m of rainfall annually. Cloud forests occur at cloud level (usually > 1000 m). These forests have low canopies (10-15 m) and are subject to the highest humidity of any forest type. Pine forests and elfin woodlands can be found at higher elevations (from about 1000 m to >2400 m). Pine forests tend to be relatively dry, whereas elfin woodlands are characterized by windblown tangles of mossy trees and shrubs up to 5 m. Wet limestone forests, with thin soils and canopies up to 15-25 m in height, cover karst regions, usually in areas receiving less than 2 m of annual precipitation. Dry scrub forests, often associated with coastal areas, have low canopies (to 10 m), many thorny shrubs and cacti, and grow on exposed rock or very thin soil in areas receiving less than 1 m rainfall annually. These occur in the drier south and those areas shielded by mountain ranges from the prevailing winds.

Essentially all the historically widespread lowland rainforests have been destroyed, and were the first to disappear after colonization of the island (Powell and Henderson, 1996a). Most other forest types are disappearing but may still be found in patches throughout the island. Wet limestone and dry scrub forests are being destroyed at rates slower than those of most other types, mainly because of difficulties posed for human access and exploitation. Still, many areas, such as those in the vicinity of Monte Cristi, have been sufficiently altered that the subsequent secondary growth produces an impenetrable tangle of cacti and thorny shrubs. Montane rainforests remain largely on only the steepest mountain slopes, and even in regions where the forest superficially appears to be natural many components consist of cultivated species (Lenart *et al.*, 1997). Haiti has been almost entirely deforested. Some recent estimates of remaining forested areas are as low as 1% (e.g., Hedges, 1996b; see Chapter 7). The only remaining extensive tracts of natural forest in the Dominican Republic are in the Cordillera Central, but patches, some of moderate size, exist in other areas (e.g., Sierra de Baoruco).

Navassa is less xeric than most small West Indian islands, although it is comparable to the moderately xeric scrub forests on Hispaniola, and supports a relatively luxuriant flora dominated by *Ficus* trees and occasional savanna-like areas (Proctor, 1959; Turner, 1960), although the latter may be attributable to human activities (Thomas, 1966a).

Table 4.1. Amphibians of Hispaniola.^a

Species	Status	Origin or other distribution
Class Amphibia, Order Anura		
Family Bufonidae		
<i>Bufo fluviaticus</i>	Endemic	North Island
<i>Bufo fractus</i>	Endemic	North Island
<i>Bufo guentheri</i>	Endemic	North Island invader of South Island
<i>Bufo marinus</i>	Introduced	Central/South America (widely introduced)
Family Hylidae		
<i>Hyla heilprini</i>	Endemic	Islandwide
<i>Hyla pulchrilineata</i>	Endemic	Islandwide
<i>Hyla vasta</i>	Endemic	Islandwide
<i>Osteopilus dominicensis</i>	Endemic	Islandwide
Family Leptodactylidae		
<i>Eleutherodactylus abbotti</i>	Endemic	Islandwide
<i>Eleutherodactylus alcoae</i>	Endemic	South Island
<i>Eleutherodactylus amadeus</i>	Endemic	South Island
<i>Eleutherodactylus apostates</i>	Endemic	South Island
<i>Eleutherodactylus armstrongi</i>	Endemic	South Island
<i>Eleutherodactylus audanti</i>	Endemic	South Island invader of North Island
<i>E. a. audanti</i>		South Island
<i>E. a. melatrigonum</i>		North Island
<i>E. a. notidodes</i>		North Island
<i>Eleutherodactylus auriculatoides</i>	Endemic	North Island
<i>Eleutherodactylus bakeri</i>	Endemic	South Island
<i>Eleutherodactylus brevirostris</i>	Endemic	South Island
<i>Eleutherodactylus caribe</i>	Endemic	South Island
<i>Eleutherodactylus chlorophenax</i>	Endemic	South Island
<i>Eleutherodactylus corona</i>	Endemic	South Island
<i>Eleutherodactylus counouspeus</i>	Endemic	South Island
<i>Eleutherodactylus darlingtoni</i>	Endemic	South Island
<i>Eleutherodactylus dolomedes</i>	Endemic	South Island
<i>Eleutherodactylus eunaster</i>	Endemic	South Island
<i>Eleutherodactylus flavescens</i>	Endemic	North Island
<i>Eleutherodactylus fowleri</i>	Endemic	South Island
<i>Eleutherodactylus furcyensis</i>	Endemic	South Island
<i>Eleutherodactylus glandulifer</i>	Endemic	South Island
<i>Eleutherodactylus glanduliferoides</i>	Endemic	South Island
<i>Eleutherodactylus glaphycompus</i>	Endemic	South Island
<i>Eleutherodactylus grahami</i>	Endemic	North Island
<i>Eleutherodactylus haitianus</i>	Endemic	North Island
<i>Eleutherodactylus heminota</i>	Endemic	South Island

Table 4.1 cont.

<i>Eleutherodactylus hypostenor</i>	Endemic	South Island
<i>Eleutherodactylus inoptatus</i>	Endemic	Islandwide
<i>Eleutherodactylus jugans</i>	Endemic	South Island
<i>Eleutherodactylus lamprotes</i>	Endemic	South Island
<i>Eleutherodactylus leoncei</i>	Endemic	South Island
<i>Eleutherodactylus lucioi</i>	Endemic	North Island
<i>Eleutherodactylus minutus</i>	Endemic	North Island
<i>Eleutherodactylus montanus</i>	Endemic	North Island
<i>Eleutherodactylus</i> sp. "N"	Endemic	North Island
<i>Eleutherodactylus nortoni</i>	Endemic	South Island
<i>Eleutherodactylus oxyrhynchus</i>	Endemic	South Island
<i>Eleutherodactylus parabates</i>	Endemic	North Island
<i>Eleutherodactylus parapelates</i>	Endemic	South Island
<i>Eleutherodactylus patriciae</i>	Endemic	North Island
<i>Eleutherodactylus paulsoni</i>	Endemic	South Island
<i>Eleutherodactylus pictissimus</i> *	Endemic	South Island invader of North Island
<i>E. p. pictissimus</i>		South Island
<i>E. p. apantheatas</i>		Valle de Neiba / Plaine de Cul-de-Sac
<i>E. p. eremus</i>		North Island
<i>Eleutherodactylus pituinus</i>	Endemic	North Island
<i>Eleutherodactylus poolei</i>	Endemic	North Island
<i>Eleutherodactylus probolaeus</i>	Endemic	North Island
<i>Eleutherodactylus rhodesi</i>	Endemic	North Island
<i>Eleutherodactylus rufifemoralis</i>	Endemic	South Island
<i>Eleutherodactylus ruthae</i> *	Endemic	Islandwide
<i>E. r. ruthae</i>		North Island
<i>E. r. aporostegus</i>		South Island
<i>E. r. bothroboans</i>		North Island
<i>E. r. tychathrous</i>		North Island
<i>Eleutherodactylus schmidti</i>	Endemic	North Island
<i>E. s. schmidti</i>		North Island
<i>E. s. limbensis</i>		North Island
<i>E. s. rucillensis</i>		North Island
<i>Eleutherodactylus sciagraphus</i>	Endemic	South Island
<i>Eleutherodactylus semipalmatus</i>	Endemic	South Island
<i>Eleutherodactylus thorectes</i>	Endemic	South Island
<i>Eleutherodactylus ventrilineatus</i>	Endemic	South Island
<i>Eleutherodactylus warreni</i>	Endemic	North Island
<i>Eleutherodactylus weinlandi</i> *	Endemic	North Island
<i>E. w. weinlandi</i>		North Island
<i>E. w. chersonesodes</i>		North Island
<i>E. w. paralius</i>		North Island

Table 4.1 cont.

<i>Eleutherodactylus wetmorei</i>	Endemic	South Island invader of North Island
<i>E. w. wetmorei</i>		South Island
<i>E. w. ceraemerus</i>		South Island
<i>E. w. diplasius</i>		South Island
<i>E. w. sommeri</i>		North Island
<i>Leptodactylus dominicensis</i>	Endemic	North Island

*Some North Island isolates are not assigned to subspecies.

Family Ranidae

<i>Rana catesbeiana</i>	Introduced	Eastern North America (widely introduced)
-------------------------	------------	--

^aThe species list is taken from Powell *et al.* (1996a), but with the addition of a species currently being described (Hedges, 1997). Origins of endemic species are from Schwartz (1980b), except for taxa subsequently described or when the taxonomy has changed. Non-Hispaniolan distributions are from Schwartz and Henderson (1991). Please note that footnotes are inserted after each family.

Diversity and Endemism

The Hispaniolan herpetofauna consists of 65 currently recognized species of amphibians (Table 4.1) and 146 recognized species of reptiles (Table 4.2). The taxonomy used is that of Powell *et al.* (1996a), with exceptions noted in the tables. Anurans are placed in only four families and six genera. Twenty subspecies in six species are recognized. Reptiles are placed in three orders, 15 families, and 24 genera [but see comments regarding iguanian lizard families *sensu* Frost and Etheridge (1989), in Powell *et al.* (1996a) and Hedges (1996a)]. Two hundred and thirty-one subspecies in 55 species are recognized, but six of these represent situations in which a single subspecies of a polytypic species inhabits Hispaniola. Of these, one subspecies is introduced (*Anolis c. cristatellus*), three are endemic (*Trachemys stejnegeri vicina*, *Sphaerodactylus elegans punctatissimus*, and *Cyclura c. cornuta*), one clearly is native (*Gonatodes albogularis notatus*), and whether the last (*Mabuya bistrriata sloanei*) is native or introduced is uncertain.

The relative paucity of families and genera compared to the diversity of species on Hispaniola is consistent with the general pattern seen throughout the Greater Antilles, in large part because both the insular and the regional faunas are dominated by three genera: *Eleutherodactylus*, *Sphaerodactylus*, and *Anolis* (see Chapter 7). One likely cause of this preponderance of so few genera is the difficulty and subsequent rarity of

overwater colonization of an island [Hedges, 1996a, and references therein; but also see comments in Crother and Guyer (1996a,b) and in rebuttals by Hedges (1996d,e)]. This generalization may apply, even though West Indian *Eleutherodactylus* apparently arose by proto-Antillean vicariance (Hedges, 1989a,b, 1996a,b,c), and Guyer and Savage (1996a,b) have proposed an alternative scenario. Nevertheless, that ancestral Hispaniolan *Anolis* and *Sphaerodactylus* underwent spectacular radiations is apparent from evidence suggesting closer affinities among Hispaniolan forms than among more morphologically similar species from different islands (see discussion of the ecomorph concept in Chapter 7).

The possible differences in origins may also explain to some degree the obvious differences in the patterns seen among the species densities of amphibians (Fig. 4.2) and reptiles (Fig. 4.3). Amphibian distribution evidently and logically is correlated to a large degree with moisture (which, in turn, is correlated to some extent with elevation) (Hedges, 1997). Increased species densities, indicated by darker shades in Fig. 4.2, easily can be superimposed on the uplands indicated in Fig. 4.1. Historical factors, however, may complicate this issue and apparently are largely responsible for the fact that the highest elevations (e.g., the Cordillera Central) do not support a greater number of species than lower ranges on the South Island. Conversely, low, xeric regions, less hospitable to amphibians, exhibit consistently lower densities. The greatest reptilian species densities, in contrast, are in many instances associated with areas along the edges of biogeographic regions (Fig. 4.3). This ecotonal effect is most obvious where junctions between regions coincide with the greater boundary between the two paleoislands. Reptiles may be more likely than amphibians to move readily into areas bordering their historical ranges, creating concentrations of diversity along boundary zones.

The dramatically greater number of reptilian subspecies is testament to the remarkable adaptability of many reptiles when faced with the myriad habitats on Hispaniola, and testament to the pronounced differences between even geographically proximate habitats – differences even the most mobile reptiles frequently fail to overcome. The relatively few amphibian subspecies (all in the genus *Eleutherodactylus*), in contrast, speaks to the confinement of many anurans largely to moist upland regions (Fig. 4.2) (Hedges, 1999). In addition, the many ecologically versatile reptilian species with broad distributions contrasts sharply with the larger percentage of amphibian species endemic to ranges which are

small and often restricted to one type of habitat (Schwartz and Henderson, 1991; see Chapter 7).

Table 4.2. Reptiles of Hispaniola.^a

Species	Status	Origin or other distribution
CLASS REPTILIA, ORDER TESTUDINES		
Family Emydidae		
<i>Trachemys decorata</i>	Endemic	Uncertain (see discussion in Schwartz, 1980b)
<i>Trachemys stejnegeri</i>	West Indian	North Island, Bahamas, Puerto Rico (widely introduced)
<i>T. s. vicina</i>	Endemic	North Island
ORDER SQUAMATA, SUBORDER AMPHISBAENIA		
Family Amphisbaenidae		
<i>Amphisbaena caudalis</i>	Endemic	South Island
<i>Amphisbaena gonavensis</i> ^b	Endemic	Île de la Gonâve
<i>Amphisbaena innocens</i>	Endemic	South Island invader of North Island
<i>Amphisbaena hyporissor</i> ^b	Endemic	South Island
<i>A. h. hyporissor</i>		South Island
<i>A. h. leberi</i>		South Island
<i>Amphisbaena manni</i>	Endemic	North Island invader of South Island
SUBORDER LACERTILIA		
Family Anguidae		
<i>Celestus agasepsoides</i>	Endemic	South Island invader of North Island
<i>Celestus anelpistus</i>	Endemic	North Island
<i>Celestus carraui</i>	Endemic	North Island
<i>Celestus costatus</i>	Endemic	Islandwide; Navassa
<i>C. c. costatus</i>		South Island
<i>C. c. aenetergum</i>		North Island - Isla Catalinita
<i>C. c. chalcorhabdus</i>		North Island
<i>C. c. emys</i>		North Island - Île de la Tortue
<i>C. c. leionotus</i>		North Island
<i>C. c. melanchrous</i>		North Island
<i>C. c. neiba</i>		North Island
<i>C. c. nesobous</i>		South Island - Île-à-Vache

Table 4.2 cont.

<i>C. c. oreistes</i>		South Island
<i>C. c. psychonothos</i>		North Island
<i>C. c. saonae</i>		North Island - Isla Saona
<i>Celestus curtissi</i>	Endemic	North Island invader of South Island
<i>C. c. curtissi</i>		North Island, Île de la Gonâve
<i>C. c. aporus</i>		South Island
<i>C. c. diastatus</i>		North Island
<i>C. c. hylonomus</i>		North Island
<i>Celestus darlingtoni</i>	Endemic	North Island
<i>Celestus haetianus</i>	Endemic	South Island
<i>C. h. haetianus</i>		South Island
<i>C. h. mylicus</i>		South Island
<i>C. h. surdus</i>		South Island
<i>Celestus macrotus</i>	Endemic	South Island
<i>Celestus marcanoi</i>	Endemic	North Island
<i>Celestus sepsoides</i>	Endemic	Islandwide
<i>Celestus stenurus</i>	Endemic	Islandwide
<i>C. s. stenurus</i>		South Island
<i>C. s. alloeides</i>		North Island
<i>C. s. rugosus</i>		North Island
<i>C. s. weinlandi</i>		North Island, Valle de Neiba/Plaine de Cul-de-Sac
<i>Celestus warreni</i>	Endemic	North Island
Family Gekkonidae		
<i>Aristelliger expectatus</i>	Endemic	Islandwide
<i>Aristelliger lar</i>	Endemic	Islandwide
<i>Gonatodes albogularis</i>	Native	Central and South America, Cuba
<i>G. a. notatus</i>	West Indian	Jamaica, Cayman Islands
<i>Hemidactylus haitianus</i>	West Indian	Cuba, Puerto Rico
<i>Hemidactylus mabouia</i>	Native (?)	North and South islands; Africa, Neotropics
<i>Phyllodactylus wirshingi</i>	West Indian	Puerto Rico
<i>P. w. hispaniolae</i>	Endemic	North Island, Valle de Neiba
<i>P. w. sommeri</i>	Endemic	North Island

Table 4.2 cont.

<i>Sphaerodactylus altavelensis</i>	Endemic	North Island invader of South Island
<i>S. a. altavelensis</i>		South Island, Isla Alto Velo
<i>S. a. brevirostratus</i>		North and South islands, Plaine de Cul-de-Sac
<i>S. a. enriquilloensis</i>		North and South islands, Valle de Neiba
<i>S. a. lucioi</i>		North Island
<i>Sphaerodactylus armstrongi</i>	Endemic	South Island
<i>S. a. armstrongi</i>		South Island
<i>S. a. hysinephes</i>		South Island
<i>Sphaerodactylus asterulus</i>	Endemic	North Island
<i>Sphaerodactylus callocricus</i>	Endemic	North Island
<i>Sphaerodactylus cinereus</i>	Endemic	North Island invader of South Island
<i>S. c. cinereus</i>		Plaine de Cul-de-Sac
<i>S. c. stejnegeri</i>		North Island
<i>Sphaerodactylus clenchi</i>	Endemic	North Island
<i>S. c. clenchi</i>		North Island
<i>S. c. apocoptus</i>		North Island
<i>Sphaerodactylus cochranæ</i>	Endemic	North Island
<i>Sphaerodactylus copei</i>	Endemic	South Island invader of North Island
<i>S. c. copei</i>		South Island, Plaine de Cul-de-Sac
<i>S. c. astreptus</i>		South Island
<i>S. c. cataplexis</i>		South Island, Bahamas (introduced)
<i>S. c. deuterus</i>		Île de la Gonâve
<i>S. c. enochrus</i>		South Island
<i>S. c. pelates</i>		South Island
<i>S. c. picturatus</i>		South Island
<i>S. c. polyommatus</i>		South Island, Île Grande Cayemite
<i>S. c. websteri</i>		South Island
<i>Sphaerodactylus cryphius</i>	Endemic	South Island, Valle de Neiba
<i>Sphaerodactylus darlingtoni</i>	Endemic	North Island
<i>S. d. darlingtoni</i>		North Island
<i>S. d. bobilini</i>		North Island

Table 4.2 cont.

<i>S. d. mekistus</i>		North Island
<i>S. d. noblei</i>		North Island
<i>Sphaerodactylus difficilis</i>	Endemic	North Island invader of South Island
<i>S. d. difficilis</i>		North Island
<i>S. d. anthracomus</i>		South Island
<i>S. d. diolenius</i>		North Island
<i>S. d. euopter</i>		North Island - Île de la Tortue
<i>S. d. lycauges</i>		North Island
<i>S. d. peratus</i>		North Island
<i>S. d. typhlopous</i>		North Island
<i>Sphaerodactylus elasmorhynchus</i>	Endemic	South Island
<i>Sphaerodactylus elegans</i>	West Indian	Cuba
<i>S. e. punctatissimus</i>	Endemic	Islandwide, Île de la Gonâve
<i>Sphaerodactylus epiurus</i>	Endemic	North Island
<i>Sphaerodactylus ladae</i>	Endemic	North Island
<i>Sphaerodactylus lazelli</i>	Endemic	North Island
<i>Sphaerodactylus leucaster</i>	Endemic	North Island
<i>Sphaerodactylus nycteropus</i>	Endemic	South Island
<i>Sphaerodactylus ocoae</i>	Endemic	North Island
<i>Sphaerodactylus omoglaux</i>	Endemic	South Island, Plaine de Cul-de-Sac
<i>Sphaerodactylus perissodactylius</i>	Endemic	North Island
<i>Sphaerodactylus plummeri</i>	Endemic	South Island
<i>Sphaerodactylus randi</i>	Endemic	South Island
<i>S. r. randi</i>		South Island
<i>S. r. methorius</i>		South Island
<i>S. r. strahmi</i>		South Island
<i>Sphaerodactylus rhabdotus</i>	Endemic	North Island invader of South Island
<i>Sphaerodactylus samanensis</i>	Endemic	North Island
<i>Sphaerodactylus savagei</i>	Endemic	North Island
<i>S. s. savagei</i>		North Island
<i>S. s. juanilloensis</i>		North Island
<i>Sphaerodactylus shrevei</i>	Endemic	North Island
<i>Sphaerodactylus sommeri</i>	Endemic	North Island
<i>Sphaerodactylus streptophorus</i>	Endemic	South Island

Table 4.2 cont.

<i>S. s. streptophorus</i>		South Island
<i>S. s. sphenophanes</i>		South Island
<i>Sphaerodactylus thompsoni</i>	Endemic	South Island
<i>Sphaerodactylus williamsi</i>	Endemic	North Island
<i>Sphaerodactylus zygaena</i>	Endemic	South Island
Family Iguanidae		
<i>Cyclura cornuta</i>	West Indian	Navassa, Isla Mona
<i>C. c. cornuta</i>	Endemic	Islandwide
<i>Cyclura ricordii</i>	Endemic	Uncertain (see discussion in Schwartz, 1980b)
Family Polychrotidae		
<i>Anolis aliniger</i>	Endemic	North Island invader of South Island
<i>Anolis altavelensis</i>	Endemic	South Island, Isla Alto Velo
<i>Anolis alumina</i>	Endemic	South Island
<i>Anolis armouri</i>	Endemic	South Island
<i>Anolis bahorucoensis</i>	Endemic	South Island
<i>A. b. bahorucoensis</i>		South Island
<i>A. b. southerlandi</i>		South Island
<i>Anolis baleatus</i>	Endemic	North Island
<i>A. b. altager</i>		North Island
<i>A. b. caeruleolatus</i>		North Island
<i>A. b. fraudator</i>		North Island
<i>A. b. lineatacervix</i>		North Island - Isla Saona
<i>A. b. litorisilva</i>		North Island
<i>A. b. multistruppus</i>		North Island
<i>A. b. samanae</i>		North Island
<i>A. b. scelestus</i>		North Island
<i>A. b. sublimis</i>		North Island
<i>Anolis barahonae</i>	Endemic	South Island
<i>A. b. barahonae</i>		South Island
<i>A. b. albocellatus</i>		South Island
<i>A. b. inquinatus</i>		South Island
<i>A. b. mulitus</i>		South Island
<i>Anolis barboursi</i>	Endemic	South Island invader of North Island
<i>Anolis brevirostris</i>	Endemic	N. Island invader of S. Island
<i>A. b. brevirostris</i>		Valle de Neiba / Plaine de Cul-de-Sac

Table 4.2 cont.

<i>A. b. deserticola</i>		North Island
<i>A. b. wetmorei</i>		South Island
<i>Anolis caudalis</i>	Endemic	North Island invader of South Island, Île de la Gonâve
<i>Anolis chlorocyanus</i>	Endemic	North Island invader of South Island
<i>A. c. chlorocyanus</i>		North Island invader of South Island
<i>A. c. cyanostictus</i>		North Island
<i>Anolis christophei</i>	Endemic	North Island
<i>Anolis coelestinus</i>	Endemic	South Island
<i>A. c. coelestinus</i>		South Island
<i>A. c. demissus</i>		South Island, Île Grande Cayemite
<i>A. c. pecuarius</i>		South Island, Île-à-Vache
<i>Anolis cristatellus</i>	Introduced	Puerto Rico Bank
<i>A. c. cristatellus</i>		Puerto Rico
<i>Anolis cybotes</i>	Endemic	Islandwide
<i>A. c. cybotes</i>		Islandwide
<i>A. c. doris</i>		Île de la Gonâve
<i>A. c. ravifaux</i>		North Island - Isla Saona, Isla Catalinita (?)
<i>Anolis darlingtoni</i>	Endemic	South Island
<i>Anolis distichus</i>	West Indian	Islandwide; Bahamas (introduced elsewhere)
<i>A. d. aurifer</i>	Endemic	South Island
<i>A. d. dominicensis</i>	Endemic	North and South islands
<i>A. d. favillarum</i>	Endemic	South Island
<i>A. d. ignigularis</i>	Endemic	North Island (introduced in Florida)
<i>A. d. juliae</i>	Endemic	South Island - Île-à-Vache
<i>A. d. patruelis</i>	Endemic	South Island - Île Grande Cayemite
<i>A. d. properus</i>	Endemic	North Island
<i>A. d. ravitergum</i>	Endemic	North Island
<i>A. d. sejunctus</i>	Endemic	North Island - Isla Saona
<i>A. d. suppar</i>	Endemic	South Island
<i>A. d. tostus</i>	Endemic	North Island - Isla Catalina

Table 4.2 cont.

<i>A. d. vinosus</i>	Endemic	South Island
<i>Anolis dolichocephalus</i>	Endemic	South Island
<i>A. d. dolichocephalus</i>		South Island
<i>A. d. portusalus</i>		South Island
<i>A. d. sarmenticola</i>		South Island
<i>Anolis etheridgei</i>	Endemic	North Island
<i>Anolis eugenegrahami</i>	Endemic	North Island
<i>Anolis fowleri</i>	Endemic	North Island
<i>Anolis haetianus</i>	Endemic	South Island
<i>Anolis hendersoni</i>	Endemic	South Island
<i>A. h. hendersoni</i>		South Island
<i>A. h. ravidormitans</i>		South Island
<i>Anolis insolitus</i>	Endemic	North Island
<i>Anolis koopmani</i>	Endemic	South Island
<i>Anolis longitibialis</i>	Endemic	South Island
<i>A. l. longitibialis</i>		South Island - Isla Beata
<i>A. l. specuum</i>		South Island
<i>Anolis marcanoii</i>	Endemic	North Island
<i>Anolis marron</i>	Endemic	South Island
<i>Anolis monticola</i>	Endemic	South Island
<i>A. m. monticola</i>		South Island
<i>A. m. quadrisartus</i>		South Island
<i>Anolis olssoni</i>	Endemic	North Island invader of South Island
<i>A. o. olssoni</i>		North Island
<i>A. o. alienus</i>		North Island
<i>A. o. dominigensis</i>		North Island
<i>A. o. extensus</i>		Islandwide
<i>A. o. ferrugicauda</i>		North Island
<i>A. o. insulanus</i>		Île de la Gonâve
<i>A. o. montivagus</i>		North Island
<i>A. o. palloris</i>		South Island
<i>Anolis placidus</i>	Endemic	North Island
<i>Anolis porcatus</i>	Introduced	Cuba
<i>Anolis ricordii</i>	Endemic	Islandwide
<i>A. r. ricordii</i>		North and South islands
<i>A. r. leberi</i>		South Island
<i>A. r. subsolanus</i>		South Island
<i>A. r. viculus</i>		South Island

Table 4.2 cont.

<i>Anolis rimarum</i>	Endemic	North Island
<i>Anolis rupinae</i>	Endemic	South Island
<i>Anolis semilineatus</i>	Endemic	Islandwide
<i>Anolis sheplani</i>	Endemic	South Island
<i>Anolis shrevei</i>	Endemic	North Island
<i>Anolis singularis</i>	Endemic	South Island invader of North Island (?), Île de la Gonâve
<i>Anolis strahmi</i>	Endemic	South Island
<i>A. s. strahmi</i>		South Island
<i>A. s. abditus</i>		South Island
<i>Anolis websteri</i>	Endemic	North Island
<i>Anolis whitemani</i>	Endemic	North Island
<i>A. w. whitemani</i>		North Island, Valle de Neiba/Plaine de Cul-de-Sac
<i>A. w. breslini</i>		North Island
<i>A. w. lapidosus</i>		North Island
Family Scincidae		
<i>Mabuya lineolata</i>	Endemic	North Island
<i>Mabuya bistrriata</i> (?)	Native (?)	Northern South America
<i>M. b. sloanei</i>	Native (?)	North Island (one record); Puerto Rico Bank, Turks and Caicos islands, Jamaica
Family Teiidae		
<i>Ameiva chrysoleaema</i>	Endemic	North Island invader of South Island
<i>A. c. chrysoleaema</i>		Islandwide
<i>A. c. abbotti</i>		South Island - Isla Beata
<i>A. c. alacris</i>		North Island
<i>A. c. boekeri</i>		North Island
<i>A. c. defensor</i>		North Island
<i>A. c. evulsa</i>		South Island - Grosse Caye
<i>A. c. ficta</i>		South Island
<i>A. c. jacta</i>		North Island
<i>A. c. parvioris</i>		North Island
<i>A. c. procax</i>		North Island
<i>A. c. quadrijugis</i>		South Island

Table 4.2 cont.

<i>A. c. regularis</i>		North Island
<i>A. c. richardthomasi</i>		North Island - Isla Saona
<i>A. c. secessa</i>		Île de la Gonâve
<i>A. c. umbratilis</i>		Valle de Neiba
<i>A. c. woodi</i>		North Island - Île de la Tortue
<i>Ameiva leberi</i>	Endemic	South Island
<i>Ameiva lineolata</i>	Endemic	North Island invader of South Island
<i>A. l. lineolata</i>		Islandwide
<i>A. l. beatensis</i>		South Island - Isla Beata
<i>A. l. meracula</i>		North Island
<i>A. l. perplicata</i>		North Island
<i>A. l. privigna</i>		South Island
<i>A. l. semota</i>		North Island - Isla Catalina
<i>Ameiva taeniura</i>	Endemic	Islandwide; Navassa Island
<i>A. t. taeniura</i>		South Island
<i>A. t. aequorea</i>		South Island - Île-à-Vache
<i>A. t. azuae</i>		North Island
<i>A. t. barbouri</i>		North Island, Île de la Gonâve
<i>A. t. ignobilis</i>		North Island
<i>A. t. meyerabichi</i>		North Island
<i>A. t. pentamerinthus</i>		South Island - Île Grande Cayemite
<i>A. t. regnatrix</i>		South Island
<i>A. t. rosamondae</i>		North Island - Isla Saona
<i>A. t. tofacea</i>		North Island
<i>A. t. vafra</i>		North Island
<i>A. t. varica</i>		South Island
<i>A. t. vulcanalis</i>		North and South islands
Family Tropicuridae		
<i>Leiocephalus barahonensis</i>	Endemic	South Island
<i>L. b. barahonensis</i>		South Island
<i>L. b. aureus</i>		South Island
<i>L. b. beatanus</i>		South Island - Isla Beata
<i>L. b. oxygaster</i>		South Island

Table 4.2 cont.

<i>Leiocephalus endomychus</i>	Endemic	North Island
<i>Leiocephalus lunatus</i>	Endemic	North Island
<i>Leiocephalus melanochlorus</i>	Endemic	South Island
<i>L. m. melanochlorus</i>		South Island
<i>L. m. hypsistus</i>		South Island
<i>Leiocephalus personatus</i>	Endemic	North Island invader of South Island
<i>L. p. personatus</i>		South Island
<i>L. p. actites</i>		North Island
<i>L. p. agraulus</i>		North Island
<i>L. p. budeni</i>		North Island
<i>L. p. elatoprosopon</i>		South Island
<i>L. p. mentalis</i>		North Island
<i>L. p. poikilometes</i>		North Island
<i>L. p. pyrrholaemus</i>		North Island
<i>L. p. scalaris</i>		North Island
<i>L. p. socioensis</i>		North Island
<i>L. p. tarachodes</i>		North Island
<i>L. p. trujilloensis</i>		North Island
<i>Leiocephalus pratensis</i>	Endemic	North Island
<i>L. p. pratensis</i>		North Island
<i>L. p. chimarus</i>		North Island - Île-à-Cabrit
<i>Leiocephalus rhutidira</i>	Endemic	North Island
<i>Leiocephalus schreibersii</i>	Endemic	North Island invader of South Island
<i>L. s. schreibersii</i>		Islandwide
<i>L. s. nesomorus</i>		North Island - Île de la Tortue
<i>Leiocephalus semilineatus</i>	Endemic	North Island invader of South Island
<i>Leiocephalus vinculum</i>	Endemic	Islandwide (?)
<i>L. v. vinculum</i>		Île de la Gonâve
<i>L. v. altavelensis</i>		South Island - Isla Alto Velo

SUBORDER SERPENTES

Family Boidae

<i>Epicrates fordii</i>	Endemic	North Island invader of South Island
-------------------------	---------	--------------------------------------

Table 4.2 cont.

<i>E. f. fordii</i>		North Island, Valle de Neiba/ Plaine de Cul-de-Sac, Île de la Gonâve
<i>E. f. agametus</i>		North Island
<i>E. f. manototus</i>		North Island - Île-à-Cabrit
<i>Epicrates gracilis</i>	Endemic	Islandwide
<i>E. g. gracilis</i>		North Island
<i>E. g. hapalus</i>		South Island
<i>Epicrates striatus</i>	West Indian	Islandwide; Bahamas
<i>E. s. striatus</i>	Endemic	North and South Islands, Île de la Gonâve
<i>E. s. exagistus</i>	Endemic	South Island
<i>E. s. warreni</i>	Endemic	North Island - Île de la Tortue
Family Colubridae		
<i>Alsophis anomalus</i>	Endemic	Islandwide
<i>Alsophis melanichmus</i>	Endemic	Islandwide
<i>Antillophis parvifrons</i>	Endemic (?)	Islandwide; Bahamas (status unknown)
<i>A. p. parvifrons</i>		South Island
<i>A. p. alleni</i>		Île de la Gonâve, Île Petite Gonâve
<i>A. p. lincolni</i>		South Island
<i>A. p. niger</i>		North Island
<i>A. p. paraniger</i>		North Island
<i>A. p. protenus</i>		North and South islands
<i>A. p. rosamondae</i>		South Island - Île-à-Vache
<i>A. p. stygius</i>		North Island - Isla Saona
<i>A. p. tortuganus</i>		North Island - Île de la Tortue
<i>Darlingtonia haetiana</i>	Endemic	South Island
<i>D. h. haetiana</i>		South Island
<i>D. h. perfector</i>		South Island
<i>D. h. vaticinata</i>		South Island
<i>Hypsirhynchus ferox</i>	Endemic	Islandwide
<i>H. f. ferox</i>		North and South islands
<i>H. f. exedruss</i>		North Island - Isla Saona

Table 4.2 cont.

<i>H. f. paracrousis</i>		Île de la Gonâve
<i>H. f. scalaris</i>		South Island
<i>Ialtris agyrtes</i>	Endemic	Islandwide
<i>Ialtris dorsalis</i>	Endemic	Islandwide
<i>Ialtris parishi</i>	Endemic	Islandwide
<i>Uromacer catesbyi</i>	Endemic	Islandwide
<i>U. c. catesbyi</i>		South Island
<i>U. c. cereolineatus</i>		South Island - Île Grande Cayemite, Île Petite Cayemite (?)
<i>U. c. frondicolor</i>		Île de la Gonâve
<i>U. c. hariolatus</i>		North Island
<i>U. c. incháusteguii</i>		North Island - Isla Saona
<i>U. c. insulaevaccarum</i>		South Island - Île-à- Vache
<i>U. c. pampineus</i>		North Island
<i>Uromacer frenatus</i>	Endemic	South Island invader of North Island, Île de la Gonâve
<i>U. f. frenatus</i>		South Island
<i>U. f. chlorauges</i>		South Island invader of North Island
<i>U. f. dorsalis</i>		Île de la Gonâve
<i>U. f. wetmorei</i>		South Island - Isla Beata
<i>Uromacer oxyrhynchus</i>	Endemic	North Island invader of South Island
Family Leptotyphlopidae		
<i>Leptotyphlops asbolepis</i>	Endemic	North Island
<i>Leptotyphlops calypso</i>	Endemic	North Island
<i>Leptotyphlops leptipilepta</i>	Endemic	South Island
<i>Leptotyphlops pyrites</i>	Endemic	South Island
Family Tropicodphiidae		
<i>Tropicodphis haetianus</i>	West Indian	Islandwide; Cuba, Jamaica
<i>T. h. haetianus</i>	Endemic (?)	North Island invader of South Island, Île de la Gonâve; Cuba
<i>T. h. hemerus</i>	Endemic	North Island
<i>T. h. tiburonensis</i>	Endemic	South Island
Family Typhlopidae		
<i>Typhlops capitulatus</i>	Endemic	South Island, Plaine de Cul- de-Sac

Table 4.2 cont.

<i>Typhlops gonavensis</i>	Endemic	Île de la Gonâve
<i>Typhlops hectus</i>	Endemic	South Island invader of North Island
<i>Typhlops pusillus</i>	Endemic	North Island invader of South Island, Île de la Gonâve
<i>Typhlops schwartzi</i>	Endemic	North Island
<i>Typhlops sulcatus</i>	Endemic	South Island invader of North Island, Île de la Gonâve
<i>Typhlops syntherus</i>	Endemic	South Island
<i>Typhlops tetrathyreus</i>	Endemic	Uncertain, Plaine de Cul-de-Sac
<i>Typhlops titanops</i>	Endemic	South Island
ORDER CROCODYLIA		
Family Crocodylidae		
<i>Crocodylus acutus</i>	Native	Islandwide (historically); Neotropics

^aThe species list is taken from Powell *et al.* (1996), except where noted. Origins of endemic species are from Schwartz (1980b), except for taxa subsequently described or when the taxonomy has changed. Non-Hispaniolan distributions are from Schwartz and Henderson (1991). Endemism of subspecies is noted only for non-Hispaniolan endemic species.

^bBased on allopatry, clear and consistent meristic and morphological differences (Thomas, 1965a), and comments in Thomas (1965c) and Powell (1992b, 1993), we recognize the main island populations as a species distinct from that of Île de la Gonâve.

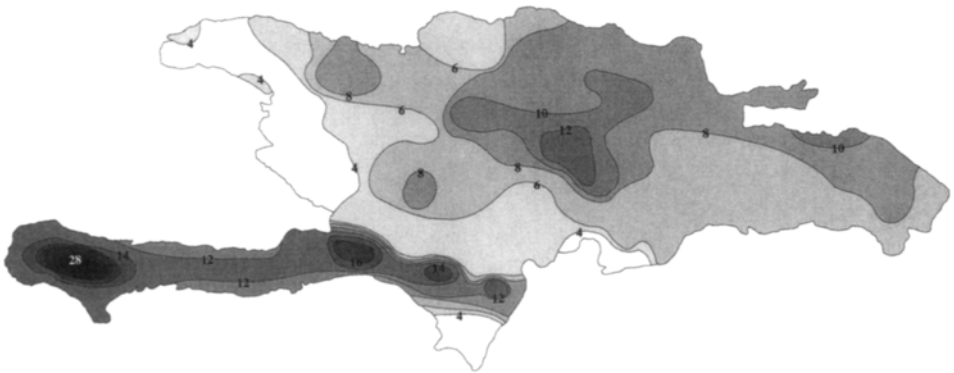


Figure 4.2. Amphibian species density on Hispaniola. Species density is the number of overlapping species distributions. This entity is approximately equal to the number of sympatric species but often is greater than the number of syntopic species. Contours with intervals of two species were determined by outlining and superimposing ranges based on records in Schwartz and Henderson (1991) with modifications by Hedges (1999). This figure is modified from Fig. 12 in Hedges (1999).

However, another consideration relative to the large number of reptilian (and a few amphibian) subspecies was raised by Powell (1993). Reptiles tend to have more morphological characters (e.g., scales) capable of being examined for variation (molecular data suggest that amphibians may actually demonstrate as much geographic variability as reptiles; S. B. Hedges, personal communication).

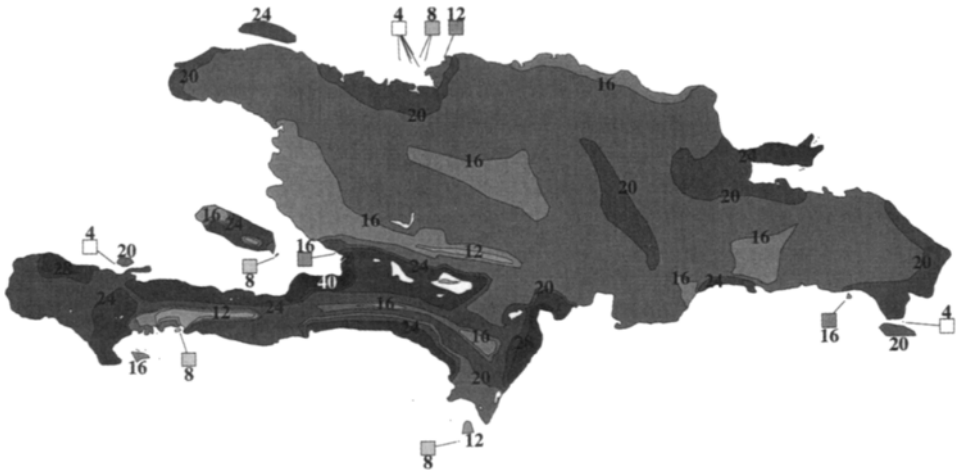


Figure 4.3. Squamate reptilian species density on Hispaniola. Species density is the number of overlapping species distributions. This entity is approximately equal to the number of sympatric species but often is greater than the number of syntopic species. Numbers are conservative estimates because species for which few records exist were not assumed to be present in all intervening areas of apparently suitable habitat. Contours with intervals of four species were determined by outlining and superimposing ranges based on records in Schwartz and Henderson (1991).

Based on an application of the evolutionary species concept (Wiley, 1978; Frost and Hillis, 1990), Powell (1993) identified subspecies with ranges that are apparently allopatric with those of conspecifics and formal diagnoses of morphological distinctiveness had been published (e.g., *Ameiva chrysoleama* in Fig. 4.4). Largely without reexamination and

further study of each situation, he recognized 43 Hispaniolan species which might actually represent complexes containing as many as 92 specific taxa. Five of the identified subspecies have been elevated to full species status: *Leiocephalus endomychus* by Pregill (1992), *Bufo (Peltophryne) fractus*, *Aristelliger expectatus*, and *Hemidactylus haitianus* by Powell *et al.* (1996a), and *Amphisbaena hyporissor* (this chapter).



Figure 4.4. Ranges of representative species on Hispaniola. *Ameiva chrysoleama* (light gray) has a broad distribution on both North and South paleoislands. This species was considered a North Island invader of the South Island by Schwartz (1980b). *Sphaerodactylus samanensis* (medium gray) is a North Island endemic (Schwartz, 1980b) with a limited range restricted to the Los Haitises region along the southern shore of the Bahía de Samaná. *Eleutherodactylus chlorophenax* (black) is a South Island endemic (Schwartz, 1980b) with a limited range restricted to the western portion of the Massif de la Hotte. Range outlines are modified from Schell *et al.* (1993), Schwartz and Henderson (1991), and Cunningham *et al.* (1998), respectively.

If only a fraction of the additional taxonomic questions survive the scrutiny of detailed morphological and biochemical evaluations, the number of subspecies elevated to full species status will increase substantially the species richness of Hispaniola – with considerable implications for the development of conservation programs on the island.

The following is a summary of the distribution and presumed endemism of currently recognized taxa of Hispaniolan amphibians and reptiles. Most of the attributions regarding paleoisland origins are from Schwartz (1980b) and those based on subsequently described taxa are based largely on distribution. However, S. B. Hedges (personal commu-

nication) pointed out the difficulty in making such decisions based solely on distributional data. Obviously, the best information would come from phylogenetic studies, but sufficiently detailed analyses are lacking for many Hispaniolan forms. In addition, a valid alternative to "invasion" hypotheses in some instances (e.g., *Celestus agasepsoides* or *Typhlops pusillus*) is vicariance (see Crother and Guyer, 1996a,b): A continuous range in the Pleistocene, for example, may have been split by a change in the intervening habitat (e.g., drying or inundation of the Valle de Neiba/Plaine du Cul de Sac). As a result, the assignment of biogeographic histories presented here should be treated as preliminary until more detailed phylogenies become available for a greater number of the Hispaniolan taxa. Distributional data included in Schwartz and Henderson (1991) are not cited repeatedly; only subsequently published references are listed.

Amphibians

Of the 65 recognized species, one is an introduced ranid, four are bufonids, four are hylids, and the remainder are leptodactylids (Table 4.1). *Rana catesbeiana* has been introduced, probably intended as a food source, from North America. This species is widely distributed across the island wherever suitable ponds or slow-moving streams are available for breeding.

Of the four bufonids, one (*Bufo marinus*) was introduced to the island and has spread widely in lowland areas, especially in agricultural regions, but also in the arid Valle de Neiba/Plaine de Cul-de-Sac. The other three species are endemic to Hispaniola and all specifically to the North Island. *Bufo fluviaticus* has a restricted range in the Valle de Cibao, and *B. fractus* has a similarly restricted range in the eastern Dominican lowlands. Both areas are moderately to strongly mesic. *Bufo guentheri*, in contrast, is widely distributed across the North Island and has invaded the South Island via the Valle de Neiba/Plaine de Cul-de-Sac. Unlike the other two endemic species, *B. guentheri* is largely xerophilic. Hedges (1996a) recognized a single West Indian lineage (the *Bufo peltocephalus* group) of native bufonids, centered in Cuba, to which all endemic Hispaniolan species are assigned.

All of the four hylid species are Hispaniolan endemics and have islandwide distributions (i.e., North or South island origins cannot be determined). *Osteopilus dominicensis* is found in a variety of habitats, apparently not even requiring an arboraceous area (although these frogs are

usually arboreal). Very adaptable and obviously capable of effectively utilizing severely altered habitats, females often deposit ova in artificial pools and fountains and adults are not infrequently edificarian in urban areas and resorts. The three species of *Hyla* are almost always associated with mesic broadleaf forests, although *H. pulchrrilineata* is known to occupy forest remnants and "situations much disturbed by man" (Schwartz and Henderson, 1991). *Hyla vasta* is most common in gallery forests near streams, and *H. heilprini* nearly always is associated with rapidly flowing or torrential waters. Hedges (1996a) recognized two West Indian lineages of native hylids (*Hyla heilprini* and *Osteopilus*, to which he assigned the other Hispaniolan species currently placed in the genus *Hyla*).

The lone Hispaniolan leptodactylid in the genus *Leptodactylus*, *L. dominicensis*, is a North Island endemic restricted to the southern shore of the Bahía de Samaná. Hedges (1996a) placed this species with the closely related Puerto Rican Bank endemic, *L. albilabris*, into a single lineage.

All other Hispaniolan leptodactylids are assigned to the genus *Eleutherodactylus*. Of the 55 species, only three have islandwide distributions, 17 are North Island endemics (none of which have invaded the South Island), and 36 are endemic to the South Island. Of the latter, three have invaded the North Island and all three have endemic North Island subspecies plus one subspecies restricted to the Valle de Neiba/Plaine de Cul-de-Sac.

When all Hispaniolan amphibians are considered, two species (*Bufo marinus* and *Rana catesbeiana*) were introduced and 63 (96.9%) are endemic (Hedges, 1997). Of the endemic forms, seven (11.1%) have islandwide distributions and their origins cannot be attributed to either of the paleoislands, 20 (31.7%) are North Island endemics, and 36 (57.1%) are South Island endemics. Only one North Island form has invaded the South Island, whereas three South Island forms have established populations on the North Island. Of the 20 subspecies, all of which are *Eleutherodactylus*, 10 are in three species with South Island origins, six in two species with North Island origins, and four in a single species with an islandwide distribution.

When distribution is related to physiographic provinces, high levels of endemism are also evident. By correlating the biogeographic regions of Henderson and Schwartz (1984a) with distributions of native amphibians, a picture of regional endemism develops (Table 4.3). Of the 62

Table 4.3 cont.

Species	Biogeographic Region															
	MH	MS	SB	PB	CS	GG	SN	PC	PN	CC	VC	NH	ER	LA	MG	TP
<i>Eleutherodactylus hypostenor</i>	X	X	X													
<i>Eleutherodactylus inoptatus</i>	X	X	X				X			X	X	X	X			X
<i>Eleutherodactylus jugans</i>			X													
<i>Eleutherodactylus lamprotes</i>	X															
<i>Eleutherodactylus leoncei</i>			X													
<i>Eleutherodactylus lucioi</i>									X							
<i>Eleutherodactylus minutus</i>										X						
<i>Eleutherodactylus montanus</i>										X						
<i>Eleutherodactylus</i> sp. "N"							X									
<i>Eleutherodactylus nortoni</i>			X													
<i>Eleutherodactylus oxyrhynchus</i>	X	X														
<i>Eleutherodactylus parabates</i>								X								
<i>Eleutherodactylus parapelates</i>	X															
<i>Eleutherodactylus patriciae</i>										X						
<i>Eleutherodactylus paulsoni</i>	X															X
<i>Eleutherodactylus pictissimus</i>	X	X	X		X									X		X
<i>Eleutherodactylus pituinus</i>										X						
<i>Eleutherodactylus poolei</i>										X						
<i>Eleutherodactylus probolaeus</i>													X			
<i>Eleutherodactylus rhodesi</i>								X								
<i>Eleutherodactylus ruffifemoralis</i>			X													
<i>Eleutherodactylus ruthae</i>	X	X	X				X			X		X	X			
<i>Eleutherodactylus schmidti</i>										X						
<i>Eleutherodactylus sciagraphus</i>	X															
<i>Eleutherodactylus semipalmatus</i>	X	X														
<i>Eleutherodactylus thorectes</i>	X															
<i>Eleutherodactylus ventrilineatus</i>	X															
<i>Eleutherodactylus weinlandi</i>							X			X		X	X			
<i>Eleutherodactylus wetmorei</i>	X	X								X						
<i>Leptodactylus dominicensis</i>												X				

*These regions are less detailed than those discussed in the text and illustrated in Figure 1 (see comments in Henderson and Schwartz, 1984a:3). However, the use of these regions, with subdivided areas combined and no consideration of satellite islands (see text), provides an adequate appreciation of regional endemism among Haitian and Dominican frogs. *Eleutherodactylus warreni* is omitted from this table because it is endemic and restricted to Île de la Tortue. Key to abbreviations: MH = Massif de la Hotte, MS = Massif de la Selle, SB = Sierra de Baoruco, PB = Península de Barahona, CS = Valle de Neiba/Plaine de Cul-de-Sac, GG = Golfe de la Gonâve, SN = Sierra de Neiba and Montagnes du Trou-d'Eau, PC = Plateau Central, PN = Presqu'île du Nord-Ouest, CC = Cordillera Central and Massif du Nord, VC = Valle de Cibao, NH = Plaine du Nord (north Haitian littoral plain extending eastward to include much of the northern and eastern República Dominicana), ER = eastern República Dominicana (largely mesic coastal plain), LA = Llanos de Azua, MG = Sierra Martín García, TP = Tiburon Peninsula.

species considered (see caption to Table 4.3), 42 (67.7%) are restricted to a single one of the 16 regions considered. Another seven (11.3%) are found only in two regions, and in all instances the regions are adjacent and similar in habitat (e.g., the Massif de la Hotte and Massif de la Selle). Six (9.7%) additional species are limited to three to five regions, all of which are either contiguous and similar in habitat, consist of upland areas and adjacent lowlands, and/or represent very similar situations (e.g., uplands) separated by apparently uninhabitable intervening areas of distinctly different habitats. Only seven species (11.3%) are found in as many as six regions and in areas with varying ecological conditions, and only two (*Osteopilus dominicensis* and *Eleutherodactylus abbotti*) are found in as many as three-fourths (≥ 12) of the regions considered. The hylid frogs were by far the most geographically diverse group, with no species found in fewer than five regions, and *O. dominicensis* in all but one.

Few amphibians have established populations on satellite islands. *Eleutherodactylus alcoae* occurs on Isla Beata, *E. inoptatus* has been found on Île de la Tortue, and *E. pictissimus* is known from Île-à-Vache. *Eleutherodactylus warreni* is endemic and restricted to Île de la Tortue and is the only satellite island endemic amphibian of Hispaniola. *Osteopilus dominicensis*, the only species found on more than one satellite, possibly a tribute to its ecological versatility, has populations on Île de la Gonâve, Île-à-Vache, Île Grande Cayemite, Île de la Tortue, and Isla Saona. No amphibians are known to occur on Navassa Island.

Reptiles

Of the 146 recognized species, one is a crocodylian, two are emydid turtles, and the remainder are squamates, with all three traditionally recognized suborders represented (Table 4.2). *Crocodylus acutus* has a broad historical distribution which includes southern Florida, the western Greater Antilles (Cuba, Hispaniola, and Jamaica), and the Neotropical mainland south along both coasts of southern México and Central and South America to northern Perú on the Pacific side and to Venezuela on the Caribbean (Atlantic) side (Groombridge, 1987). On Hispaniola, native populations once were found along many coastal areas as well as in a number of inland streams and lakes (Thorbjarnarson, 1988). Current populations appear to be restricted to saline Etang Saumatre, hypersaline Lago Enriquillo, and a few geographically proximate freshwater streams and lakes in the Valle de Neiba/Plaine de Cul-de-Sac (Schubert and

Santana, 1996), although evidence supporting the extirpation of crocodiles from the Montecristi region is inconclusive and further (nocturnal) surveys will be required to confirm this conclusion.

Hispaniola is the only West Indian island on which two species of native emydid turtles are known to occur (Seidel and Incháustegui Miranda, 1984). Both species are native to Hispaniola, and *Trachemys decorata* is endemic. Most populations are found in the Valle de Neiba/Plaine de Cul-de-Sac, but others are known from the Vallée de l'Artibonite on the North Island and from the western Tiburon Peninsula of the South Island. Schwartz (1980b) discussed the difficulty of tracing the origin of this species to either the North or South paleoisland. *Trachemys stejnegeri*, on the other hand, is endemic to the Bahamas, Puerto Rico, and Hispaniola (Seidel, 1996), with the populations on the latter constituting a subspecies (*T. s. vicina*) widespread on and endemic to the North Island. Hedges (1996a) placed all West Indian species of *Trachemys* in a single lineage, presumably of North American origin.

Amphisbaenian squamates are represented on Hispaniola by five species (Table 4.2), all of which are endemic. The only North Island endemic (*Amphisbaena manni*) has invaded the South Island, but only one (*A. innocens*) of the three South Island endemics contains North Island populations. The only currently recognized subspecies are of *A. hyporissor*, and the ranges of both are limited to the South Island. The fifth species (*A. gonavensis*) is restricted to Île de la Gonâve. In addition to the latter, *A. caudalis* is found only on satellite islands, Île Grande Cayemite and Presqu'île de Baradères off the northern coast of the Tiburon Peninsula (Haiti). Other insular populations include those of *A. hyporissor leberii* (Isla Beata), *A. innocens* (Île Grande Cayemite) and *A. manni* (Île de la Tortue); none are endemic to these islands. Hedges (1996a) placed all West Indian species in the genus *Amphisbaena* (including Cuban species formerly in *Cadea*) in a single lineage.

Lacertilian squamates constitute the most abundant and diverse of the herpetofaunal elements on Hispaniola. Seven families are represented: anguids by 12 species and 22 subspecies in one genus, gekkonids by 38 species and 41 subspecies in five genera, iguanids (*sensu strictu*) by two species and one subspecies in one genus, polychrotids by 42 species and 66 subspecies in one genus, scincids by two species and one subspecies in one genus, teiids by four species and 35 subspecies in one one, and tropidurids by 10 species and 24 subspecies in one genus.

All 12 species of Anguidae are placed in the genus *Celestus* and all are Hispaniolan endemics. Three are islandwide forms, six are North Island endemics, and three appear to have originated on the South Island. Two (*C. costatus* and *C. stenurus*) of the three islandwide species contain both North and South Island endemic subspecies. Of the North Island species, one (*C. curtissi*) is represented on the South Island by a South Island endemic subspecies. Of the South Island endemic species, one (*C. agasepsoides*) has invaded the North Island. All three subspecies of any South Island endemic, all in *C. haitianus*, are restricted to the South Island. Four subspecies of *C. costatus* are endemic to satellite islands (Île-à-Vache, Île de la Tortue, Isla Catalinita, Isla Saona), and another is endemic to Navassa Island (see below). All were listed by Powell (1993) as possible candidates for full-species recognition. Another population is known from the Cayos Siete Hermanos (Burns *et al.*, 1992). *Celestus curtissi* contains populations on Île de la Gonâve, Île de la Tortue, Isla Catalina, and Isla Saona. None represent endemic subspecies, although the populations on the two latter islands are unassigned subspecifically. Satellite island populations of *C. stenurus* are known to occur on Île-à-Vache, Île Grande Cayemite, Île-à-Cabrit, and the Cayos Siete Hermanos. Although the population on Île Grande Cayemite is unassigned, none appear to represent endemic subspecies. Hedges (1996a) assigned all Hispaniolan species in *Celestus*, including those formerly placed in *Diploglossus*, *Sauresia*, and *Wetmorena*, to a single West Indian lineage. In contrast, Savage and Lips (1993), resurrecting the classification which places species with claw sheaths in *Diploglossus* and those lacking sheaths in *Celestus*, retained *Sauresia* and *Wetmorena* and noted their affinity with *Diploglossus*. On this basis, Hispaniolan anguids represent two distinct lineages.

The 38 species and 41 subspecies of Hispaniolan gekkonids are placed in five genera. Both species of *Aristelliger* are islandwide endemics, and neither is polytypic. Populations of *A. expectatus* are known from Île de la Gonâve, Île Grande Cayemite, Île-à-Cabrit, Île de la Tortue, Isla Beata, Isla Alto Velo, and Cayo Pisaje. The only known satellite populations of *A. lar* are on Isla Catalina and the Cayos Siete Hermanos (but see Burns *et al.*, 1992 and Powell and Parmelee, 1992). Both are in an Antillean *Aristelliger* lineage (Hedges, 1996a). *Gonatodes albugularis* is widely distributed through Central and northern South America and also is native to the western Antilles. All Hispaniolan populations, including those on Île de la Gonâve and Île-à-Cabrit, are

assigned to the West Indian endemic subspecies, *G. a. notatus*, which also is found on Jamaica and the Cayman Islands. Hedges (1996a) recognized a single West Indian lineage. Two species in the genus *Hemidactylus* are found on Hispaniola, each representing separate West Indian lineages (Hedges, 1996a). *Hemidactylus haitianus* is a West Indian endemic also found on Cuba and Puerto Rico. The only known Hispaniolan satellite island population is on Isla Saona. *Hemidactylus mabouia* is widespread in the Neotropics, where it probably is native (but see Kluge, 1969 and Vanzolini, 1978). On Hispaniola, the species is known only from the area around Port-au-Prince and the extreme northeastern Dominican Republic. Because this species is known as an extremely successful colonizer (Meshaka *et al.*, 1994a,b,c), these populations are presumed to be native. *Phyllodactylus wirshingi* is a West Indian endemic found on Puerto Rico and Hispaniola. Populations on the latter constitute two North Island endemic subspecies, one of which (*P. w. hispaniolae*) has extended its range into the Valle de Neiba. Hedges (1996a) recognized this species as a distinct West Indian lineage.

The remaining 32 species and 38 subspecies of Hispaniolan gekkonids are all in the genus *Sphaerodactylus*. Of the 32 species, all are native and all but one are endemic. That species, *S. elegans* (known also from Cuba), is represented by an endemic subspecies (*S. e. punctatissimus*) which has an islandwide distributional pattern and also contains populations on Île de la Gonâve and Île Grande Cayemite. Twenty species are North Island endemics, four of which have invaded the South Island, and two of these contain South Island endemic subspecies. Eleven species are South Island endemics, of these, three (*S. copei*, *S. cryphius*, *S. omoglaux*) have invaded the Valle de Neiba/Plaine de Cul-de-Sac and one (*S. copei*) has extended its range onto the North Island and to Île de la Gonâve. *Sphaerodactylus altavelensis* has satellite island populations on Île-à-Cabrit and on Isla Alta Velo, the latter constituting an endemic (n nominate) subspecies (although Powell *et al.*, 1989 tentatively attributed a specimen taken on the Península de Barahona to this subspecies). Populations of *S. copei* are on Île de la Gonâve, Île Grande Cayemite, and Île-à-Vache, with the first two recognized as satellite island endemic subspecies. *Sphaerodactylus difficilis euopter* is endemic to Île de la Tortue, but the population of *S. d. typhlopous* reported from the Cayos Siete Hermanos is not restricted to the islands. *Sphaerodactylus randi strahmi* has been found on, but is not restricted to Cayo Pisaje. Similarly, *S. savagei juanilloensis* is known from Isla Saona but

also is found on the main island. However, the populations of *S. savagei* on Isla Catalinita is unassigned subspecifically. A population of *S. thompsoni* occurs on Isla Beata. All of the satellite island endemics were listed by Powell (1993) as deserving consideration as full species. Hedges (1996a) placed all West Indian *Sphaerodactylus* in a single lineage.

Both iguanid species are in the West Indian endemic genus *Cyclura*. *Cyclura ricordii* is restricted to Hispaniola, but its paleoisland affinities are uncertain (Schwartz, 1980b). *Cyclura cornuta* is an islandwide species on Hispaniola, where it is represented by the nominate subspecies. Endemic subspecies also occur on Isla Mona between Hispaniola and Puerto Rico and on Navassa Island. Powell (1993) suggested that these forms may be deserving of full-species status. These and all West Indian species of *Cyclura* represent a single lineage (Hedges, 1996a).

All 42 species and 66 subspecies of polychrotid lizards on Hispaniola are currently placed in the genus *Anolis*. Two species are introduced: *A. cristatellus* from the Puerto Rico Bank and *A. porcatum* from Cuba (Hispaniolan populations of both are assigned to nominate subspecies). The former is well established in and around La Romana and the latter in Santo Domingo. In both instances, their distributions end with the onset of largely native vegetation, in which they are replaced, respectively, by the ecologically similar endemics, *A. cybotes* and *A. chlorocyanus*. *Anolis distichus*, also known from the Bahamas and introduced into Florida, is native. With 12 endemic Hispaniolan subspecies on both the North and South islands, *A. distichus* is considered an islandwide form. Of the 39 endemic species, *Anolis cybotes*, *A. ricordii*, and *A. semilineatus* also are islandwide species. Seventeen species are recognized as North Island endemics. Of these, five (*A. aliniger*, *A. brevirostris*, *A. caudalis*, *A. chlorocyanus*, and *A. olssoni*) have invaded the South Island and another (*A. whitemani*) the Valle de Neiba/Plaine de Cul-de-Sac. Two of the South Island invaders (*A. brevirostris* and *A. olssoni*) contain subspecies endemic to that area. Of the 19 South Island endemics, only two (*A. barbouri* and *A. singularis*) have invaded the North Island and the latter is questionable; Schwartz and Henderson (1991) noted that North Island specimens of *A. singularis* from the Sierra de Martín García may represent a related form. *Anolis coelestinus* also is known from the Plaine de Cul-de-Sac. *Anolis altavelensis* is endemic to Isla Alto Velo. Satellite island endemic subspecies are known from Île de la Gonâve (*A. cybotes doris* and *A. olssoni insulanus*), Île Grand Cayemite [*A. coelestinus de-*

missus and *A. distichus patruelis* (which also may refer to the population on Île Petit Cayemite)], Île-à-Vache (*A. coelestinus pecuarius* and *A. distichus juliae*), Isla Beata (*A. l. longitibialis*), Isla Catalina (*A. distichus tostus*), and Isla Saona [*A. baleatus lineatacervix* and *A. cybotes ravifaux* (which also may refer to the population on Isla Catalinita), *A. distichus sejunctus*]. Non-endemic satellite island populations occur on Île de la Gonâve (*A. caudalis*, *A. chlorocyanus*, and *A. singularis*), Île de la Petit Gonâve (*A. caudalis*), Île de la Tortue (*A. chlorocyanus*, *A. cybotes*, *A. distichus*, and *A. semilineatus*), Île-à-Cabrit (*A. brevirostris*, *A. caudalis*, *A. cybotes*, *A. olssoni*, and *A. semilineatus*), Île Grande Cayemite (*A. cybotes* and *A. semilineatus*), Île-à-Vache (*A. cybotes*), Isla Catalina (*A. cybotes* and *A. semilineatus*), Isla Beata (*A. brevirostris*), Isla Saona (*A. chlorocyanus*), and the Cayos Siete Hermanos (*A. distichus*, *A. whitemani*) — although Schwartz had planned to recognize the population of *A. cybotes* on Isla Catalina at the subspecific level (R. W. Henderson, personal communication). Powell (1993) listed 16 satellite island endemics and other diagnosed allopatric subspecies as possibly deserving full-species status. Hedges (1996a) placed all West Indian species of *Anolis* (including those until recently assigned to *Chamaeleolis* and *Chamaelinorops*) in a single lineage.

Both Hispaniolan skinks are in the genus *Mabuya*. *Mabuya lineolata* is endemic and restricted to the North Island. This species constitutes a single West Indian lineage (Hedges, 1996a). The status of the other familial representative is much less certain. *Mabuya bistrriata* is found widely throughout the West Indies, with the Hispaniolan population from near Santo Domingo assigned to *M. b. sloanei*, presumably the same subspecies found on Jamaica, the Turks and Caicos Islands, and the Puerto Rico Bank. The broad distribution of this lizard argues for consideration of the Hispaniolan specimens as being native, but a distribution centered on a cosmopolitan port suggests an introduction. Hedges (1996a) considered all of the West Indian populations of this lizard to be part of another single lineage.

All four species of teiids are in the genus *Ameiva*, and all are endemic to Hispaniola. *Ameiva taeniura* is an islandwide form, with endemic subspecies on both the North and South islands. Satellite island endemic subspecies occur on Île de la Gonâve, Île Grande Cayemite, Île-à-Vache, and Isla Saona (although that on Île de la Gonâve has also been recorded from the adjacent North Island). Additional insular populations are on Île Petite Cayemite and Isla Catalina. *Ameiva chrysolaema* and *A.*

lineolata are North Island endemics that have invaded the South Island. Both contain satellite island endemic subspecies: *A. chrysolaema* on Île de la Gonâve, Île de la Tortue, Grosse Caye, Isla Beata, and Isla Saona; and *A. lineolata* on Isla Beata and Isla Catalina. Other satellite island populations are known from Île-à-Cabrit (both species) and from Isla Catalina and the Cayos Siete Hermanos (*A. chrysolaema*). *Ameiva leberi* is a South Island endemic without any satellite island populations. Powell (1993) listed 21 satellite island endemics and other diagnosed allopatric subspecies as possibly deserving full-species status. All West Indian species of *Ameiva*, except populations of *A. ameiva* in the western Caribbean and on the southern Lesser Antilles, are assigned to a single lineage (Hedges, 1996a).

Five families of ophidian squamates are represented on Hispaniola: boids by three species and eight subspecies in one genus, colubrids by 11 species and 27 subspecies in six genera, leptotyphlopids by four species in one genus, tropidophiids by a single genus with one species and three subspecies, and typhlopids by nine species in one genus. No species of snakes have been introduced.

All three native species of Boidae are in the genus *Epicrates*. *Epicrates fordii* is a North Island endemic which has invaded the South Island through the Valle de Neiba/Plaine de Cul-de-Sac. Satellite island populations occur on Île de la Gonâve, Isla Saona, and Île-à-Cabrit, with the latter considered a subspecies endemic to this tiny coastal island. *Epicrates gracilis* and *E. striatus* are both islandwide species, with subspecies endemic to both paleoislands. The former is restricted to Hispaniola, but *E. striatus* also is native to the Bahamas, where an additional five subspecies occur. Hispaniolan satellite island populations of *E. striatus* are known from Isla Saona, Île de la Gonâve, Île Grande Cayemite, Île-à-Vache, and Île de la Tortue, but only the last is considered an endemic subspecies (listed by Powell, 1993 as possibly deserving of full-species status). Hedges (1996a) followed Kluge (1989) in concluding that all West Indian species of *Epicrates* form a monophyletic group.

The 11 species and 27 subspecies of Hispaniolan colubrids currently are placed in six genera. Both species of *Alsophis* are islandwide endemics. The few scattered localities for both species (only two for *A. melanichnus*) make any further estimation of origin impossible. Satellite island populations of *A. anomalus* are known from Île de la Tortue and Isla Beata. *Antillophis parvifrons* is another islandwide Hispaniolan spe-

cies, although this snake also has been recorded from Little Inagua Island in the Bahamas (the status of this population is unknown; Schwartz and Henderson, 1991). Of the nine endemic subspecies, four are associated solely with satellite islands: *A. p. alleni* on Île de la Gonâve and Île Petit Gonâve, *A. p. rosamondae* on Île-à-Vache, *A. p. stygius* on Isla Saona, and *A. p. tortuganus* on Île de la Tortue. Powell (1993) considered these to be candidates for recognition as full species. Other satellite island populations are known from Île Grande Cayemite and Grosse Caye (*A. p. parvifrons*), Isla Beata (*A. p. lincolni*), and Isla Catalina (*A. p. parani-ger*). South Island endemic *Darlingtonia haetiana* contains three subspecies, all restricted to the uplands of the three sequential ranges of the South Island. *Hypsirhynchus ferox* is another islandwide form, with subspecies on both paleoislands and two composed of satellite island populations: *H. f. exedrus* on Isla Saona and *H. f. paracrousis* on Île de la Gonâve. All three species of *Ialtris* also are considered islandwide forms, with the dearth of locality records for *I. agyrtes* (N = 3) and *I. parishi* (N = 2) precluding a more accurate picture of their origins. Both *I. dorsalis* and *I. parishi* are known from Île de la Tortue, and *I. dorsalis* also is known to occur on Île de la Gonâve and Île-à-Vache. The three species of *Uromacer* paint a more disparate picture. *Uromacer catesbyi* is an islandwide form with seven subspecies, five of which are restricted to satellite islands: *U. c. cereolineatus* to Île Grand Cayemite and presumably Île Petit Cayemite, *U. c. frondicolor* to Île de la Gonâve, *U. c. Inchâusteguii* to Isla Saona, *U. c. insulaevaccarum* to Île-à-Vache, and *U. c. scandax* to Île de la Tortue. All may represent full species (Powell, 1993). *Uromacer frenatus* is a South Island endemic which has invaded the North Island (Smith and Powell, 1991). Endemic satellite populations are known from Île de la Gonâve (*U. f. dorsalis*) and Isla Beata (*U. f. wetmorei*). The monotypic species, *U. oxyrhynchus*, was considered by Schwartz (1980b) to be a North Island endemic which has invaded the South Island. Satellite island populations are known from Île de la Tortue, Isla Catalina, and Isla Saona. Powell (1993) included all satellite island endemic subspecies in his list of taxa deserving full-species status. Hedges (1996a) placed all the Hispaniolan colubrids into a single West Indian alsophine lineage.

The four species of *Leptotyphlops* are all endemic to Hispaniola, two (*L. asbolepis* and *L. calypso*) to the North Island and two (*L. leptipilepta* and *L. pyrites*) to the South Island. Hedges (1996a) followed Thomas *et*

al. (1985) in placing all four Hispaniolan species with *L. bilineata* from the Lesser Antilles into a single West Indian radiation.

The lone Hispaniolan tropidophiid, *Tropidophis haetianus*, also is known from Cuba (site of a single record of the otherwise Hispaniolan nominate subspecies) and Jamaica, with three endemic subspecies. On Hispaniola, the species is an islandwide form containing subspecies endemic to both paleoislands. Satellite island populations are known from Île de la Gonâve and Île de la Tortue. All West Indian tropidophiids were placed in a single lineage by Hedges (1996a).

All nine species in the genus *Typhlops* are endemic to Hispaniola. Two (*T. pusillus* and *T. schwartzi*) are considered to be North Island endemics, and one of these (*T. pusillus*) has invaded the South Island and also is found on Île de la Gonâve. Five species are South Island endemics; of these, two (*T. syntherus* and *T. titanops*) are restricted to the South Island, two (*T. hectus* and *T. sulcatus*) have invaded the North Island, with *T. sulcatus* also known from Île de la Gonâve and from Navassa Island. The fifth (*T. capitulatus*) also has been recorded from the Plaine de Cul-de-Sac. *Typhlops gonavensis* is endemic to Île de la Gonâve, and the status of *T. tetrathyreus*, the range of which is largely restricted to the Plaine de Cul-de-Sac, is uncertain. In addition to those forms on Île de la Gonâve, satellite island populations are known from Île Grande Cayemite (*T. hectus*, *T. pusillus*, and *T. sulcatus*), Île de la Tortue (*T. pusillus*), Isla Alto Velo (*T. sulcatus*), and Isla Saona (*T. pusillus*). Hedges (1996a), citing Thomas (1989), placed all West Indian species of *Typhlops* in a single lineage.

When all 146 species of Hispaniolan reptiles are considered, only two species (*Anolis cristatellus* and *A. porcatus*) clearly were introduced [the status of *Mabuya bistrriata* and *Hemidactylus mabouia* is unknown, but we will follow Schwartz and Henderson (1991) in treating the Hispaniolan populations of these species as native]. Of the native species, 14 have native populations elsewhere (other than Navassa Island) and, of these, eight are represented on Hispaniola by endemic subspecies. Conservatively treating only these eight and those restricted to Hispaniola as endemics, 94.5% of Hispaniolan reptiles are endemic. Of the endemic forms, 24 (17.4%) have islandwide distributions, 62 (44.9%) are endemic to the North Island, 48 (34.8%) are endemic to the South Island, two (1.4%) are endemic and restricted to Île de la Gonâve, and the status of two (1.4%) is uncertain. Of the North Island species, 19 (30.6%) have invaded the South Island and another the Valle de Neiba/Plaine de Cul-

Table 4.4 cont.

<i>Typhlops</i>	X	X	X
<i>tetrathyreus</i>			
<i>Typhlops titanops</i>	X	X	

* These regions are less detailed than those discussed in the text and illustrated in Figure 1 (see comments in Henderson and Schwartz, 1984a:3). However, the use of these regions, with subdivided areas combined, provides an adequate appreciation of regional endemism among Haitian and Dominican amphisbaenians, lizards, and snakes. Key to abbreviations: MH = Massif de la Hotte, MS = Massif de la Selle, SB = Sierra de Baoruco, PB = Península de Barahona, CS = Valle de Neiba/Plaine de Cul-de-Sac, GG = Golfe de la Gonâve, SN = Sierra de Neiba and Montagnes du Trou-d'Eau, PC = Plateau Central, PN = Presqu'île du Nord-Ouest, CC = Cordillera Central and Massif du Nord, VC = Valle de Cibao, NH = Plaine du Nord (north Haitian littoral plain extending eastward to include much of the northern and eastern República Dominicana), ER = eastern República Dominicana (largely mesic coastal plain), LA = Llanos de Azua, MG = Sierra Martín García, TP = Tiburon Peninsula, IT = Île de la Tortue, IG = Île de la Gonâve, PG = Île de la Petit Gonâve, GC = Île Grande Cayemite, IP = Île Petit Cayemite, IV = Île-à-Vache, GR = Grosse Caye, IC = Île-à-Cabrit, SH = Cayos Siete Hermanos, CA = Isla Catalinita, IS = Isla Saona, CT = Isla Catalina, IB = Isla Beata, AV = Isla Alto Velo.

de-Sac, whereas only eight (16.6%) of the South Island endemics have invaded the North Island and another three are established widely in the Valle de Neiba/Plaine de Cul-de-Sac. Of the 228 endemic subspecies, 79 are in 15 polytypic species classified as islandwide forms, 95 are in 20 polytypic North Island endemic species, and 54 are in 18 South Island species.

In relating reptilian distributions to the biogeographic provinces, including satellite islands, defined in Henderson and Schwartz (1984a) (Table 4.4), regional endemism is less evident than that in amphibians (i.e., more reptilian than amphibian species are found in more regions or, conversely, fewer are restricted to one or very few regions). Only 141 native species were considered; the aquatic emydid turtles and one crocodylian were omitted, because any correlation with physiographic region would merely represent a coincidental juxtaposition of lakes, rivers, or coastal areas with the defined terrestrial regions. Of the species meeting the criteria for inclusion, 51 (36.2%) are restricted to only one of the 16 main island and 14 satellite island regions included in the analysis. Another 23 (16.3%) are found in only two, usually proximate and ecologically similar regions, and 25 more (17.7%) are found in three to five regions, which, as for amphibians, either are contiguous or similar in habitat. Forty-two (29.8%) species are found in six or more regions, but only two (*Anolis cybotes* and *Antillophis parvifrons*) are found in more than three-fourths (≥ 23) of the regions considered. However, if the satellite islands are disregarded, ten species are found in at least three-

fourths (≥ 12) of the main-island regions, with specimens of both *Anolis cybotes* and *Antillophis parvifrons* having been recorded from all 16. All familial groups except two contained at least one species found in as many as nine regions. *Mabuya lineolata* (Scincidae) was found in three regions (*M. bistrriata* in only one) and only one species of *Leptotyphlops* was found in more than one region (*L. pyrites* in two).

Navassa Island

The known herpetofauna of Navassa Island (Table 4.5) consists of eight squamate species in two suborders, seven families, and eight genera. However, four have not been collected recently and may be extinct. Six other species have been recorded from the island but were regarded by Thomas (1966a) as not having been originally collected there (i.e., erroneous collection data had been recorded). *Aristelliger cochranae* and *Celestus costatus* have distinct Hispaniolan affinities. The former is closely related to and until recently was thought to be conspecific with Hispaniolan *A. expectatus*. The latter is a Navassan endemic subspecies (*C. c. badius*) of a widely distributed Hispaniolan form. *Sphaerodactylus becki* and *Anolis longiceps* are both endemic to Navassa Island. The former appears to be most closely related to Hispaniolan *S. copei*, and was, in fact, placed in the *S. copei* species group by Hass (1996). *Anolis longiceps* appears to be most closely related to *A. maynardi* of the Cayman Islands, and Schmidt (1921b) stated that *A. longiceps* is "apparently related to the Cuban *Anolis porcatus* through *Anolis maynardi*." Of the species for which recent specimens are not available, the iguanid *Cyclura cornuta* was represented on Navassa by an endemic subspecies [*C. c. onchiopsis* (= *C. c. nigerrima*)]. *Leiocephalus eremitus* has been confused with *L. melanochlorus* (Schmidt, 1921b), but Thomas (1966a) seemed confident that a distinct Navassan form once existed. *Typhlops sulcatus* is endemic to Hispaniola, whereas Navassan *Tropidophis melanurus* apparently represents an endemic subspecies (*T. m. bucculentus*) of a snake species endemic to Cuba. Thus, of the eight species rather reliably attributed to Navassa Island, six have Hispaniolan and two have Cuban affinities, although one of the latter may have reached Navassa via the Cayman Islands.

Historical Perspectives

Although pre-Columbian native Americans undoubtedly exploited some members of the herpetofauna (at least iguanas and turtles) as food (see Powell and Henderson, 1996b), the first historical references to any Hispaniolan amphibian or reptiles are those of Christopher Columbus (cited in Major, 1870; Morison, 1963), who made frequent references to iguanas and turtles in his writings. Also, Gonzalo de Oviedo (cited in Stoudemire, 1959), an early European chronicler, described an encounter with a very large (undoubtedly exaggerated) snake "at the foote of the Mountains called Pedernales." However, a formal taxonomy of Hispaniolan species was many years in coming, and realized only with the publication of *The Herpetology of Hispaniola* by Doris M. Cochran in 1941. Arguably the most comprehensive herpetofaunal account in the entire West Indian region, this book provided a foundation to which every subsequent authority has first turned when addressing questions pertaining to the amphibians and reptiles of the island.

Amphibians

Taxonomic authorities for Hispaniolan amphibians are listed in Table 4.6. The widespread toad, *Bufo marinus*, introduced on Hispaniola, was described by Linnaeus (1758), albeit not from a West Indian specimen. The next Hispaniolan species to be described, *Rana catesbeiana* by George Shaw (1802), also is introduced and was described from non-West Indian (North American) material. During the first half of the nineteenth century, European scientists, soliciting materials from any source, placed a large number of West Indian species into a systematic order (Powell and Henderson, 1996b). However, only two Hispaniolan endemics were included: *Osteopilus dominicensis*, described by Johann J. von Tschudi (1838), and *Eleutherodactylus oxyrhynchus*, described by Constant Duméril and Gabriel Bibron (1841) in their massive *Érpetologie Générale*. All other species and all subspecies occurring on the island have been described subsequently by North Americans. Edward Drinker Cope (1869, 1871) described two endemic hylids (*Hyla pulchilineata* and *H. vasta*) before the turn of the century, but 59 of the 65 Hispaniolan species have been described since 1900. Thomas Barbour (1914, 1942) described three species of *Eleutherodactylus*, and another was described by Karl P. Schmidt (1919). However, Hispaniolan anuran

Table 4.5. Reptiles of Navassa Island (from Thomas, 1966b).

Species	Status	Faunal Association
CLASS REPTILIA ORDER SQUAMATA		
SUBORDER LACERTILIA		
Family Anguidae		
<i>Celestus costatus</i>	Native	Hispaniola
<i>C. c. badius</i>	Endemic	
Family Gekkonidae		
<i>Aristelliger cochranae</i>	Endemic	Hispaniola (<i>A. expectatus</i>)
<i>Sphaerodactylus becki</i>	Endemic	Hispaniola (<i>S. copei</i> group)
Family Iguanidae		
<i>Cyclura cornuta</i> *	Native	Hispaniola
<i>C. c. nigerrima</i> *	Endemic	
Family Polychrotidae		
<i>Anolis longiceps</i>	Endemic	Cuba (<i>A. porcatus</i>) via Cayman Islands (<i>A. maynardi</i>)
Family Tropiduridae		
<i>Leiocephalus eremitus</i> *	Endemic	Hispaniola (<i>L. melanochlorus</i>)
SUBORDER SERPENTES		
Family Tropidophiidae		
<i>Tropidophis melanurus</i> *	Native	Cuba
<i>T. m. bucculentus</i> *	Endemic	
Family Typhlopidae		
<i>Typhlops sulcatus</i> *	Native	Hispaniola

*Not collected recently; these species may be extinct or the Navassan populations may be extirpated.

taxonomy in the first half of the twentieth century was dominated by Cochran, G. Kingsley Noble, and Benjamin Shreve. Shreve named four species of *Eleutherodactylus* in a 1936 paper and three more with Ernest E. Williams in 1963. Cochran (1923a,b, 1932d, 1934b, 1935, 1937, 1938a, 1939, 1941) named 11 species and one currently recognized sub-species, including the sole representative of the genus *Leptodactylus* and an endemic toad. Noble, alone (1923a) and with William G. Hassler (1933), named eight species of *Eleutherodactylus*.

However, an appreciation for the tremendous diversity of Hispaniolan amphibians was left for the second half the century and was largely due to the prodigious efforts of Albert Schwartz. In a series of papers, Schwartz (1964a, 1964/1965, 1965a, c,d, 1966a, 1968c, 1971b, 1972, 1973c, 1976a,b, 1977a, 1979b, 1980a) named 21 species and 12 subspecies of *Eleutherodactylus* (including *E. neodreptus*, recently placed in the synonymy of *E. audanti*; Hedges, 1996d) and two species of endemic toads. In doing so, he was solely responsible for over one third of the known species and over 85% of the subspecies on the island. A pure taxonomist, Schwartz nevertheless sought to know the animals he studied in the field as well as in the more traditional context of museum specimens. As a result, his taxa often are identified by habitat use as well as by morphological and meristic characters. Although some workers have criticized the vast number of subspecies described by Schwartz, even the most ardent critics cannot deny that our understanding of geographic variation in the taxa addressed by him has been immeasurably enhanced.

Table 4.6. Taxonomic Authorities for Hispaniolan Amphibians.^a

Authority	Number of taxa described	Percentages
Barbour (1914–1942)	3	4.6 (0) 3.8
Cochran (1923–1941)	11 (1)	16.9(7.1) 15.2
Cope (1869–1871)	2	3.1 (0) 2.5
Duméril and Bibron	1	1.5 (0) 1.3
Hedges (1988)	1	1.5 (0) 1.3
Hedges and Thomas (1987–1992)	4	6.2 (0) 5.1
Hedges, Thomas, and Franz (1987)	1	1.5 (0) 1.3
Linnaeus (1758)	1	1.5 (0) 1.3
Lynn (1958)	0 (1)	0 (7.1) 1.3
Noble (1923)	6	9.2 (0) 7.6
Noble and Hassler (1933)	2	3.1 (0) 2.5
Schmidt (1919)	1	1.5 (0) 1.3
Schwartz (1964–1980)	23 (12)	35.4 (85.7) 44.3
Shaw (1802)	1	1.5 (0) 1.3
Shreve (1936)	4	6.2 (0) 5.1
Shreve and Williams (1963)	3	4.6 (0) 3.8
Tschudi (1838)	1	1.5 (0) 1.3

^a The range of publication dates is given in parentheses after each name. The number of currently recognized species described is followed by the number of subspecies currently accepted as valid (n nominate subspecies are not included). Percentages are for species (subspecies) and for all currently recognized taxa at these levels.

William G. Lynn named another subspecies in 1958, but nearly all of the recent taxonomic work by scientists other than Schwartz has and is being done by S. Blair Hedges and colleagues. Hedges alone (1988), in three papers with Richard Thomas (1987, 1992a,b), and with Thomas and Richard Franz (1987) has named six species of *Eleutherodactylus*, and another (*E. sp.* "N" in Table 4.1) is being described at this time; based on an extrapolation of a graph illustrating the rate of species discoveries in the West Indies (Hedges, 1996a) and new discoveries not yet reported (S. B. Hedges, personal communication), more taxa apparently remain to be discovered.

In addition to the introduced *Bufo marinus*, Hispaniola supports three endemic toads. *Bufo guentheri* was described by Cochran in 1941, and *B. fluviaticus* and *B. fracta* by Schwartz in 1972 (the latter as a subspecies of *B. guentheri*). Charles S. Sonnini and Pierre-André Latreille (1801) and François-Marie Daudin (1802b) had described *B. gutturosus* and *B. strumosus*, respectively, but Cochran (1941) quoted a manuscript note by Leonhard Stejneger which stated that their inadequate descriptions were sufficient to preclude application of those names to any Hispaniolan (or West Indian) species. Nevertheless, both were subsequently and inappropriately used by Gravenhorst (1829) and Günther (1858 [1859]) in published references to Hispaniolan specimens. Shreve and Williams (1963) assigned *B. guentheri* to the "Caribbean section" of New World bufonids, and Pregill (1981a) resurrected the genus *Peltophryne* to accommodate these species. The latter generally was accepted, although Schwartz and Henderson (1988) emended the name to *Peltaphryne* (see comment in Powell, 1992a). Based on molecular evidence, Hedges *et al.* (1992b) questioned the validity of *Peltophryne*, and placed the Hispaniolan species (and other Antillean congeners) back into the genus *Bufo*. The latter action was formally acknowledged in Powell *et al.* (1996a).

Four hylid frogs, all endemic, occur on Hispaniola. *Hypsiboas* (= *Osteopilus*) *dominicensis* was described by Tschudi (1838) and *Hyla pulchrrilineata* and *H. vasta* by Cope (1869, 1871, respectively) during the nineteenth century. *Hyla heilprini* was described by Noble (1923a). Dunn (1926a) proposed that the Hispaniolan and Jamaican species represented independent radiations, but that they were "allied." Trueb and Tyler (1973), using morphological evidence, concluded that at least six independent invasions of the West Indies occurred, all from South America. Hedges *et al.* (1992b) and Hedges (1996a), based on molecular

evidence, supported as "most plausible" the South American origin of these frogs, but differ from earlier workers by suggesting that all West Indian hylids (except *H. heilprini*) represent a monophyletic group, that is to take the generic name *Osteopilus*). In contrast, Anderson (1996) used karyological data to strongly support the monophyly of *Osteopilus*, *sensu strictu*.

All other Hispaniolan anurans are in the family Leptodactylidae, and all but one are in the genus *Eleutherodactylus*. Heyer (1978) synonymized *Leptodactylus dominicensis*, originally described by Cochran (1923b), with *L. albilabris*, a closely related frog endemic to the Puerto Rico Bank. However, Schwartz and Henderson (1991) and Powell *et al.* (1996a) continued to recognize as full species these apparently closely related forms.

Tschudi (1838) named *Cornufer unicolor*, but this name has subsequently been placed in the synonymy of *Platymantis*, an Australasian ranid. Zweifel (1967) showed that Tschudi's lectotype was not a ranid and suggested it was an older name for *Eleutherodactylus inoptatus*, which had been described subsequently by Barbour (1914) as *Leptodactylus inoptatus*. This taxon was reassigned to *Eleutherodactylus* by Schmidt (1921a). To prevent confusion, Zweifel sought to suppress Tschudi's names, and the Commission of Zoological Nomenclature (Anonymous, 1978) removed *Cornufer* and *unicolor* from further consideration for eleutherodactyline frogs.

The first name coined for and still assigned to a Hispaniolan *Eleutherodactylus* is attributable to Duméril and Bibron (1841), who described *Hylodes oxyrhynchus*. *Eleutherodactylus ventrilineatus* was originally assigned to *Leptodactylus* by Shreve (1936). Substitute names were assigned by Barbour (1942; *E. haitianus* for *E. intermedius* Cochran 1941, not *E. intermedius* Barbour and Shreve 1937) and by Cochran (1937; *E. jugans* for *Leptodactylus* [= *Eleutherodactylus*] *darlingtoni* Cochran 1935:372, not *E. darlingtoni* Cochran 1935:368). In general and despite the confusion noted previously, the taxonomy of Hispaniolan forms in this speciose genus has been remarkably stable. Disagreements have centered largely around relationships within the genus and among subgenera.

Dunn (1926a) developed a taxonomic framework for the genus *Eleutherodactylus*. Various refined in the intervening years, that initial plan was expressed more fully by Schwartz (e.g., 1965a) and Joglar (1989) on the basis of morphological data and largely supported by elec-

trophoretic data presented in Hedges (1989a). Three of five currently recognized subgenera are represented in the West Indies, and all include Hispaniolan forms (Hedges, 1989a). The subgenus *Euhyas* is endemic to the western Greater Antilles (Cuba, Jamaica, and Hispaniola) and Isla Mona, although species have been introduced into México and the United States. All six species assigned to *Pelorius* are restricted to Hispaniola. The subgenus *Eleutherodactylus* occurs throughout the West Indies (except Jamaica) and on the mainland, although Hedges (1996a) cited evidence that the West Indian species form a monophyletic group. While recommending the retention of the currently assigned names until more information is available, Lynch (1996), based on cranial morphology, disagreed with Hedges (1989a) and Joglar (1989) in assignment of species to groups and in the suggested monophyly of *Pelorius*.

Reptiles

Taxonomic authorities for Hispaniolan reptiles are listed in Table 4.7. The earliest formal description of a Hispaniolan species was of *Lacerta* (= *Cyclura*) *cornuta* by Pierre-Joseph Bonnaterre in 1789. In retrospect, this was the first of a number of Hispaniolan species described during an era in which museum specialists, primarily European, solicited specimens from every continent in order to impose a taxonomic order on the reptiles of the world. A list of subsequent authorities reads almost like a "who's who in nineteenth century herpetology." François-Marie Daudin (1803) described *Scincus sloanii* (= *Mabuya bistrriata sloanei*). Georges Cuvier (1807) described *Crocodilus* [sic] *acutus* from "St.-Dominique." Johann von Spix described *Scincus* (= *Mabuya*) *bistriatus* in 1825. *Hemidactylus mabouia* was described from a Lesser Antillean specimen by Alexandre Moreau de Jonnés in 1818. In 1830, Johann Wagler described *Sphaerodactylus cinereus*. William S. MacLeay (1834) described *Sphaeriodactylus* [sic] *elegans*, albeit from a Cuban specimen. Constant Duméril and Gabriel Bibron (1836, 1837, 1839) and Duméril *et al.* (1854) included in their *Érpetologie Générale* seven species and one subspecies still recognized today. *Dendrophis* (= *Uromacer*) *catesbyi* and *Pristinotus* (= *Leiocephalus*) *schreibersii* were added to the growing list of Hispaniolan reptiles by Hermann Schlegel (1837) and Johann L. C. Gravenhorst (1837 [1838]), respectively. John Edward Gray (1840, 1852) described *Anolis porcatus* (from Cuba) and *Sauresia sepsoides*. Johann Gustav Fischer (1856, 1888) described *Homalochilus* (= *Epicrates*) *striatus*, *Chilabothrus* (= *Epicrates*) *gracilis*,

Table 4.7. Taxonomic Authorities for Hispaniolan Reptiles.^a

Authority	Number of taxa described	Percentages
Arnold (1980)	2 (1)	1.4 (0.6) 0.9
Barbour (1914-1925)	3 (1)	2.1 (0.6) 1.2
Barbour and Carr (1940)	1	0.7 (0) 0.3
Bocourt (1870)	1	0.7 (0) 0.3
Bonnaterre (1789)	1 (^b)	0.7 (0) 0.3
Cochran (1923-1939)	15 (12)	10.3 (6.6) 8.3
Cope (1861-1879)	16 (3)	11.0 (1.7) 5.8
Cullom Schwartz (1980)	0 (2)	0 (1.1) 0.6
Cuvier (1807)	1	0.7 (0) 0.3
Daudin (1803)	0 (1)	0 (0.6) 0.3
Duméril and Bibron (1836-1854)	7 (1)	4.8 (0.6) 2.4
Dunn (1920)	1 (4)	0.7 (2.2) 1.5
Fischer (1856-1888)	2 (1)	1.4 (0.6) 0.9
Gali and Schwartz (1982)	0 (1)	0 (0.6) 0.3
Gali <i>et al.</i> (1988)	0 (1)	0 (0.6) 0.3
Gans and Alexander (1962)	1	0.7 (0) 0.3
Garman (1887-1888)	1 (1)	0.7 (0.6) 0.6
Graham (1981)	1	0.7 (0) 0.3
Grant (1951)	1	0.7 (0) 0.3
Gravenhorst (1837)	1	0.7 (0) 0.3
Gray (1840-1852)	2	1.4 (0) 0.6
Günther (1858-1865)	4	2.7 (0) 1.2
Hedges and Thomas (1989)	1	0.7 (0) 0.3
Hertz (1976)	1	0.7 (0) 0.3
Incháustegui <i>et al.</i> (1985)	1	0.7 (0) 0.3
Jan (1867)	0 (1)	0 (0.6) 0.3
Kerster and Smith (1955)	1	0.7 (0) 0.3
Lazell (1961)	1	0.7 (0) 0.3
MacLeay (1834)	1	0.7 (0) 0.3
Meerwarth (1901)	1	0.7 (0) 0.3
Mertens (1938-1950)	1(5)	0.7 (2.8) 1.8
Moreau de Jonnés (1818)	1	0.7 (0) 0.3
Noble (1923)	1(3)	0.7 (1.7) 1.2
Noble and Hassler (1933)	5 (1)	3.4 (0.6) 1.8
Peters (1863)	1	0.7 (0) 0.3
Rand (1961)	1	0.7 (0) 0.3
Reinhardt and Lütken (1863)	0 (2)	0 (1.1) 0.6
Richmond (1964)	2	1.4 (0) 0.6

Table 4.7 cont.

Ruibal (1946)	1	0.7 (0) 0.3
Schlegel (1837)	1	0.7 (0) 0.3
Schmidt (1919-1928)	4	2.7 (0) 1.2
Schwartz (1964-1983)	10 (106)	6.8 (58.6) 35.6
Schwartz and Franz (1976)	1	0.7 (0) 0.3
Schwartz and Graham (1980)	1	0.7 (0) 0.3
Schwartz <i>et al.</i> (1979)	1	0.7 (0) 0.3
Schwartz and Henderson (1982)	0 (1)	0 (0.6) 0.3
Schwartz and Incháustegui (1976)	1	0.7 (0) 0.3
Schwartz and Jacobs (1989)	0 (1)	0 (0.6) 0.3
Schwartz and Klinikowski (1966)	1 (9)	0.7 (5.0) 3.1
Schwartz and Rossman (1976)	1	0.7 (0) 0.3
Schwartz and Thomas (1965-1977)	2 (3)	1.4 (1.7) 1.5
Sheplan and Schwartz (1974)	0 (4)	9 (2.2) 1.2
Shreve (1936-1968)	6 (4)	4.1 (2.2) 3.1
Spix (1825)	1	0.7 (0) 0.3
Steindachner (1867)	1	0.7 (0) 0.3
Thomas (1962-1989)	10 (2)	6.8 (1.1) 3.7
Thomas and Hedges (1988-1993)	5	3.4 (0) 1.5
Thomas <i>et al.</i> (1985)	3	2.1 (0) 0.9
Thomas and Schwartz (1965-1983)	5 (8)	3.4 (4.4) 4.0
Wagler (1830)	1	0.7 (0) 0.3
Weinland (1862)	1	0.7 (0) 0.3
Williams (1960-1975)	7 (1)	4.8 (0.6) 2.4
Williams and Rand (1969)	1	0.7 (0) 0.3
Williams and Webster (1974)	1	0.7 (0) 0.3

*The range of publication dates is given in parentheses after each name. The number of currently recognized species described is followed by the number of subspecies currently accepted as valid (nominated subspecies are not included). Percentages are for species (subspecies) and for all currently recognized taxa at these levels.

^bHispaniolan endemic nominate subspecies.

and *Ameiva regularis* (= *A. chrysoleama regularis*). Four additional reptilian species [*Pelophilus* (= *Epicrates*) *fordii*, *Hypsirhynchus ferox*, *Phylodryas* (= *Ialtris*) *dorsalis*, *Ahaetulla* (= *Uromacer*) *frenata*] were described by Albert Günther (1858, 1861, 1865). David Weinland (1862) described *Amphisbaena innocens*, and Johannes Reinhardt and Christian Lütken (1862) named two taxa currently recognized as subspe-

cies; included among the latter was the exceedingly abundant *Anolis (distichus) dominicensis*, which Powell (1993) suggested was specifically distinct from Bahamian *A. distichus*. Wilhelm Peters (1863) described *Alsophis anomalus*. In 1867, Giorgio Jan and Ferdinando Sordelli described *Dromicus protenus* (= *Antillophis parvifrons protenus*) and Franz Steindachner *Sphaerodactylus copei*. Marie-Firmin Bocourt (1870) described *Anolis brevirostris*, and the end of the period of European dominance came with the description of *Hemidactylus haitianus* (as *H. brookii haitianus*) by Hermann Meerwarth in 1901.

In the latter half of the nineteenth century, a transition from European to North American scientists, and the North Americans would continue to dominate the taxonomy of Hispaniolan reptiles through the present day. Edward Drinker Cope (1861, 1861 [1862], 1862a,b, 1864, 1868, 1879) described 19 taxa, 16 currently recognized as species and three more as subspecies. Among Cope's species are the nearly ubiquitous green anoles, *Anolis chlorocyanus* and *A. coelestinus*, the widespread grass anole, *A. semilineatus*, as well as the exceedingly common *A. cybotes*, *A. distichus*, *Celestus stenurus*, *Leiocephalus personatus*, and *Dromicus* (= *Antillophis*) *parvifrons*. Samuel Garman (1887a,b) described *Sphaerodactylus picturatus* (= *S. copei picturatus*) and *Anolis haetianus*. As for amphibians, the pace accelerated after the turn of the century, and many of the same persons were responsible. Thomas Barbour (1914, 1925) named five species (one, *Anolis doris*, now considered a subspecies of *A. cybotes*), and with Archie Carr (1940) described [*Pseudemys* (= *Trachemys*) *decorata*]. Emmett R. Dunn (1920a,b) named a lizard and four snakes (all of the latter now are considered subspecies of *Antillophis parvifrons* or *Uromacer frenatus*). G. Kingsley Noble, alone (1923b) and with William G. Hassler (1933), described six species and four currently recognized subspecies. However, dominating this period in Hispaniolan taxonomy was Doris M. Cochran. In a series of papers (1923c, 1927, 1928b,c, 1931a,b, 1932a,b,c, 1933, 1934a,b, 1935, 1939), she named 27 currently recognized taxa (15 species and 12 subspecies), and represented among them were lizards, snakes, and an amphisbaenian. Acknowledging the importance of Cochran's contributions, Rodolfo Ruibal (1946) described *Sphaerodactylus cochranae*. Also prominent during this time, and an exception to the general dominance by North Americans of 20th century Hispaniolan taxonomy, was Robert Mertens, who described in three papers (1938, 1939, 1950) one

currently recognized species (*Anolis aliniger*) and five additional taxa currently considered subspecies.

However, and again as in amphibians, an acknowledgment of the tremendous diversity among Hispaniolan reptiles came only in the second half of this century – and, again, was largely due to the efforts of Albert Schwartz. In an impressively long list of papers from 1964 to 1989, alone (1964b, 1965b,e, 1966b, 1967a,b, 1968a,b,d, 1969a,b, 1970a,b,c,d, 1971c,d,e, 1973a,b,d, 1974a,b, 1975a,b,c, 1976c,d, 1977b,c, 1978b,c, 1979a,c,d,e,f, 1980c, 1981, 1983) or with one or more of many collaborators (Cullom and Schwartz, 1980; Gali and Schwartz, 1982; Gali *et al.*, 1988; Incháustegui *et al.*, 1985; Schwartz and Franz, 1976; Schwartz and Graham, 1980; Schwartz *et al.*, 1979; Schwartz and Henderson, 1982; Schwartz and Incháustegui, 1976; Schwartz and Jacobs, 1989; Schwartz and Klinikowski, 1966; Schwartz and Rossman, 1976; Schwartz and Thomas, 1964 [1965], 1965, 1977; Sheplan and Schwartz, 1974; Thomas and Schwartz, 1965, 1967, 1977, 1983a,b), he named a total of 24 species and 136 subspecies, fully 16.4% of the species and an astounding 75.1% of the reptilian subspecies on Hispaniola. Although difficult to ascertain solely from reading and using the keys included in guides such as Henderson and Schwartz (1984a) and Henderson *et al.* (1984), Schwartz brought to his taxonomy a sense of an organism's total gestalt, along with its ecology and the often very detailed characters identified in his keys (Thomas, 1996). Herpetologists who have not worked on Hispaniola simply cannot appreciate the clarity of the differences among geographic races often separated more distinctly by habitat than by distance.

Still, Schwartz was not alone in elucidating the growing knowledge of reptilian diversity on Hispaniola. Although better known for his work elsewhere in the West Indies, Chapman Grant (1951) described *Celestus curtissi*. Harold Kerster and Hobart Smith (1955) described *Phyllodactylus wirshingi* from Puerto Rican material. James "Skip" Lazell, usually associated with the Lesser Antilles, named *Sphaerodactylus shrevei* in 1961. Carl Gans and A. A. Alexander (1962) described *Amphisbaena gonavensis* in their analysis of Hispaniolan amphisbaenians. Two species of *Typhlops* were described by Neil Richmond (1964) in a paper that began the process of elucidating the relationships among these scolecocephidians. In 1976, Paul Hertz, better known for his studies of saurian thermal biology, named *Anolis alumina*. Sixto Incháustegui, a Dominican naturalist, with Schwartz (1976) and with Schwartz and Robert W.

Henderson (1985) identified *Diploglossus* (= *Celestus*) *marcanoii* and *D.* (= *Celestus*) *carraulti*, respectively. Douglas Arnold (1980) identified two species and a subspecies in his analysis of the *Anolis brevirostris* complex.

However, despite of these many, frequently prominent contributors, a few persons (in addition to Schwartz) stand out. Ernest E. Williams (1960, 1962a,b, 1963a,b, 1965a,b, 1975) named seven species and one subspecies of *Anolis*, plus another species (*A. rupinae*) with Thomas Preston Webster in 1974 and still another (*A. insolitus*) with A. Stanley Rand in 1969. Rand, who with Williams applied the ecomorph concept to West Indian anoles (see Chapter 7), also described *A. koopmani* in 1961. Richard Thomas, alone (1965a,b,c, 1966b, 1968, 1971, 1974a, 1982, 1989), with Schwartz (Schwartz and Thomas, 1964 [1965], 1965, 1977; Thomas and Schwartz, 1965, 1967, 1977, 1983a,b), more recently with S. Blair Hedges (Hedges and Thomas, 1989a; Thomas and Hedges, 1988, 1989, 1992, 1993), and with Roy W. McDiarmid and Fred G. Thompson (1985) described a total of 26 species and 13 subspecies of Hispaniolan reptiles. Thus Thomas is second only to Schwartz in the total number of taxa described and, when only species are considered, he is responsible for more than any other single person. Hedges, also, is worthy of more than casual mention. With Thomas, he has identified six reptilian species, but of particular note are his recent synoptic work on origins and relationships of the herpetofauna and his explorations into many nearly inaccessible areas searching for additional taxa that may still lurk there.

The two species of *Trachemys*, described in this century by Barbour and Carr (1940, *T. decorata*) and Schmidt (1928, *T. stejnegeri*), have been subjected to various generic reassignments as a result of our frequently changing, but growing understanding of relationships among the American emydid turtles. Originally described as *Pseudemys*, McDowell (1964) placed both forms (along with species currently in *Chrysemys*, *Pseudemys*, and *Trachemys*) in the genus *Chrysemys*, *sensu lato*, but recognized at a subgeneric level an affinity between North American *T. scripta* and the West Indian forms. Seidel and Inchaustegui Miranda (1984) elevated McDowell's subgenera to full generic status. In addition, considerable confusion has existed regarding the specific and subspecific assignments of the Hispaniolan forms. Mertens and Wermuth (1955) and Wermuth and Mertens (1961, 1977) had considered both taxa to be subspecies of *T. terrapen*; Pritchard (1979) had indicated that only

one form, *T. decorata*, occurred on Hispaniola; and Williams (1956) and Schwartz and Thomas (1975) had considered *T. s. vicina* a subspecies of Cuban *T. decussata*. Seidel and Incháustegui Miranda (1984) established the current usage, which has been followed by all subsequent workers.

In their initial descriptions, all Hispaniolan amphisbaenians were placed in the genus *Amphisbaena*, and for all but two, *A. gonavensis* and *A. hyporissor*, the original assignment to species is still recognized. *Amphisbaena gonavensis* first was described by Gans and Alexander (1962) as a subspecies of *A. innocens*, and *A. hyporissor* by Thomas (1965a) as a main island subspecies of *A. gonavensis*. In addition, Gans and Alexander (1962) considered *A. caudalis* a subspecies of *A. innocens*, but Schwartz and Thomas (1975) noted that the two forms had been collected in sympatry on Île Grande Cayemite. Only minor confusion was caused by Schmidt (1928), who inadvertently created a *nomen nudum* by using *Amphisbaena weinlandi* for *A. innocens* Weinland.

Historically, Hispaniolan anguids have been assigned to the genera *Celestus*, *Diploglossus*, *Panolopus*, *Sauresia*, and *Wetmorena*. *Panolopus costatus* was described by Cope (1861 [1862]), but more recent confusion has centered on the differences between *Celestus* and *Diploglossus*, among which many of the species have been repeatedly and variously assigned. Following Strahm and Schwartz (1977), Henderson and Schwartz (1984a) and Henderson *et al.* (1984) distinguished these genera by the presence (*Diploglossus*) or absence of a "well-developed osteodermal radix." Savage and Lips (1993) rejected assignments to genera on the basis of this character because the presumed differences represent ontogenetic stages. Instead, they used the presence (*Diploglossus*) or absence (*Celestus*) of claw sheaths to distinguish these forms and, in doing so, placed all Hispaniolan forms formerly referred to *Diploglossus* in *Celestus*.

Sauresia sepsoides was described by Gray (1852) as a new genus of "Scincidae." Weinland (1862) described the same form as *Embryopus habichii* from Haitian material. In both instances, these lizards were distinguished from other known taxa in having only four toes. The monotypic genus *Wetmorena* was established by Cochran (1927) for *W. haetiana*, and was segregated from other West Indian anguids by the lack of external ear openings. As species in both genera have claw sheaths, Savage and Lips (1993) suggested that they are allied with *Diploglossus* and resulted from speciation within that group. Based on albumin ID and DNA sequence data [Hedges *et al.*, 1992b; Hass, Maxson, and Hedges,

manuscript in preparation (cited in Hedges, 1996a)], both Hispaniolan endemic genera were placed in a monophyletic radiation of Antillean *Celestus*. These changes first appeared formally in Powell *et al.* (1996a).

The gekkonid genus *Aristelliger* was erected for *A. lar* by Cope (1861 [1862]). Grant (1931a) described *A. cochranae* from Navassa Island, and Cochran (1933) described *A. expectatus* from Haiti. Mertens (1939), without comment, sunk the latter to a subspecies of the former, and this was followed subsequently until *A. expectatus* was elevated back to full-species status by Powell *et al.* (1996a). Noble and Klingel (1932) erected the genus *Aristelligella* for small members of the genus *Aristelliger* with two or more asymmetrical terminal scancers (Bauer and Russell, 1993). Although not explicitly stated, that description implicated *A. barbouri* as the type species, although two species (the other being *A. cochranae*) were assigned to the new genus. Hecht (1951) and others have used *Aristelligella* as a subgenus, but others [R. I. Crombie, personal communication (cited in Bauer and Russell, 1993)] regard it as valid at the generic level.

Gymnodactylus (= *Gonatodes*) *notatus* was described by Reinhardt and Lütken (1862) from Haitian material. Barbour (1937), in his third list of West Indian amphibians and reptiles, placed the species in the genus *Gonatodes*, previously applied to *G. albigularis* [sic] by Fitzinger (1843). Vanzolini and Williams (1962) sunk *notatus* to a subspecies of *G. albigularis*.

The two Hispaniolan representatives of the genus *Hemidactylus* have obvious African affinities (Kluge, 1969). *Hemidactylus mabouia*, with a nearly pan-Neotropical distribution, was described from Lesser Antillean material (Duméril and Duméril, 1851; Stejneger, 1904; Smith and Taylor, 1950) as *Gecko mabouia* by Moreau de Jonnés in 1818, but still is considered conspecific with populations in Africa. *Hemidactylus haitianus* was described as *H. brookii haitianus* by Meerwarth (1901). The *H. brookii* complex [R. I. Crombie, personal communication (cited in Powell *et al.*, 1996a)] has a broad distribution in the Eastern Hemisphere, across equatorial Africa (including the Cape Verde Islands), probably several Indian Ocean islands (the taxonomic status of these populations is unclear), the Indian subcontinent, and through the Malay–Indonesian Archipelago. The Antillean populations were elevated to full-species status by Powell *et al.* (1996a).

Geckos in the genus *Phyllodactylus* originally were described by Gray (1830) from “tropical South America?” (Gray, 1845), but the type

species, *P. pulcher*, is restricted to Barbados (Parker, 1935). These lizards are characterized by leaf-like terminal subdigital lamellae and have a broad, but spotty distribution throughout the Neotropics (Dixon, 1973a). The presence of this species in the Greater Antilles was established with the description of *P. wirshingi* from Isla Caja de Muertos, Puerto Rico, by Kerster and Smith in 1955. The first published acknowledgment that geckos in the genus *Phyllodactylus* occurred on Hispaniola was in a remark in Schwartz and Thomas (1975), in which they noted that "These lizards are similar to *Ph. wirshingi*, but we merely point out that the genus occurs on Hispaniola." Additional populations had been discovered by the time Schwartz (1979f) described the two endemic Hispaniolan subspecies, *P. w. hispaniolae* and *P. w. sommeri*.

The genus *Sphaerodactylus* was named by Wagler (1830), and all 32 Hispaniolan species were properly assigned at the time of their original descriptions. Morphological studies resulted in the recognition of several species groups (Thomas and Schwartz, 1966, 1983a; Shreve, 1968; Thomas, 1975; Schwartz and Graham, 1980; Schwartz and Garrido, 1981a, 1985; Schwartz, 1983), but these studies provided few insights into relationships among the groups. Hass (1991), based on starch-gel electrophoretic data and immunological comparisons of serum albumin, suggested that West Indian forms are monophyletic (*S. sputator* section of Hass, 1996) and that Greater Antillean *Sphaerodactylus* were partitioned into two main groups, the *cinereus* and *argus* series, both including Hispaniolan species. Using DNA sequence data, Hass (1996) modified her previous classification; Hispaniolan species were largely unaffected, except that *S. elasmorhynchus* was not assigned to a subsection.

The iguanid (*sensu strictu*) genus *Cyclura* is represented on Hispaniola by two species. *Cyclura cornuta* was the first Hispaniolan reptile to be described, as *Lacerta cornuta* by Bonnaterre (1789). Lacépède (1789) soon thereafter placed this species in the genus *Iguana*, but Wagler (1830) erected the monotypic genus *Metopoceros* for *cornuta*. Fitzinger (1843), however, treated *Metopoceros* as a subgenus of *Hypsilophus*. These iguanas were placed in the genus *Cyclura*, erected by Harlan (1824 [1825]) for *C. carinata* from the Bahamas, by Cope (1886).

Aloponotus ricordii was named by Duméril and Bibron (1837). Fitzinger (1843) recognized *Aloponotus* as a subgenus of *Hypsilophus*, but Boulenger (1885) placed *Aloponotus ricordii* Duméril and Bibron (1837), *Hypsilophus* (*Aloponotus*) *ricordii* Fitzinger (1843), and *Aloponotus ricordi* Gray (1845) in the synonymy of *Metopoceros cor-*

nutus (Bonnaterre, 1789); in the process misspelling each name. This species was placed in the genus *Cyclura* by Cochran (1924).

The names *Cyclura onchiopsis* and *C. nigerrima*, both described from Navassa Island by Cope (1885, 1886), have been used interchangeably; the confusion apparently resulting from the fact that the holotype of *C. nigerrima* (USNM 9974) bears a lower number than any syntype of *C. onchiopsis* (USNM 9977), although the latter name was used in an earlier publication. *Cyclura stejnegeri* was described by Barbour and Noble (1916) from Isla Mona in the Mona Channel between Hispaniola and Puerto Rico. Barbour (1937) relegated *C. stejnegeri* to subspecific status within *C. cornuta*, and Schwartz and Thomas (1975) did the same for *C. onchiopsis*. A number of efforts to elevate either or both to full species have not met with wide acceptance (see remark in Schwartz and Henderson, 1988:122). As a result, Hispaniolan populations continue to be recognized as constituting a nominate subspecies.

All 42 species of Hispaniolan anoles currently are placed in the genus *Anolis*. However, this consistency has not always characterized this group. Cope erected a number of genera and subgenera for various West Indian forms, and three Hispaniolan species were included: *Eupristus baleatus* (1864), *Anolis (Ctenocercus) coelestinus* (1862b), and *Anolis (Anolis) cybotes* (1862b). Cochran (1934b) erected the genus *Audantia* for *A. armouri*, presumably on the basis of a distinct "transverse gular fold." She subsequently described *Audantia shrevei* in 1939. Etheridge (1960), by inference, placed these species in the genus *Anolis*. Later, Williams (1963b) treated *A. armouri* as a subspecies of *A. cybotes*, but he elevated it back to a full species in 1976. In 1935, Cochran erected the genus *Xiphocercus* for *X. darlingtoni*. Williams (1962b) placed this taxon in *Anolis*, creating a nomenclatural conflict with *A. darlingtoni* Cochran (1939), for which Williams provided the substitute name, *A. etheridgei*. However, the most dramatic effort to generically partition the anoline lizards, by Guyer and Savage (1986), was based largely on morphology. Schwartz and Henderson (1988) applied these generic assignments to West Indian forms. As a result, Hispaniolan species were placed in the genera *Ctenonotus* (*altavelensis*, *armouri*, *brevirostris*, *caudalis*, *cristatellus*, *cybotes*, *distichus*, *eugenegrahami*, *haetianus*, *longitibialis*, *marcanoi*, *marron*, *shrevei*, *whitemani*) and *Semiurus* (*baleatus*, *barahonae*, *ricordii*), but none were assigned to the genera *Dactyloa* or *Norops* (see comments below regarding *Chamaelinorops*). Cannatella and de Queiroz (1988) and Williams (1989a) raised substantive questions

regarding the validity of these genera, which despite their frequent use by workers on the Neotropical mainland (see note added in proof in Vitt and Zani, 1996b), have not subsequently been applied widely to West Indian forms. Hedges (1996a) suggested that classifications such as those of Williams (1976) and Burnell and Hedges (1990), in which informal categories like series and groups were used, are preferable, as they allow for disagreement and realignment without affecting formal classification.

The morphologically (Forsgaard, 1983) and ecologically (Flores *et al.*, 1994) distinctive *Anolis barbouri* was placed in the monotypic genus *Chamaelinorops* by Schmidt (1919). Although presumably from Navassa Island, Thomas (1966a) restricted the type locality to the Massif de la Hotte in Haiti. In the interim, *C. wetmorei* had been described by Cochran (1928a) from Hispaniola. Hass *et al.* (1993), based largely on DNA sequence data, synonymized *Chamaelinorops* (and Cuban *Chamaeleolis*) within *Anolis*. The lack of agreement over taxonomy reflects the dichotomy of opinions regarding the origin of the species. Etheridge (1960) and Wyles and Gorman (1980a) suggested that this lizard is very derived, having arisen within *Anolis*, but Williams (1977) and Case and Williams (1987) argued that the species is very primitive, maybe the most primitive of all extant anoles.

General relationships among West Indian anoline lizards, postulated by Etheridge (1960) and Williams (1976), largely were based on osteology and karyotypes. Subsequent work, based on albumin ID data (e.g., Wyles and Gorman, 1980; Shochat and Dessauer, 1981; Hass *et al.*, 1993), allozyme data (Burnell and Hedges, 1990), and DNA sequence data (Hass *et al.*, 1993), has refined our understanding of those relationships. Many of the series and species groups recognized by Williams (1976) were supported, although his major alpha/beta dichotomy was not.

The following Hispaniolan species originally were described as subspecies: *Anolis aliniger* (*A. chloro-cyanus aliniger* Mertens, 1939), *A. altavelensis* (*A. dominicensis altavelensis* Noble and Hassler, 1933), *A. barahonae* (*A. ricordii barahonae* Williams, 1962a), *A. caudalis* (*A. dominicensis caudalis* Cochran, 1932b), and *A. dolichocephalus* (*A. hendersoni dolichocephalus* Williams, 1963a). *Anolis bahorucoensis* has been sunk to subspecies (*A. hendersoni baharucoensis* [sic] Williams, 1963a) and subsequently elevated back to a full species. *Anolis cybotes doris* (Barbour, 1925) and *A. distichus dominicensis* (Reinhardt and Lütken, 1862) originally were described as species, but have been relegated

subsequently to subspecific status. Most described species which later were synonymized can be attributed to scattered collecting activities during a time when an understanding of the diversity and biogeography of Hispaniolan forms was woefully incomplete; included in this group are *A. laeviceps* (= *A. chlorocyanus*) (Lichtenstein, 1856), *A. latirostris* (= *A. coelestinus*) from Navassa Island (Schmidt, 1919) (also see below), and *A. biauritus* (= *A. distichus*) (Meerwarth, 1901). However, the situation involving *A. cochranæ* (= *A. semilineatus*), described by Williams and Rand (1961), occurred more recently. Additional confusion has prevailed regarding the status of closely related forms, notably among the distichoids (*A. distichus* and the *A. brevirostris* complex) (Arnold, 1980; Williams and Case, 1986) and cybotoids (Williams, 1963b; Case and Williams, 1988; also see remarks in Schwartz and Henderson, 1988).

Mabuya lineolata was described as an endemic Hispaniolan species by Noble and Hassler (1933). Unfortunately, the taxonomic status of *Mabuya bistrriata*, known only from the vicinity of Santo Domingo, is considerably less clear-cut. Until Powell *et al.* (1996a), this species was listed among the Hispaniolan fauna as *M. mabouya*. Schwartz and Henderson (1988) presented a rather detailed synonymy, but also said that "the taxonomy of Antillean *Mabuya* is not so simple as current nomenclature indicates; however, the study of this group is complicated by the extinction or virtual extinction of a number of island populations." They then referred the reader to "more complete synonymies" in Dunn (1935) and Peters and Donoso-Barros (1970). Although the latter has been updated (Peters and Donoso-Barros, 1986), no clarification of the status of West Indian populations was forthcoming. Avila-Pires (1995) addressed the status of Amazonia *Mabuya* in what is now considered the *M. mabouya* complex, with revisions suggesting that the West Indian forms are, in fact, *M. bistrriata*. Although this nomenclature was used by Hedges (1996a), Murphy (1996), and Powell *et al.* (1996a), the latter stated that "the assignment of Antillean populations remains unresolved." The Hispaniolan material currently is assigned to *M. bistrriata sloanei*, originally described as *Scincus sloanii* by Daudin (1803).

The polytypic species *Ameiva chrysolæma* was described by Cope (1868). *Amiva* [sic] *vittipunctata* (Cope, 1871) and *Cnemidophorus affinis* (Fischer, 1883) were subsequently placed in the synonymy of the nominate subspecies. *Ameiva abbotti* was described by Noble (1923b) as a form endemic to Isla Beata, and *Ameiva regularis* by Fischer (1888) from northern Haiti; both are now relegated to subspecific status. Addi-

tional subspecies were described by Cochran (1934b, *A. c. woodi*), Mertens (1938, *A. c. boekeri*), Schwartz (1968a, *A. c. quadrijugis*), and Schwartz (1973a, *A. c. evulsa*), but the definitive analysis of this species is attributed to Schwartz and Klinikowski (1966). They named nine currently recognized subspecies, clarified the status of those forms previously described, and also named *A. leberi* as a patternless subspecies of *A. chrysolaeama*. Schwartz and Thomas (1975) elevated the latter, stating "that *A. ch. ficta* and *A. leberi* are broadly syntopic without intergradation for about 30 kilometers along the Pedernales–Oviedo road, and *A. leberi* occurs at the type-locality of *A. ch. ficta*." However, Sproston *et al.* (1998) found no ecological segregation between the two forms and suggested that the status of *A. leberi* be reevaluated.

Ameiva lineolata was described by Duméril and Bibron (1939). Noble (1923b) named *A. beatensis* from Isla Beata, but Mertens (1939) relegated it to a subspecies of *A. lineolata*. Four additional subspecies were defined by Schwartz (1965e). A similar history applies to *A. taeniura*. Originally described by Cope (1862a), insular species subsequently were described on Île de la Gonâve (*A. barboursi* by Cochran, 1928a), Isla Saona (*A. rosamondae* by Cochran, 1934b), and Navassa Island (*A. navassae* by Schmidt, 1919). *Ameiva t. meyerabichi* was named by Mertens (1939) in a paper in which he also relegated *A. barboursi* and *A. rosamondae* to subspecific status. However, the definitive work on the species was by Schwartz (1967b), in which he described eight additional subspecies and sunk *A. navassae* into the synonymy of *A. taeniura*. Another insular subspecies, *A. t. pentamerinthus* from Île Grande Cayemite, was named by Schwartz (1968a).

Frost and Etheridge (1989) determined that lizards in the endemic West Indian genus *Leiocephalus* were in the same clade as South American *Tropidurus* and *Liolemus*. They referred to this clade as the family Tropiduridae, and placed *Leiocephalus* in the subfamily Leiocephalinae. Pregill (1992), in his analysis of the West Indian forms, followed this treatment. However, Hedges *et al.* (1992b) and C. A. Hass, *et al.* [manuscript in preparation (cited in Hedges, 1996a)] presented albumin ID and DNA sequence data that suggested a closer affinity between *Leiocephalus* and North American Crotaphytidae. Although unresolved (may the distinctiveness of these lizards warrant recognition at the familial level?), little doubt exists that Hispaniola, with ten extant species, is the center of their radiation, which currently encompasses the West Indies north of Puerto Rico [although material known only from fossils indicates that the

range of the genus once included all of the major islands and banks of the West Indies, as least as far south as Martinique (Pregill *et al.*, 1988)].

The first Hispaniolan species to be described was *Pristonotus schreibersii* (Gravenhorst, 1837 [1838]), later assigned to *Steironotus* by Fitzinger (1843), and finally to *Leiocephalus* Gray, 1827 by Cope (1868). Cope also described *L. melanochlorus* (1862b), *Liocephalus* [sic] *macropus* (1862b), *Liocephalus* [sic] *trigeminatus* (= *L. personatus*) (1862b), and *Liocephalus* [sic] *eremitus* (1868), the last from Navassa Island. *Leiocephalus barahonensis* was described by Schmidt (1921a), treated as a subspecies of *L. personatus* by Mertens (1939) and Cochran (1941), and reelevated to full-species status by Schwartz (1967a). Noble and Hassler (1933) described *L. altavelensis* from Isla Alto Velo, the placement of which has subsequently caused some confusion. Schwartz (1967a) suggested that the population might represent (1) an aberrant form of *L. barahonensis*, (2) a distinct species, or (3) a subspecies of *L. vinculum* – despite its peculiar distribution far removed from the nearest putative *L. vinculum* on Île de la Gonâve. He least liked the first and eventually settled for the third option, although Pregill (1992) presented a case for the first (currently used) and Powell (1993) also suggested that full-species status might be most accurate. Cochran (1928b) erected the genus *Hispaniolus* for *H. pratensis*. She distinguished *Hispaniolus* from *Leiocephalus* on the basis of the former lacking dorsal or caudal crests. Etheridge (1966) placed *Hispaniolus* into the synonymy of *Leiocephalus*. Cochran (1932a) first used the combination *L. personatus*, and she also named *L. lunatus* (1934a), *L. semilineatus* (1941), and *L. vinculum* (1941) as subspecies of the former. All were elevated to full species by Schwartz (1967a), who also suggested that these species, with *L. barahonensis*, constituted the *L. personatus* group; Pregill (1992), however, found little evidence to support that characterization. *Leiocephalus endomychus*, originally described as a subspecies of *L. vinculum* by Schwartz (1967a), was elevated to a full species by Pregill (1992). Schwartz (1979c) described *L. ritudira*.

The three Hispaniolan boids, all currently placed in the genus *Epicrates*, were originally described as *Pelophilus fordii* by Günther (1861), *Homalochilus striatus* by Fischer (1856), and *Chilabothrus gracilis* by Fischer (1888). In that same paper, Fischer (1888) also allocated the equally slender and largely arboreal *P. fordii* to *Chilabothrus*. Although Steindachner assigned *striatus* to *Epicrates* in 1863, the morphologically distinct *fordii* and *gracilis* were not reassigned until

Boulenger (1893) did so in his *Catalogue of the Snakes in the British Museum*. Cope (1862a) described *Homalochilus multisectus*, the type locality of which was restricted to the vicinity of Santo Domingo by Sheplan and Schwartz (1974); it currently is relegated to the synonymy of the nominate subspecies of *E. striatus* (Stull, 1935).

The sole Hispaniolan tropidophiid, *Tropidophis haetianus*, was described as *Ungualia* [sic] *haetiana* by Cope (1879), the confusion due to the similarity with the Neotropical mainland genus *Ungaliophis*, which is placed with *Trophidophis* in a distinct family. *Tropidophis conjunctus*, described by Fischer (1888), is relegated to the synonymy of the nominate subspecies, *T. h. haetianus*. Cochran (1924) treated *T. haetianus* as a subspecies of Cuban *T. maculatus*, a designation followed by Stull (1928) and others until Schwartz and Marsh (1960) justified treatment of these forms as a distinct species.

Among Hispaniolan colubrids, two species of racers have been described: *Alsophis melanichnus* by Cope (1862a) and *Zamenis* (= *Alsophis*) *anomalus* by Peters (1863). *Dromicus parvifrons* also was described by Cope (1862a), and was allocated to *Antillophis* by Maglio (1970). Subsequent to the original description, a number of species and subspecies were described and assigned variously to *Dromicus*, *Leimadophis*, and *Leptophis*. Jan and Sordelli (1867) described *Dromicus protenus*, and Fischer (1883) *Leptophis frenatus* (with an erroneous type locality listed as Sierra Leone). Both were placed within *Antillophis parvifrons* (as *A. p. protenus*) by Maglio (1970). Dunn (1920b) named *Leimadophis alleni* from Île de la Gonâve and *L. tortuganus* from Île de la Tortue; both forms were relegated to subspecific status by Maglio (1970). Currently recognized subspecies originally placed in genera other than *Antillophis* are: *Leimadophis parvifrons niger* (Dunn, 1920b), *Leimadophis parvifrons lincolni* (Cochran, 1931a), *Dromicus parvifrons rosamondae* (Cochran, 1934b), and *Dromicus parvifrons paraniger* and *Dromicus parvifrons stygius* (Thomas and Schwartz, 1965). The monotypic genus *Darlingtonia* was erected by Cochran (1935) for *D. haetiana*. Recently, Hedges and Garrido (1992b) suggested that this species may be reassigned to *Arrhyton*. In contrast, unpublished allozyme data (B. I. Crother, personal communication) indicate that including *Darlingtonia* in *Arrhyton* would render the genus polyphyletic; instead, these data suggest that *Darlingtonia* and Jamaican *Arrhyton* form a group independent of Cuban and Puerto Rican *Arrhyton*. The monotypic genus *Hypsirhynchus* was erected and *H. ferox* was described by Günther (1858). Cope

(1862a) described *H. scalaris* from Haitian material, but Schwartz (1971c) relegated it to subspecific status. Günther (1858) also described *Philodryas* (= *Ialtris*) *dorsalis*. Cope (1862a) erected the genus *Ialtris* for *I. vultuosa*, which subsequently was placed into the synonymy of *I. dorsalis*, although confusion over the generic name did not cease until well into the twentieth century (Powell and Henderson, 1994). Günther's (1862) *Dromicus mentalis* and Werner's (1909) *Dromicus w-nigrum* contributed to the confusion, as did Cope's (1863b) own unjustified emendation of *Ialtris* to *Jaltris* (Smith and Wallach, 1993). *Ialtris parishi* (Cochran, 1932c) and *I. agyrtes* (Schwartz and Rossman, 1976) were described as such. Similar confusion has existed regarding snakes in the endemic genus *Uromacer*. *Dendrophis catesbyi* was described by Schlegel (1837), but Duméril *et al.* (1854) placed it in the newly erected genus *Uromacer* (Duméril, 1853) along with the newly described *U. oxyrhynchus* (although the latter was erroneously assigned a type locality of Senegal). In 1861, Duméril erected the monotypic genus *Megalocercus* for Schlegel's *D. catesbyi*. Shortly thereafter, Günther (1865) described *Ahaetulla frenata* and Garman (1887b) *U. inornatus*. The former was reassigned to *Uromacer* and the latter sunk into the synonymy of *U. frenatus* by Boulenger (1894). Further complicating an understanding of these snakes were the descriptions of three satellite island forms as new species. Dunn (1920b) named *U. scandax* from Île de la Tortue (relegated to a subspecies of *U. catesbyi* by Mertens, 1939). Dunn (1920b) also described *U. dorsalis* from Île de la Gonâve, and Cochran (1931a) described *U. wetmorei* from Isla Beata. The former was sunk to subspecific status (within *U. frenatus*) by Schwartz (1979d) and the latter by Horn (1969). Interestingly, Schwartz (1979d) subsequently recognized the validity of *U. wetmorei*, but Schwartz and Thomas (1975) followed Horn, and Henderson and Schwartz (1984b) formally acknowledged that action. Lazell (1983) suggested that the Antillean xenodontine genera *Alsophis*, "*Dromicus*," *Arrhyton*, and *Liophis* were weakly differentiated and used the name *Liophis* for members of this complex; nevertheless, subsequent workers largely have followed Schwartz and Henderson (1988) in using the more "classical" names.

The history of scolecophidian taxonomy of Hispaniolan forms is simultaneously straightforward and relatively complex. The first leptotyphlopoid discovered on the island was *Leptotyphlops pyrites*, described by Thomas (1965b). All of the other three known species were described by Thomas *et al.* (1985). In contrast, the situation regarding Hispaniolan

typhlopoid snakes is more circuitous, with the current state of knowledge incomplete until publication of the analysis by Thomas (1989). Much of the confusion occurred because the name *Typhlops lumbricalis* (but not *Anguis lumbricalis* of Linnaeus, 1758) was variously attributed to Hispaniolan material (Cochran, 1924, 1941; Richmond, 1964; Thomas, 1974a) and subsequently assigned to several species. Cope (1868) described *T. sulcatus*, originally from Navassa Island, and Barbour (1914) described *T. pusillus*, and only these names and *T. lumbricalis* were used until Richmond's (1964) study, in which he named *T. capitulatus*, *T. gonavensis*, and *T. haitiensis*. The latter was placed in the synonymy of *T. sulcatus* in a paper by Thomas (1965c), in which he also named *T. syntherus* and relegated *T. gonavensis* to a subspecies of *T. capitulatus*. *Typhlops hectus* was described by Thomas (1974a) for a part of the material which had been referred to as *T. lumbricalis*. In his doctoral dissertation, Thomas (1976) made reference to three new forms, all previously designated in the literature as *T. lumbricalis*, as *Typhlops* I. Subspecies A, *Typhlops* I. Subspecies B, and *Typhlops* II. With their formal descriptions by Thomas (1989) as *T. schwartzi*, *T. tetrathyreus*, and *T. titanops*, respectively, the currently accepted status of the nine Hispaniolan species finally was resolved.

The final component of the Hispaniolan herpetofauna and the lone-crocodilian representative on the island is *Crocodylus acutus*, described by Cuvier (1807) from "St.-Dominique."

Navassa Island

The three taxonomic authorities for the known Navassan reptiles are listed in Table 4.8. Thomas (1966a) summarized the state of the herpetofauna and provided a history of collecting activity on the island. Cope (1868), based on specimens collected by W. J. Rasin, described *Celestus badius*, *Liocephalus* [sic] *eremitus*, *Typhlops sulcatus*, and *Ungalia* (= *Tropidophis*) *bucculentus*. In 1885, he described *Cyclura onchiopsis*. Mertens (1939) relegated *C. badius* to a subspecies in *Celestus costatus*, an allocation with which Schwartz (1964b), in his study of Hispaniolan *Diploglossus costatus*, concurred. *Liocephalus eremitus* was described from a single specimen in the Rasin collection. A later specimen, collected by R. H. Beck, was actually a specimen of *L. melanochlorus* (Thomas, 1966a), and was probably taken from the Les Cayes region of Haiti. It was the latter specimen on which comments in Etheridge (1966) were based. Thomas (1966a) examined the original holotype, com-

mented on its unique features, and stated that: "The distinctness of this form plus the confidence generated by the W. J. Rasin specimens indicate that Navassa once possessed an endemic species of *Leiocephalus* which is now extinct." *Typhlops sulcatus* also was described from a single specimen. Richmond (1964) noted the existence of *T. sulcatus*, but did not compare it to *T. haitiensis*, which he named in the same paper. Thomas (1965c) first allocated a Hispaniolan specimen to *T. sulcatus*, and later (1966b) confirmed his suspicion that *T. haitiensis* belonged in the synonymy of *T. sulcatus*. Cope (1868) suggested an affinity between *Ungalia bucculentus* and *Tropidophis melanurus*, but Boulenger (1893) listed it in the synonymy of *T. maculata*, and Stull (1928) considered it a subspecies of *T. pardalis*. Bailey (1937) noted that Stejneger (1917) had supported Cope's original contention, as did he. Schwartz and Marsh (1960) concurred. Formal publication of the currently recognized trinomial appeared first in Thomas (1966a). Cope (1886) described *Cyclura nigerrima*, and the name often has been confused with *C. onchiopsis* (Cope, 1885), even after clarification in Schwartz and Thomas (1975). Regardless of name, the Navassan *Cyclura* was regarded as a distinct species by Barbour and Noble (1916), Schmidt (1919, 1921b), and Cochran (1941). However, Barbour (1937) treated it as a subspecies of *C. cornuta* and expressed doubts of its distinctiveness at even that level. Mertens (1939) agreed with the latter, but Cochran (1941) remarked on "striking differences." Thomas (1966a), without having examined specimens, "arbitrarily" used the trinomen, the convention largely followed since.

Table 4.8. Taxonomic Authorities for Native Navassan Reptiles (Thomas, 1966).^a

Authority	Number of taxa described	Percentages
Cope (1868)	2 (3)	40.0 (100.0) 62.5
Grant (1931)	1	20.0 (0) 12.5
Schmidt (1921)	2	40.0 (0) 25.0

^aThe publication date is given in parentheses after each name. The number of currently recognized species described is followed by the number of subspecies currently accepted as valid (nominate subspecies are not included). Percentages are for species (subspecies) and for all currently recognized taxa at these levels.

R. J. Beck spent a week on Navassa in 1917 (Wetmore and Swales, 1931) and collected the greatest number of Navassan specimens currently in any collection. On the basis of this material, Schmidt (1919) described *Sphaerodactylus becki*, *Anolis longiceps*, *A. latirostris*, *Chamaelinorops* (= *Anolis*) *barbouri*, *Ameiva navassae*, and noted the "probable" presence on the island of *Anolis olssoni*. *Anolis longiceps* is a large anole in the *carolinensis* group, and its Cuban (via the Cayman Islands) affinity has been discussed previously. In contrast, *A. latirostris*, characterized by a broad spatulate snout, was probably taken in Haiti, where Beck spent about a month prior to his time on Navassa Island. Thomas (1966a) examined the specimen, determined it was damaged (crushing the snout), and placed *A. latirostris* in the synonymy of *A. coelestinus*. *Chamaelinorops* (= *Anolis*) *barbouri* is another example of a specimen the origin of which was erroneously recorded. Undoubtedly these lizards were taken in Haiti by Beck from mesic upland habitats which are absent on Navassa. The questions regarding *Ameiva navassae* are unresolved. Described from a single specimen, Thomas (1966a) assumed that the collection data were incorrect and placed this taxon in the synonymy of *A. taeniura*, with a type locality restricted to the western part of the Tiburon Peninsula. He noted, however, that Schwartz, in a personal communication, had indicated that this specimen was at or above the range in several counts for *A. taeniura* in the Les Cayes–Camp Perrin region. Subsequently, Schwartz (1967b) treated *navassae* as a subspecies of *A. taeniura* endemic to Navassa Island, and that allocation has been followed since. That Schmidt (1919) had some suspicions regarding the origin of Beck's specimens is obvious from his note that *Anolis olssoni* was "probably from Navassa Island." In addition to the question of its origin, the specimen turned out to be an example of *A. semilineatus* [Schmidt, 1921b; E. E. Williams, personal communication (cited in Thomas, 1966a)]. Schmidt (1921b) also noted the presence on Navassa of *Sphaerodactylus cinereus* and *A. distichus*. Like *A. semilineatus*, these undoubtedly were collected by Beck in Haiti (Thomas, 1966a).

Aristelliger cochranae was described by Grant (1931a), and shortly thereafter Cochran (1933) described *A. expectatus* from Hispaniola. Cochran (1941) discussed the distinctiveness of the two forms, and counts by Thomas (1966b) generally supported her contentions. Nevertheless, Thomas (1966b) chose to follow Mertens (1939) in considering these geckos conspecific. This was generally accepted until Powell *et al.* (1996a), based largely on published data, comments in Powell (1993),

and prior use by Powell and Parmerlee (1993), once again elevated the Hispaniolan form to full-species status.

Fossil Record

In general, the West Indian fossil record is sparse. However, due to the fortunate series of "accidents" that led to the formation of amber deposits on Hispaniola, at least a few specimens are preserved in their entirety, safely encased in hardened resin. The only known frog is an *Eleutherodactylus* sp. from Upper Eocene to late Pleistocene amber (Poinar and Cannatella, 1987), although Poinar (1992) referred to six other Dominican frogs in amber, none have been "verified for authenticity or described." That tortoises, *Geochelone* sp., were found historically on Navassa Island and Hispaniola is supported by late Pleistocene to Holocene material reported by Auffenberg (1967) and Franz and Woods (1983), respectively. Etheridge (1965) described late Pleistocene material from the Dominican Republic as *Aristelliger lar*, *Celestus costatus*, *C. stenurus*, *Diploglossus* (= *Celestus*) sp., *Anolis chlorocyanus*, *A. cybotes*, *A. ricordii*, *Leiocephalus apertosulcus* (†), *L. personatus*, *Ameiva chrysolaeama*, and *A. taeniura*. Etheridge (1965) referred to the material listed above as *Diploglossus* (= *Celestus*) sp. as *D. stenurus*, but based on its large body size, Schwartz (1970a) assumed the specimen was *D.* (= *Celestus*) *warreni*. However, the subsequent discovery of two additional large anguids on Hispaniola (Schwartz *et al.*, 1979; Incháustegui *et al.*, 1985) and the fact that Holocene dwarfism has occurred in some Antillean lizards (Pregill, 1986) renders identification of this material to species impossible. *Leiocephalus apertosulcus* was described from a right dentary, but numerous cranial and postcranial bones were also referred to this species, the extinction of which was presumed to be pre-Columbian. The holotype of *Anolis dominicanus* (†) (Rieppel, 1980) is preserved in early Miocene amber. Pregill (1984) described *Leiocephalus anonymous*, also from a right dentary, which was found by A. J. Poole around 1927 "in cave sediment probably no older than latest Pleistocene" (Pregill, 1992). Pregill (1984) referred more material to this species, and speculated that *L. anonymous* may have persisted into historical times. In the same year, Böhme (1984) described *Sphaerodactylus dommeli* from Oligocene amber.

Current Status of the Herpetofauna

Hispaniola has been subjected to the generally negative impact of non-native Americans for a longer period than any other area in the Western Hemisphere. Still, many species of Hispaniolan amphibians and especially reptiles appear to thrive despite (or maybe because of) human alterations to habitats. *Osteopilus dominicensis* is known to breed in ornamental fountains and even in heavily contaminated drainage ditches in the center of urban areas (R. Powell, personal observation). Schwartz and Henderson (1991) remarked on the fact that *Hyla pulchrrilineata* frequently occurs "in situations much disturbed by man." *Eleutherodactylus weinlandi* is encountered commonly in resort areas along the southeastern Dominican coast, where it freely takes advantage of mesic situations created by the construction of ornamental pools and by the watering of lawns and golf courses (R. Powell, personal observation). Several species of *Eleutherodactylus* often are found frequenting the debris resulting from agricultural practices and human wastefulness (Schwartz and Henderson, 1991). Some anoles, notably *Anolis distichus* and members of the *A. brevirostris* complex, *A. chlorocyanus* and its South Island counterpart *A. coelestinus*, *A. cybotes*, and the grass anoles are nearly ubiquitous, coexist successfully with humans (see Chapter 7), and may, in some instances, actually function as human commensals — a role more commonly associated with some geckos. And a few geckos similarly do, in fact, thrive. *Hemidactylus haitianus* is almost as ubiquitous at night in urban areas as are some of the anoles by day. *Sphaerodactylus difficilis* frequently occurs in human habitations and often makes use of the debris that inevitably accompanies human activities (R. Powell, personal observation). Even the generally more secretive *Aristelliger expectatus* occasionally plays the role of human commensal (Lynxwiler *et al.*, 1991). A few anguids (e.g., *Celestus costatus* and *C. stenurus*) take advantage of human-made clearings and the resultant litter, and *Ameiva chrysolaeama* and *A. taeniura* often are seen foraging in open areas of human design, even in streets and on construction sites in urban areas. When weeds and scrub were allowed to grow in a previously "manicured" park in Barahona, the density of the *A. chrysolaeama* population declined (R. Powell, personal observation). Snakes, in general, fare less well in the company of humans, who often actively persecute them, but *Antillophis parvifrons* occurs frequently in urban areas (e.g., the main street in Sosua, downtown Barahona, and an intensely developed resort

area near La Romana; R. Powell, personal observation). An adult *Hypsirhynchus ferox* was collected crossing the main highway along the coast in Barahona, *Uromacer catesbyi* is not infrequently encountered in the urban parks of Santo Domingo, and *Typhlops syntherus*, to the consternation of the local inhabitants, has been taken around an old foundation in a heavily cultivated garden along Highway 44 in Paraiso (Powell, personal observation).

However, many species fare less well. Powell and Henderson (1996a) noted (admittedly in discussing the entire West Indies, but the implications for Hispaniola are evident):

Habitat destruction is rampant and forests are rapidly disappearing in order to produce charcoal and plant crops... The introduction of the mongoose is correlated with the decline or disappearance of several snake taxa and populations... The introduction of cats, dogs, and rats has had a catastrophic effect on some *Cyclura* populations. Human numbers are booming and development (largely for the tourist trade) continues unabated.

Obviously, the habitat specialists and species with restricted ranges are most vulnerable, although the most attention generally has been devoted to a few species frequently exploited by humans and formally considered endangered. The sea turtles, although not discussed elsewhere in this chapter, largely have been eliminated from Haitian beaches, primarily due to the commercial trade for "tortoise shell" but also for meat and eggs (Ottenwalder, 1996). Those on the Dominican side of the island have fared little better. Crocodiles (*C. acutus*) once were common along most coastal areas, but as a result of uncontrolled hunting for meat, hides, and parts presumably with aphrodisiac qualities, now appear to be restricted to the largely saline lakes in the Valle de Neiba/Plaine de Cul-de-Sac (Thorbjarnarson, 1988; Schubert and Santana, 1996). Both *Cyclura cornuta* and *C. ricordii* have been exploited for food, and population densities are low in most areas. Similarly, populations of the two aquatic turtles in the genus *Trachemys* have been severely depleted (Schubert, 1993; J. Ottenwalder and Gomez, unpublished manuscript; J. A. Ottenwalder, personal observation), and the appearance of turtles and their eggs in markets continues to aggravate the situation. However, in addition to these well-documented examples, several other species may be in even worse condition.

The very large anguid, *Celestus anelpistus*, may be restricted to a habitat (broadleaf lowland forest) which is nearly gone. Workers clearing the area from which the type series was taken had not previously seen these large lizards (Schwartz *et al.*, 1979), suggesting their rarity,

and none have been collected since (Henderson, 1988). The colubrid snake *Alsophis melanichnus* has not been collected for several decades and may be extinct (Schwartz and Henderson, 1991; Schubert, 1993; Powell and Henderson, 1998). Two species in the genus *Ialtris*, *I. agyrtes* and *I. parishi*, are at best very rare (males of *I. agyrtes* have never been collected). Although known locality records are scattered and this may be a reflection of secretive habits, this is unlikely, as both are large and probably are actively foraging and diurnally active (Schwartz and Henderson, 1991). Instead, the paucity of records undoubtedly speaks to the rarity of these species. Based on well-documented events on Hispaniola and elsewhere in the West Indies (Henderson and Sajdak, 1986; Henderson, 1992), the most probable cause is predation by the mongoose (*Herpestes javanicus*).

Despite recent concerns over declines in amphibian populations (Heyer *et al.*, 1994, and references cited therein), Hedges (1993) concluded that declaring any West Indian frog species extinct was premature [but see Joglar and Burrowes (1996) regarding the probable extinctions of Puerto Rican frogs]. Hedges (1993), however, stressed that habitat destruction (e.g., rampant deforestation) could have a catastrophic effect on the frog fauna once all forests had been destroyed. Both the Dominican Republic and Haiti are listed among the top ten tropical countries in terms of net annual deforestation, with rates of 2.5% and 3.9%, respectively (World Resources Institute, 1994). Whereas less than 1% of Haitian forests remain and many amphibians are known to occur only in moist upland forests, Hedges (1993) suggested that Haiti soon may become the world's "first major biodiversity disaster." On the other hand, Henderson and Powell (see Chapter 7) suggested the possibility that the frogs of Haiti (and possibly all of Hispaniola) will endure as long as second growth forests, banana plantations, and similar degraded habitats persist." Their explanation is that the fauna may be more euryoecious than what is generally thought to be the case. S. B. Hedges (personal communication) noted in response that many species clearly are dependent on natural (primary) vegetation. Among these are "leaf litter" species (e.g., *Eleutherodactylus thorectes*, *E. ventrilineatus*), stream "specialists" (stream quality being a reflection of forest quality), and forms associated closely with orchids and/or bromeliads. Also, amphibians may "explode" following clear-cutting or slash-and-burn activities (J. M. Ruder, A. K. Howard, and J. D. Forester, unpublished data; S. B. Hedges, personal communication), giving the impression that they may thrive in

heavily disturbed habitats, although this probably is correlated with a temporary explosion of invertebrates exploiting the overabundance of decaying vegetation. Inevitably, however, the "explosion" is followed by a "crash," after which only a few common species remain. Regardless, when habitat destruction is combined with the impact of introduced predators, such as the mongoose, black rat (*Rattus rattus*), feral cat (*Felis domesticus*), and even large, alien amphibians like *Rana catesbeiana* and *Bufo marinus* (Hedges and Thomas, 1991; Hedges, 1993, 1996a), the outlook for many Hispaniolan amphibians is grim — especially because these predators may operate in undisturbed habitats.

Both Haiti and the Dominican Republic have set aside protected areas, some of which contain the few remaining large stands of natural vegetation. Unfortunately, enforcement of the laws establishing and protecting these areas is spotty, especially in Haiti. Ottenwalder (1996) vividly documented the abject failure of Haitian authorities to enforce laws designed for the conservation of natural resources, and Hedges (1996a) recorded seeing "trees being cut and removed from one of the last remaining stands of natural forest in the country" — in the "center of Pic Macaya National Park." The situation is considerably better in the Dominican Republic (Ottenwalder, 1989), where the Departamento de Vida Silvestre and the Dirección Nacional de Parques (Hoppe, 1989; Valdez Sierra and Mateo Félix, 1992) are actively engaged in surveying the natural resources of that country, educating of the populace, and conserving areas of particular importance. However, and despite laws making it illegal to cut trees, clear-cutting and burning are common, even in national parks.

The situation on Navassa Island is complicated by the number of species that have been erroneously reported to occur on the island. However, Thomas (1966a), based on his own work and the results of five trips by various persons during which reptiles were collected, indicated that four species, all of which have been collected recently, are extant, and four additional species, none of which have been collected recently, are presumed to be extinct. Such a high rate of extirpation may be the combined effect of human alteration of habitats and the very small size and relative isolation of the island, with the latter resulting in some parts of the fauna never having been well established and others particularly vulnerable to perturbations of their limited habitats. Of the four presumably extinct forms, *Cyclura cornuta onchiopsis*, like the large iguanas elsewhere in the West Indies, is very sensitive to human activities,

and may have been driven to extinction (Thomas, 1966a). The reason for the extinction of *Leiocephalus eremitus* is more difficult to explain, because the lizards in this genus usually are sufficiently euryoecious to tolerate, if not thrive, under the onslaught of human habitat alterations – and this has raised some doubts regarding the validity of the taxon (but see Thomas, 1966a). Similarly difficult to explain is the status of *Typhlops sulcatus*. Although having a lifestyle that often precludes the ready collection of specimens, as a result of the number of trips and collectors that have visited Navassa over the years, surely additional material would have been discovered if the species was still extant. Again, however, confidence in the validity of the Rasin collection warrants recognition of *T. sulcatus* as a former member of the Navassan herpetofauna. Thomas (1966a) stated that it “is difficult to believe that [*Tropidophis melanurus*] *bucculentus* has been exterminated on Navassa; further collecting may yield more specimens of this form.”

Hedges (1997), relative to the entire West Indies, recommended that (1) deforestation be controlled, (2) park boundaries and laws against the illegal felling of trees be enforced, (3) human populations be controlled, and (4) additional protected areas be set aside. In particular reference to Hispaniola, he suggested the enlargement of Parc National Pic Macaya and Park National Morne La Visite in Haiti. Two regions of comparable biotic value in the Dominican Republic have benefited from recent government action. The Sierra de Neiba National Park was established in 1995 (Decree 221-95), and highlands in the eastern Sierra de Baoruco (encompassing Loma Remigio, Loma Pie de Palo, and Loma Trocha de Pey) were protected as the Reserva Biológica Padre Miguel Domingo Fuertes (category 1B IUCN) in July 1996 (Decree 233, Article 11). Removal or control of introduced predators also must receive a high priority [see Vogel *et al.* (1996) and Tolson (1996) for discussion of efforts on Jamaica and Puerto Rican satellite islands, respectively]. Finally, Hedges and Thomas (1991) and Hedges (1997) presented a strong case for the promotion of scientific collection and continued exploration, primarily by relaxing the often restrictive requirements for obtaining collecting or export permits.

The Puerto Rico Area

Richard Thomas

Biology Department

PO Box 23360

University of Puerto Rico-Río Piedras

San Juan, Puerto Rico 00931

Introduction: The Region

The Puerto Rico Area includes approximately 200 islands and cays on three submarine banks: The Mona Bank, including Monito, the Desecheo Bank, The Puerto Rico Bank, and the St. Croix Bank (Fig. 5.1). The native herpetofauna of this archipelago comprises three genera of anurans, one emydid turtle, and 15 genera of squamates, with a total of approximately 80 species. Puerto Rico, with an area of about 8682 km², is the largest island. With Pleistocene lowering of sea level, the most recent episode of which ended during the interval from 15,000 to 8000 years ago, the Puerto Rico Bank constituted a single elongate island that extended from Puerto Rico to Anegada (Heatwole and MacKenzie, 1966) having an area of around 21,000 km². The Puerto Rico Bank includes Puerto Rico and over 80 associated cays, which might harbor populations of reptiles or amphibians. To the east of Puerto Rico are the Passage Islands of Culebra and Vieques (and 22 associated cays), and the 93 islands of the American and British Virgin Islands (MacLean *et al.*, 1977; Heatwole *et al.*, 1981; Lazell, 1983). St. Croix and its three associated cays lie off the Bank 58 km to the south of St. Thomas. To the west of the Bank lie the islands of Desecheo (21 km west of Puerto Rico), Mona (61 km west of Puerto Rico), and its satellite, Monito, which have herpetofaunas partly or wholly derived from the Puerto Rico Bank.

The Puerto Rico Bank has a plutonic core that is overlain in coastal plains of the north and south coasts of Puerto Rico by thick deposits of Miocene to Oligocene limestone, giving rise to various karst terrains, particularly prominent and extreme in the north where one finds haystack karst (Monroe, 1976). The southwestern corner of Puerto Rico is an accreted wedge of sea-floor containing ultramafic and serpentine rocks

(Case *et al.*, 1984; Burke, 1988). Except for the island of Anegada, a low-lying limestone platform at the eastern extreme of the Bank, along with most of the cays surrounding Puerto Rico, and a large part of Vieques, there are no extensive limestone exposures elsewhere on the Bank. Some of the Virgin Islands do have areas of limestone. Mona and Monito Islands, which lie off the bank to the west are elevated limestone platforms. Desecheo is igneous in composition.

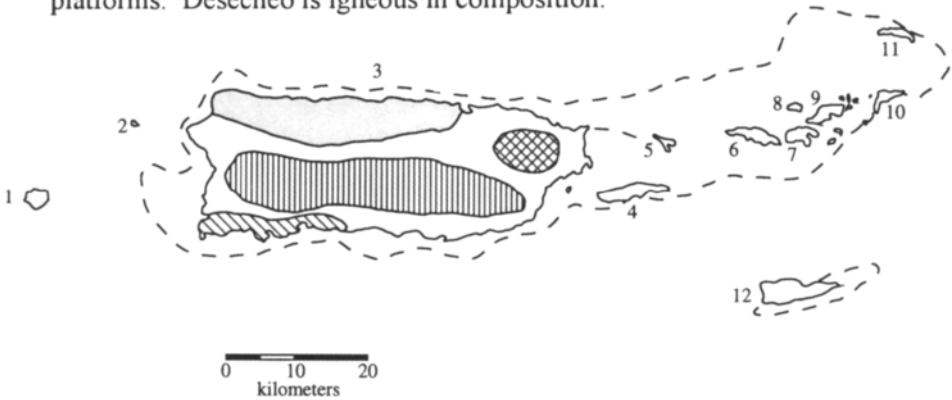


Figure 5.1. The islands of Puerto Rico area: 1, Mona; 2, Desecheo; 3, Puerto Rico; 4, Vieques; 5, Culebra; 6, St. Thomas; 7, St. John; 8, Jost Van Dyke; 9, Tortola; 10, Virgin Gorda; 11, Anegada; 12, St. Croix. The dotted line delimits the Puerto Rico Bank. Within Puerto Rico the approximate limits of major regions are indicated: northern karst area (shaded), Cordillera Central and associated highlands (vertical hatching), the Luquillo Massif (cross hatching), and the xeric extreme (diagonal hatching).

The geological subdivisions notwithstanding, the overriding determinant of distributions of amphibians and reptiles is probably moisture. Beyond Puerto Rico, the other islands of the area are more xerophytic: scrub to dry woods with the most mesic areas being found in ravines and on the highest parts of the islands. Ewel and Whitmore (1973), employing Holdridge's Life Zone system, note that on the Passage Islands and U.S. Virgin islands the areas are climatically capable of supporting Subtropical Moist Forest (= Life Zone) range from 72% on St. Thomas to 0% on Culebra. Tortola has a substantial amount of Subtropical Moist Forest habitat. St. Croix, although larger than any of the eastern bank islands, has only 16% in the Moist Tropical Forest Life Zone. Mona is reported by Ewel and Whitmore to be 100% Subtropical Dry Forest. These generally xeric conditions of the eastern islands extend to the eastern extreme of Puerto Rico, but the uplift of the Luquillo Mountains

continuing into the Cordillera Central and the northern coastal plain are mesic, with the wettest areas being in the higher elevations of the Luquillo massif, where the annual rainfall may exceed 4000 mm/year. The south coast of Puerto Rico is in a rain shadow that increases in dryness toward the west with the areas of least rainfall receiving about one meter a year and producing the most extremely xerophytic conditions where this falls on a limestone substrate. Mona and Desecheo are xerophytic but not so extreme as the driest part of the southwestern coast of Puerto Rico. Many of the Puerto Rico Bank islands appear slightly tilted toward the south. This results in many islands having rocky or sandy "high-energy" shores on the north and "drowned" coastlines on the south with mangrove areas and mangrove cays along the south coasts of the larger islands. Along the east and south coasts of Puerto Rico are numerous cays and islets of various sizes and elevations. The south coast cays are more homogeneous: low, with extensive mangrove and with limestone, coral rubble, and sand substrates, although some have small rocky areas, and Caja de Muertos, the largest and farthest offshore, is rocky and elevated (90 m).

A Brief History of Herpetology in the Puerto Rico Area

Endemic species were first described from the Puerto Rico area by Duméril and Bibron in their *Erpétologie générale* (1844). Major additions to the herpetofauna were made by Stejneger (1904), Schmidt (1928), and Grant in the 1930s. Descriptions of new species have continued with diminishing frequency into recent years (Hedges and Thomas, 1991a). Thomas and Joglar (1996) discussed the history of the development of herpetology in the area.

The Puerto Rican Herpetofauna within the Greater Antillean Assemblages

The Puerto Rico Area shares with the rest of the Greater Antilles a fauna of widespread genera. Each major Greater Antillean island bank has an *Eleutherodactylus* fauna, an *Anolis* fauna, a *Sphaerodactylus* fauna, at least one anguid lizard (*Diploglossus* or *Celestus*), at least one large *Alsophis*, one or more small generalized colubrids, at least one *Typhlops*, and a freshwater emydid turtle of the genus *Trachemys* (two on Hispaniola). In addition, most islands have one or more amphisbaenids (lacking on Jamaica and the Bahamas), one or more hylid frogs (lacking on Puerto Rico), and one or more tropidophiid snakes (lacking on Puerto

Rico). However, for post-Pleistocene extinctions on Jamaica and Puerto Rico, the iguanid genus *Leiocephalus* would also be a part of the widespread Greater Antillean fauna (Pregill, 1981b).

Puerto Rico, like Jamaica, has no endemic genera of amphibians and reptiles. In Cuba and Hispaniola a number of species in the past have been accorded generic status, because they depart strikingly in morphology from their nearest relatives. To avoid an epizootic of parphyly, several of these genera have been sunk into larger parent genera, e. g., *Chamaelinorops* and *Chamaeleolis* into *Anolis* (Hass *et al.*, 1993), the anguids *Sauresia* and *Wetmorena* into *Celestus*, *Cadea* into *Amphisbaena* (Hedges, 1996b). Despite the nomenclatural changes, the fact remains that those "genera" represent significant departures in morphology from their congeners. The Puerto Rico Area lacks any such strikingly divergent species, the most divergent certainly being the live-bearing *Eleutherodactylus jasperi*, whose uniqueness lies in its mode of reproduction. In external morphology, it is a fairly standard arboreal member of its genus.

Patterns of Distribution: The Puerto Rico Bank

Of the 80 or so species in the entire area, 57 occur on Puerto Rico, and, of those, 37 are confined to that island and its immediate offshore cays. As one progresses east of Puerto Rico into the Passage Islands and the Virgin Islands, there is a marked drop-off in species, no doubt due largely to the reduction in area of the eastern islands but also due to the absence of high-elevation, moist habitat.

Distribution within Puerto Rico

Within Puerto Rico several patterns of distribution are found among the amphibians and reptiles. First, island wide species are found at almost all elevations: *Leptodactylus albilabris*, *Eleutherodactylus antillensis*, *Eleutherodactylus coqui*, and *Anolis cristatellus*. It is unclear to what extent *L. albilabris*, *E. antillensis*, and *A. cristatellus* occur at very high elevations naturally; they prefer open habitats and may have extended their elevational ranges along roads and clearings. *L. albilabris* is arguably the most ecologically tolerant of all Puerto Rican amphibians and reptiles; it occurs from the most xeric conditions to wet montane forest, albeit in disturbed situations, and is the first frog in the assembly sequence to be found on small islands. Second, there are widespread, generally low to moderate elevation species (*Eleutherodactylus antillensis*, *E. cochranae*, *Leptodactylus albilabris*, *Anolis cristatellus*, *A. pulchellus*,

Epicrates inornatus, *Alsophis portoricensis*, *Arrhyton exiguum*, *Sphaerodactylus macrolepis*, etc.). Third, there are widespread mostly upland mesophilic species (*Eleutherodactylus brittoni*, *E. eneidae*, *E. wightmanae*, *Anolis cuvieri*, *A. gundlachi*, *A. krugi*, *A. occultus*, *Diploglossus plei*, *Sphaerodactylus klauberi*, *Typhlops rostellatus*). Fourth, there are seven species restricted to the xeric southern coastal area (*Ameiva wetmorei*, *Anolis cooki*, *Anolis poncensis*, *Phyllodactylus wirshingi*, *S. roosevelti*, *Amphisbaena xera*, *Typhlops granti*). Beyond that, there is a miscellany of restricted distributions, some whose areas of restriction are ecologically or physiographically definable (*Eleutherodactylus unicolor* restricted to the high elevations of the Luquillo Mountains, *E. jasperi* restricted to the Sierra de Cayey, *Amphisbaena schmidti*, restricted to the northern karst area, *E. cooki* restricted to the Sierra de Panduras). and some which are not: *Amphisbaena bakeri*, restricted to western, mostly interior Puerto Rico in both karst and non-karst areas; *Sphaerodactylus gaigeae* and *Typhlops hypomethes* with U-shaped distributions around the eastern end of the island; *Sphaerodactylus nicholsi*, occupying the southwestern coastal region with an outlier population on the north coast; *Sphaerodactylus townsendi*, the sister species of the former, ranging from the southeast to the east and onto the eastern cays and Vieques. Over much of the region the distribution of species is more determined by moisture and elevation (or an interaction of the two). The ranges of species liking open, lowland habitat were presumably more restricted in pre-Columbian times, when Puerto Rico was more heavily forested, and those preferring closed canopy forest were presumably more extensively distributed. In the 1940s Puerto Rico reached a maximum of deforestation due to cultivation and pasturage, such that only 6% of the area was forested (including shade crops such as coffee). With economic changes and the dwindling of agriculture, there has been a recrudescence of forest so that 35% of the land was in forest by 1987 (Birdsey and Weaver, 1987). This certainly must fall far short of the percentage of aboriginal forest. Hedges (1999) analyzed the distributions of amphibians in the Puerto Rico Area and found that the species density is greatest in the Luquillo Mountains (El Yunque), which although not the highest peaks in the island they are the wettest. The occurrence of the highest densities around high mountain peaks is a recurrent pattern (Hedges, 1999).

The largest of the eastern Puerto Rico Bank islands is St. Thomas with an area of 77 km². The large islands of the eastern part of the bank, (Culebra, Vieques, St. Thomas, St. John, Tortola, Virgin Gorda, and Anegada) all have approximately 14-19 species. Most of the species of these islands are bank wide, occurring also on Puerto Rico, but there is a

small set of eastern bank endemics: *E. schwartzi*, *Anolis ernestwilliamsi*, *Sphaerodactylus parthenopion*, *Amphisbaena fenestrata*, *Typhlops catapontus*, and *Typhlops richardi*. The giant *Anolis roosevelti*, described from Culebra in 1931 but not found alive since (Grant, 1931b), probably also belongs in this category. *Eleutherodactylus lentus* of the eastern Bank islands also occurs off the Bank on St. Croix.

Aside from the eastern bank endemics, the Passage Islands and the Virgin Islands for the most part share a group of widespread sun-tolerant species that are found in the lowlands of Puerto Rico: *Eleutherodactylus antillensis*, *Eleutherodactylus cochranæ*, *Leptodactylus albilabris*, *Ameiva exsul*, *Anolis cristatellus*, *Anolis pulchellus*, *Anolis stratulus*, *Sphaerodactylus macrolepis*, *Alsophis portoricensis*, and *Arrhyton exiguum*. The foregoing species are the common, conspicuous part of the shared herpetofauna. The skink, *Mabuya mabouya*, although not common in Puerto Rico, is also part of this shared lowland assemblage, as is the toad *Bufo lemur*, which occurs on Puerto Rico and Virgin Gorda; it was no doubt continuously distributed in the past. The ecological vicariants, *Typhlops platycephalus*, *hypomethes*, *richardi*, and *catapontus* are also part of this bank-wide assemblage of species. *Sphaerodactylus parthenopion*, known only from the Virgin Islands of Tortola and Virgin Gorda, has its closest relative in *S. nicholsi* of southwestern Puerto Rico. Several species are endemic (or possibly relict endemics) to one or more of these eastern islands. *Anolis roosevelti* and *Amphisbaena fenestrata* are eastern bank species without known or obvious sister species on Puerto Rico, although one presumes that their closest relatives are (or were) found on Puerto Rico. The emydid turtle, *Trachemys stejnegeri* extends east of Puerto Rico to Vieques and possibly Culebra (Seidel, 1988a).

Lazell (1983, Table 1) presented a listing of the British Virgin Islands, their areas, elevations and species of amphibians and reptiles. The table demonstrates, in effect, the assembly rules for the herpetofaunas of these islands. The sequence is basically hierarchical with the species occupying the very small islands occurring also on the larger islands, but there is some irregularity in what species are found on the larger islands. The species that will occupy all islands from the smallest to the largest is *Anolis cristatellus* (or its daucusite equivalent, *A. ernestwilliamsi*); next is *Sphaerodactylus macrolepis*, and next *Ameiva exsul*, followed sequentially by *Anolis stratulus*, *Mabuya mabouya*, and so on. The smallest of the British Virgin Islands to be occupied by a frog (*Leptodactylus albilabris*) is Jost Van Dyke the arwa of which is 840 ha. The assembly sequence is basically similar for the La Cordillera Cays, the Passage Islands, and the U. S. Virgin Islands (Heatwole *et al.*, 1981; R. Thomas,

unpublished data), except that west of Culebra and its cays, *Sphaerodactylus townsendi* replaces *S. macrolepis* as the small cay species, *S. macrolepis* being found only on high cays (Palominos, Piñeros). In the La Cordillera cays *Leptodactylus albilabris* occupies Cayo Icaos (Heatwole *et al.*, 1981), which is smaller at 139 ha than Jost Van Dyke. On the southern coastal cays of Puerto Rico, a similar sequence of occupancy occurs, but it appears that *Ameiva exsul* will occupy some low cays with mostly halophytic undergrowth that *Sphaerodactylus townsendi* does not. An indication of the rigidity with which the assembly rules hold sway, at least in the smaller range of island size, is the failure of anoles introduced onto a small cay by Levins and Heatwole (1973). They introduced, at different times, numbers of *Anolis stratulus* and *Anolis pulchellus* onto a small, vegetated sand cay (Cayo Palominitos) off eastern Puerto Rico, and both introductions failed. Both of these species are members of the widespread, heliothermic lizards occupying many large and small islands of the bank. This cay, although variable in size, is below the size at which the assembly sequence would predict occupancy by these species. *Anolis cristatellus* and *Ameiva exsul* occupy the island naturally.

The islands of the Puerto Rico area occupying banks separate from the Puerto Rico Bank tend to have high endemism and a greater proportion of species from beyond the Area than do islands on the Bank, although their peripheral positions near Hispaniola (Mona) or the Lesser Antilles (St. Croix) are probably crucial; Desecheo, closer to Puerto Rico has no Hispaniola-derived species. Mona has species related to Puerto Rico Bank species (six of nine), but *Cyclura cornuta* is Hispaniolan in affinity, as, very likely, is *Eleutherodactylus monensis*, which resembles *E. probolaeus*, found in similar habitats on the eastern tip of Hispaniola (R. Thomas, personal observation). Three of Mona's nine amphibians and reptiles are considered endemic species. Two of Monito's three reptiles (*Anolis* cf. *monensis* and *Mabuya mabouya*) are closest to species found on Mona, but *Sphaerodactylus micropithecus* is morphologically peculiar and of uncertain affinities. St. Croix, lying off the bank 58 km to the south of St. Thomas is much larger than any of the eastern Bank islands (230 km²) but has a smaller herpetofauna (13 species) than any of these islands. However, it has a more highly endemic fauna with four endemic species (*Ameiva polops*, *Anolis acutus*, *Sphaerodactylus beattyi*, and *Alsophis sancticrucis*).

It is of great interest to know how the fauna of the Puerto Rico area is related to other parts of the Antillean Region: What ancient connections may be indicated by patterns of phylogenetic relationship, or what pattern of overwater dispersal may likewise have formed linkages of re-

lationship. Some of the Puerto Rico Area species occur beyond the archipelago or show obvious affinities with species that do: The freshwater turtle, *Trachemys stejnegeri* occurs on the Puerto Rico Bank, Hispaniola, and Great Inagua Island in the southern Bahamas (Seidel, 1988a). A population on Marie Galante in the Guadeloupean Archipelago is apparently introduced (Seidel and Adkins, 1987; Seidel, 1988a). *Hemidactylus brooki* (Hispaniola, Cuba), *Phyllodactylus wirshingi* (Hispaniola), *Sphaerodactylus macrolepis* (the Anguilla Bank), *Mabuya mabouya* (widespread in the Antilles extending to South America); the subspecies to which the Puerto Rico area populations pertain, *M. m. sloanei*, is Greater Antillean in distribution, with *M. m. mabouya* occurring in the Lesser Antilles. However, the systematics of this "species" is probably not well represented by the nomenclature. The Mona Island rock iguana is closely related to the Hispaniolan *Cyclura cornuta*, and is considered by most to be subspecifically related to that form (Schwartz and Henderson, 1991). *Sphaerodactylus macrolepis* ranges throughout the Puerto Rico Bank but occurs off the bank on St. Croix and on the Anguilla Bank, where the populations are recognized as *S. macrolepis parvus*. A *Typhlops* occurring in the Turks and Caicos Islands is very close to *T. platycephalus* (Thomas, 1976); similarly *Anolis scriptus* of the southern Bahamas, including the Turks and Caicos islands is remarkably close to *Anolis cristatellus* in morphology (R. Thomas, unpublished data) and in genetic distance (Gorman *et al.*, 1980a). The study by Gorman *et al.* (1980a) also showed that *A. desecheensis* of Desecheo Island is only weakly, if at all, differentiated from *A. cristatellus*, a fact also remarked upon by Lazell (1983). The frog *Leptodactylus albilabris* is very close to the Hispaniolan *L. dominicensis* (Schwartz and Thomas, 1975).

A number of more explicit attempts than the preceding kinds of data have been made to assess phylogenetic relationships of taxa having species in the Puerto Rico Area. The ever popular anoles have received the lion's share of the attention. Etheridge's (1960) morphologically based study was the first but it did not propose a total evolutionary tree, although it did recognize taxonomic groupings. Williams' (1972) analysis of ecomorph evolution presumed that the Puerto Rican anoles constitute a single radiation. Shochat and Dessauer (1981) used immunological distance to assess evolutionary relationships of the anoles. Gorman *et al.* (1980a, 1983) specifically focused on Puerto Rican anoles. Guyer and Savage (1986) combined molecular and morphological data sets to obtain a phylogenetic tree of the anoles, which indicated that the Puerto Rican species have primarily Hispaniolan affinities. Burnell and Hedges (1990) analyzed West Indian *Anolis* phylogeny using electrophoretic data, and Hass *et al.* (1993) derived phylogenies based on sequence di-

vergence of serum albumin and DNA sequence data. These molecular-based studies indicate relationships of the Puerto Rico Area anoles with both the Greater Antilles (Jamaica, Cuba, Hispaniola) and the Lesser Antilles. Roughgarden (1995) combined data from a number of sources and concluded that the Puerto Rican anoles have a sister-group relationship with the *bimaculatus* group anoles from the Lesser Antilles north of Martinique. Unfortunately, his tree was made only for anoles of Puerto Rico and the Lesser Antilles.

Joglar (1989) using morphological data and Hedges (1989a) using allozyme data derived phylogenies for the West Indian *Eleutherodactylus*. Joglar's study resolved only major groups of West Indian species, which are all widely distributed; Hedges' results, on the other hand, indicated membership of the Puerto Rican species in a clade with the Lesser Antillean species.

Tolson (1987) analyzed cloacal gland secretion of *Epicrates* to derive a phylogeny, and Kluge (1988) extended this work adding morphological data to obtain a phylogeny of *Epicrates*, which indicated that *E. inornatus* form a monophyletic group with the Jamaican *E. subflavus*, and that *E. monensis* is part of a clade whose other members are in Hispaniola.

Hass (1991, 1996) used allozyme, microcomplement fixation, and mitochondrial DNA data to derive phylogenies of *Sphaerodactylus*. In both of these papers she showed that the Puerto Rican *Sphaerodactylus* are part of the *argus* series, which includes taxa from Puerto Rico (four), Hispaniola (four), Jamaica (two), and Cuba (one). Although they did not try to derive a phylogeny of the species, Murphy *et al.* (1984) examined the relationship of two species of Puerto Rican *Sphaerodactylus* using allozyme data, showing that *S. townsendi* and *S. nicholsi* are separate species, as their morphology suggests. The genetic distance of those two from *S. roosevelti* is within the range of closely related species (Gorman *et al.*, 1980a).

Thomas (1989) proposed a phylogeny of *Typhlops* based on morphological data, and Hedges and Thomas (1991) looked at the relationships of the Puerto Rican *Typhlops* using allozyme data. Thomas' results indicated that most of the Puerto Rican species belong in with a group having both Hispaniolan and Lesser Antillean members. *Typhlops rostellatus* belongs to another Hispaniolan clade and has presumably invaded Puerto Rico more recently than the other Puerto Rican species, which seem to be part of a radiation within Puerto Rico. The allozyme data confirm that *T. rostellatus* is well separate from the *platycephalus-richardi-hypomethes* complex.

It seems highly likely that the Puerto Rico Bank herpetofauna has phyletic relationships both to the west with Hispaniola, Jamaica, and the Bahamas, and to the southeast with the Lesser Antilles.

The Incursive and Anthropochore Herpetofauna

One of the problems in considering any fauna is which of its components are present by virtue of natural dispersal phenomena (whether vicariant or waif) and which have been brought into the area by humans, perhaps transported from areas far beyond reach of natural dispersal and having no import to the deciphering of biotic history that gave rise to the fauna. In general, we recognize that species found in the region and nowhere else are almost certainly native species. On Puerto Rico no one doubts that *Eleutherodactylus portoricensis* is a native species, and no one doubts that *Hyla cinerea* is an introduced one. Species present problems when their plausible histories are natural dispersal or anthropochory. Most of the introduced species on Puerto Rico present no question of their endemism: *Bufo marinus*, a South American species, was introduced in the 1920s to combat insect pests in the cane fields (Rivero, 1978). Puerto Rico is the only Greater Antillean island to lack native hylid frogs, but three, *Hyla cinerea* (North America), *Osteopilus septentrionalis* (Cuba), and *Scinax rubra* (South America), have been introduced (Schwartz and Henderson, 1991; Thomas and Joglar, 1996). *Osteopilus septentrionalis* was also introduced on St. Thomas and St. Croix, as was *Eleutherodactylus coqui* (Schwartz and Henderson, 1991). The North American *Rana catesbeiana* was introduced as a potential food source (not realized) during the early part of the century (Rivero, 1978); it is widespread and common on Puerto Rico. Likewise the crocodylian, *Caiman crocodylus*, established on the north coast of Puerto Rico and present on Vieques (whether breeding or not is not known), was certainly introduced by man, probably as the result of the pet trade, sometime in the 1950s or 1960s (personal observation). The Cuban Rock Iguana, *Cyclura nubila*, established on one small island, Isla Magueyes in the bay of Parguera, is known to have been established from captive animals (Rivero, 1978).

Perhaps the first species to be suspected of being introduced are the two species of *Hemidactylus*: *H. brooki* and *H. mabouia*, which, because populations of both species exist in west Africa, were assumed by many to have been brought over during the slave trade (see Kluge, 1969 for a review). However, Kluge (1969) demonstrated the likelihood that the New World populations of *H. brooki* and *H. mabouia* are native to the New World, having become established by natural means. As late as the

1930s, when Grant (1932) wrote about *Hemidactylus* only *H. brooki* was known from Puerto Rico (Grant had a substantial collection from widespread localities), and Schmidt in the 1920s (Schmidt, 1926, 1928, had not collected them). *Hemidactylus mabouia* was known only from Vieques and St. Thomas. The situation is quite different today; *H. mabouia* is widespread on Puerto Rico, in some places enormously abundant (personal observation). It is now known from 34 islands of the Puerto Rico area (Lazell, 1983; Heatwole *et al.* 1981; R. Thomas, unpublished data). In my experience on the cays around Puerto Rico, Vieques, and Culebra it is present only on those cays that have had human habitations and has so far been found on six out of 61 cays investigated. Puerto Rico is at the northern extreme of the distribution of this species, and it is likely that *H. mabouia* is extending its range, in part due to human agency. *Hemidactylus brooki*, on the other hand, appears not to be a small island species; it is known from a few of the offshore islands of Cuba and Hispaniola (Schwartz and Henderson, 1991).

Similar problems are presented by the tortoise *Geochelone carbonaria* and the lizards *Thecadactylus rapicaudus* and *Iguana iguana*. All three of these reptiles occur from South America through the Lesser Antilles and into the Puerto Rico region (Schwartz and Henderson, 1991). Were they were brought by Amerindian migrants, by Europeans, or by natural dispersal? It is, of course possible that a mixture of the two processes has occurred. It is pretty certain that *Iguana iguana* has become established on the Puerto Rican main island in the last 30 years or so, presumably from the pet trade. *Iguana iguana* has been in the Virgin Islands for a longer time, however, and Lazell (1973, 1983; personal communication) is convinced that it is native there and in the Lesser Antilles. *Geochelone carbonaria* is also probably native to some areas of its occurrence in the Antilles (Lazell, 1993), but almost certainly it has been introduced in some areas (see Censky, 1988 for a discussion). There are records of this species from Puerto Rico, but it is not clear that there have ever been breeding populations (Censky, 1988).

This Page Intentionally Left Blank

The Lesser Antillean Fauna

*Ellen J. Censky*¹ and *Hinrich Kaiser*²

¹*Connecticut State Museum of Natural History, University of Connecticut, Storrs, Connecticut 06269*

²*Department of Biology, La Sierra University, Riverside, California 92515*

Introduction

The Lesser Antilles extend from South America northward in a long arc of more than a dozen major islands and hundreds of small islets (Fig. 6.1). The islands form a discrete group separated from the Virgin Islands to the north by the Anegada Passage and from Trinidad and Tobago to the south by the Tobago Trough. They originated as a volcanic arc during the Late Cretaceous (Perfit and Williams, 1989) and were uplifted into their present location on the eastern edge of the Caribbean Plate in the Early Oligocene (Malfait and Dinkelman, 1972). These islands are clustered on undersea banks, each of which was emergent as one large island until the post-Pleistocene rise in sea levels. As sea levels rose, large islands fragmented into many smaller ones. The present-day islands vary in size from Guadeloupe, the largest (1706 km²), to the tiniest of cays (Willet, 1987). Furthermore, profound differences in elevation exist, ranging from 1397 m on Martinique to only about 39 m on Barbuda and Anguilla.

Two separate arcs exist, an outer, older limestone arc and an inner, younger volcanic arc. The inner one still has active volcanoes and extends from Grenada through the western half of Guadeloupe (Basse Terre) on to Saba and is composed of igneous, extrusive rock. The outer limestone arc extends from Marie Galante and eastern Guadeloupe (Grande Terre) northward to Anguilla and Sombrero. Its igneous layer has eroded away, and it is capped with oceanic limestone. The limestone arc tends to be lower in elevation (highest elevation: 484 m on St. Martin)

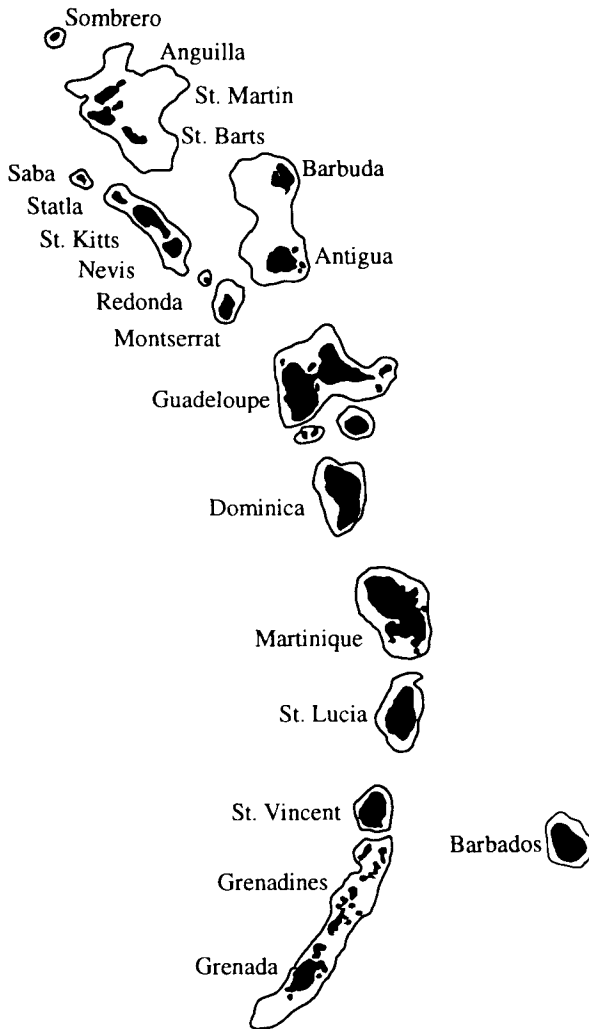


Figure 6.1. The islands of the Lesser Antilles. The map depicts the relative locations of the major islands. The maps of individual islands are contoured, with each contour line representing 1000 ft.

Table 6.1. Endemism in the Lesser Antilles

Island	Lesser Antillean Endemism		
	Bank endemic (%)	Lesser Antillean endemic (%)	Number of Species ^a
Anguilla	47	67	7/3/5
St. Martin/Maarten	38	61	5/3/5
St. Barthelemy	56	70	5/2/3
Barbuda	38	75	3/3/2
Antigua	38	66	5/3/4
Saba	14	57	1/3/3
St. Eustasius	10	73	1/7/3
St. Christophers (Kitts)	9	67	1/7/4
Nevis	8	75	1/8/3
Redonda	50		2/0/2
Montserrat	21	57	3/5/6
Guadeloupe	25	55	5/6/9
Dominica	22	78	4/10/4
Martinique	28	61	5/6/7
St. Lucia	29	62	6/7/8
St. Vincent	18	65	3/8/6
Grenadines	27	60	4/5/6
Grenada	30	45	6/3/11
Barbados	40	70	4/3/3

^a Number of Bank endemics/number of Lesser Antillean endemics excluding bank endemics/number of non-Antillean species.

Thin lines separate the Banks of islands.

than the volcanic arc (highest elevation: 1397 m on Martinique). The amount of rainfall that each island receives depends, in part, on its elevation. The low, limestone islands receive less annual rainfall (mean = 1106 mm) than the higher volcanic islands [mean = 4193 mm; Eastern Caribbean Natural Area Management Program (ECNAMP), 1980]. According to the characterization of biomes established by Holdridge (1967), all the limestone caribeas are classified as Tropical Dry Forest. Biomes on islands of the volcanic arc, on the other hand, range from Tropical Dry Forest in the low lying areas to Tropical Moist Forest at higher elevations. Guadeloupe (Basse Terre), Dominica, Martinique, and St. Vincent also have areas of Subtropical Rain Forest.

The number of amphibians and reptiles in the Lesser Antilles is related to island size, diversity of habitat, and distance from the mainland or another large island. The literature lists 95 species of amphibians and reptiles in the Lesser Antilles, six introduced, five known only from the original series of specimens, two known only from the fossil record, and two vagrants. The large islands from Guadeloupe south all have relatively high diversity, whereas the islands north of Guadeloupe, which all are smaller in size, have lower diversity (Table 6.1). Endemism in the Lesser Antilles is high and can be examined at two levels, bank endemism and Lesser Antillean endemism. Bank endemics are those species occurring on only one bank of islands (e.g., Anguilla Bank = Anguilla, St. Martin, St. Barthélemy; Dominica Bank = Dominica). Lesser Antillean endemics are those species which occur on more than one bank but are restricted to the Lesser Antilles. Bank endemism ranges from a low of 9 or 10% of species on St. Kitts Bank to a high of 38-56% of species on the Anguilla Bank (Table 6.1). Lesser Antillean endemism ranges from a low of 45% of the species on Grenada to 78% of the species on Dominica (Table 6.1). Because Grenada lies closest to the mainland, it is not surprising that it shares a greater proportion of its fauna with South America. Equally, Dominica has a high level of endemism because it is a large, tropical island with many habitat types and is relatively distant from the mainland. What is surprising is that the St. Kitts Bank, while having a majority of Lesser Antillean endemics (67-75%), has a very low level of bank endemism (8-10%); recent extinctions have probably contributed to this (e.g., *Cyclura* and *Leiocephalus*) (Table 6.1). Herein, we outline and discuss the diversity and taxonomic history of the Lesser Antillean herpetofauna. In terms of both biogeography and conservation, this fauna is very important

because of the high levels of endemism and because it is a faunal link between the diverse herpetofaunas of South America and the Greater Antilles.

Table 6.2. List of Species by Island.

Species	Island																		
	AN	SM	SB	BA	AT	SA	SE	SC	NE	RE	MO	GU	DO	MA	SL	SV	GS	GR	B
<i>Bufo marinus</i>					I			I	I		I	I		I	I	I		I	I
<i>Colostethus chalcopis</i>																			
<i>Eleutherodactylus amplinympha</i>																			
<i>E. barlagnei</i>																			
<i>E. euphronides</i>																			
<i>E. johstonei</i>	I	*		*	*	*	*	*	*		*	I	I	*	*	*	I	*	*
<i>E. martinicensis</i>			I		?														
<i>E. pinchoni</i>																			
<i>E. shrevei</i>																			
<i>Leptodactylus fallax</i>											(*)		*						
<i>L. validus</i>																	*	*	*
<i>Osteopilus septentrionalis</i>		I																	
<i>Scinax rubra</i>																I			
<i>Geochelone carbonaria</i>	*	*	*	*	*			D	D		*		D		D	D	*	*	*
<i>G. sombrenensis</i>	+																		
<i>Pelusios subniger</i>												I							
<i>Trachemys stejnegeri</i> ¹												I							
<i>T. scripta</i>												I							
<i>Ameiva ameiva</i> ¹																	(*)	*	*
<i>A. atrata</i>											*								
<i>A. cineracea</i>												(*)							
<i>A. corax</i>	**																		
<i>A. corvina</i>	**																		
<i>A. erythrocephala</i>								*	*	*									
<i>A. fuscata</i>													*						
<i>A. griswoldi</i>				*	*														
<i>A. major</i>														(*)					
<i>A. plei</i> ²	*	*	*																
<i>A. pluvianotata</i> ²										*	*								
<i>Anolis aeneus</i>																	*	*	*
<i>A. bimaculatus</i> ²				*	*			*	*	*									*
<i>A. extremus</i>															I				*
<i>A. ferreus</i>												**							
<i>A. gingivinus</i>	*	*	*																
<i>A. griseus</i>																	*		
<i>A. lividus</i>											*								
<i>A. luciae</i>															*				
<i>A. marmoratus</i> ¹¹										*		*							
<i>A. nubilis</i>										*									
<i>A. oculatus</i> ⁴													*					*	*
<i>A. richardi</i>														*				*	*
<i>A. roquet</i> ⁶														*					
<i>A. sabanus</i>						*										*			
<i>A. trinitatis</i>																I		*	*
<i>A. watters</i> ⁴	*	*		*	*		*	*	*						I			*	*
<i>Bachia heteropus</i> ¹																	*	*	*
<i>Cnemidophorus varzoi</i>															**				
<i>Diploglossus montisserrati</i>											(*)								
<i>Gecko gecko</i>															I				
<i>Gonatodes albogularis</i> ¹															(*)				

Diversity and Taxonomic History

Frogs

There are now 13 frog species extant in the Lesser Antilles (Table 6.2), and although this is not a high level of diversity by West Indian standards (Schwartz and Henderson, 1991), the biological and historical origins are remarkable (Kaiser, 1995). Three species have been introduced (*Bufo marinus*, *Osteopilus septentrionalis*, *Scinax rubra*), whereas three regionally endemic species (*Eleutherodactylus johnstonei*, *E. martinicensis*, *Leptodactylus validus*) have been introduced on various islands within the Lesser Antilles, and, in the case of *E. johnstonei*, to places well beyond (Table 6.2; Kaiser, 1997; Kaiser and Hardy, 1994; Kaiser and Henderson, 1994). The remaining seven species are single-island endemics.

French explorers and missionaries traveling in the late eighteenth century were among the first to return from the West Indies with anuran material, among it the first documented specimens from the Lesser Antilles. Resident scientists at the Muséum National d'Histoire Naturelle in Paris, foremost among them F. Daudin, A. -M. -C. Duméril, A. H. Duméril, and G. Bibron, made their workplace a focus of amphibian research and began to place systematic order upon the received specimens, soliciting assistance from colleagues throughout Europe. One of the results of these collaborations was the first description of a Lesser Antillean frog, namely that of *Hylodes martinicensis*, by J. J. Tschudi (1838). Given the technical limitations of the time, most Lesser Antillean specimens were placed into already existing taxa, and it has taken to this day to recognize the true diversity of the anuran fauna.

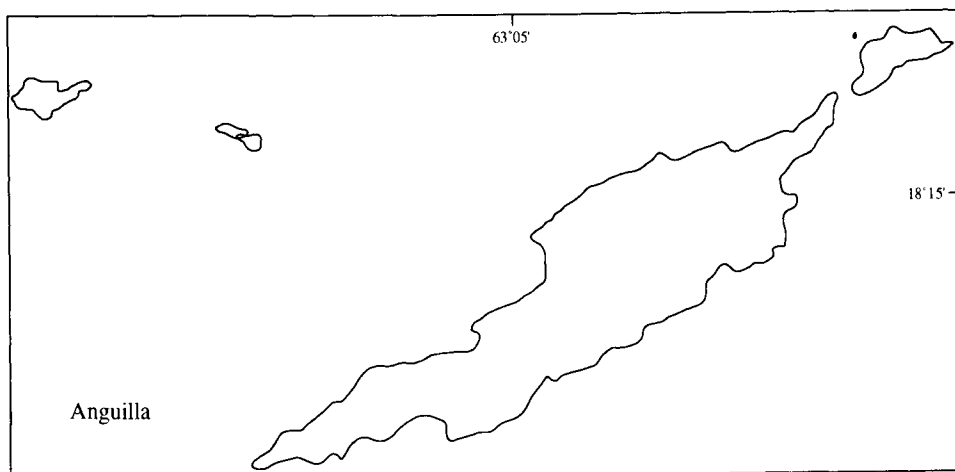
Tschudi's (1838) description of *H. martinicensis* carries with it an interesting anecdote that seems symptomatic for some of the nomenclatural hiccups in the nineteenth Century (see Lizards). During a visit to Paris, Tschudi examined a variety of specimens from the New World for his project on "Batrachians" (1838), including those cataloged by Bibron. A jar containing six small brown frogs purportedly from Martinique (but see Schwartz, 1967c) and shelved preliminarily under the name "*Hyla martinicensis*" was ready to be cataloged and described as "*Eleutherodactylus martinicensis*" by Duméril and Bibron. The specimens were of great interest to Tschudi since they were among the first ones from that island and certainly an important addition to his study of frog diversity. In his 1838 text, Tschudi used the name "*Hylodes*

Martinisensis” for these specimens, thereby providing a name priority to which Duméril and Bibron deferred in their classic herpetology text (Duméril and Bibron, 1841). However, unbeknownst to Tschudi, Bibron had changed the label of the jar to *Eleutherodactylus martinicensis* since these frogs were obviously not members of *Hyla*, indicating that this was to be the name used in a future volume of *Érpétologie Générale*. To avoid a later discrepancy between their book series and the specimen labels during relabeling of jars, they mentioned that name in their detailed description of the specimens. The name on the jar in Paris is still *Eleutherodactylus martinicensis*. A nomenclatorial clarification of the name *Eleutherodactylus* was attempted by Myers (1962), but given the number of taxonomic names that now hinge on this taxonomic eccentricity (512 at last published count; Lynch and Duellman, 1997) a final decision has not emerged.

The only other Lesser Antillean frog described in the nineteenth Century was *Leptodactylus validus* (Garman, 1887c) from St. Vincent. Placed in synonymy with *L. wagneri* (Heyer, 1970), it remained in the obscurity of classification until Heyer’s (1994) revision of the *L. wagneri* complex. The description of an endemic *Leptodactylus* from Dominica (Müller, 1926) added an interesting data point to biogeographic studies because this member of the *pentadactylus* group (Heyer, 1979) is geographically distant from the other species in that group.

After the original description of *Eleutherodactylus* frogs from the Lesser Antilles (Tschudi, 1838; Duméril and Bibron, 1841), descriptive systematic and taxonomic work resumed with the work of Thomas Barbour (1914). His monograph on the West Indian herpetofauna included the description of *Eleutherodactylus johnstonei*, and although in subsequent publications (Barbour, 1930a, 1935, 1937) the same author cast implicit doubts on the existence of the taxon (Kaiser and Hardy, 1994), its distinction has now been ascertained (Kaiser, 1992). A surge of interest in *Eleutherodactylus* occurred between 1958 and 1969, beginning with the description of *Hyla barbudensis* from a fossil ilium from Barbuda (Auffenberg, 1958). This species was placed in *Eleutherodactylus* by Lynch (1966), and synonymized with *E. johnstonei* by Pregill *et al.* (1988). The collections and work by the late Albert Schwartz (Schwartz, 1967c, 1969) became the baseline for contemporary study of *Eleutherodactylus*. *Eleutherodactylus barlagnei* was described by Lynch (1965) from material collected by J. D. Lazell and P. Barlagne. In his description, Lynch could find few external differences between the

new species and *E. martinicensis*, whereas Schwartz (1967c:51) called it “[c]ertainly the most striking and distinctive of the Lesser Antillean *Eleutherodactylus*.” Schwartz attributed Lynch’s difficulty in differentiating the new species to the state of the type series. He provided a very detailed description. In the same paper, Schwartz (1967c) described *E. pinchoni*, a miniaturized frog sympatric with *E. martinicensis* and *E. barlagnei* on the Basse Terre portion of Guadeloupe.



A recent surge in research activity has resulted in the discovery of two new species, *Eleutherodactylus amplinympha* on Dominica (Kaiser *et al.*, 1994a) and *Colostethus chalcopis* on Martinique (Kaiser *et al.*, 1994b). It also led to the removal of *E. euphronides* and *E. shrevei* from the synonymy of *E. urichi* (Kaiser *et al.*, 1994c) and of *Leptodactylus validus* from *L. wagneri* (Heyer, 1994). The dendrobatid *C. chalcopis* is a remarkable addition to the West Indian herpetofauna because it is the only dendrobatid endemic on an oceanic island. Its tadpole is atypical for the genus (Kaiser and Altig, 1994) and a systematic study of the relationship of this taxon with mainland congeners is now being completed by the junior author and colleagues. The presence of *E. amplinympha* in high forests of Dominica marks the continuation of an interesting trend among Lesser Antillean frogs. With the exception of St.

Lucia, all major islands in the volcanic arc (Guadeloupe, Dominica, Martinique, St. Vincent, Grenada) harbor at least one single-island endemic frog species, and most have a single-island endemic *Eleutherodactylus*. Looking at the distribution of taxa and simply filling the “endemism gaps,” the more widely distributed *E. martinicensis* may have originated on Martinique and the very effective colonizer *E. johnstonei* on St. Lucia.

Lizards

Compared with the relatively straightforward taxonomy of frogs, taxonomy of the lizards of these islands is less stable. For some groups, consensus is lacking at any taxonomic level above species [see Guyer and Savage (1986) and Williams (1989a) for *Anolis* and Frost and Etheridge (1989) and Lazell (1992) for Iguanidae]. The diversity of lizards on each island in the Lesser Antilles is not as great as that found in the Greater Antilles but greater than that of Lesser Antillean anurans. However, taking the Lesser Antilles as a unit, lizard diversity includes 12 genera and 42 species. In addition, one species has recently been introduced (*Gekko gecko*), whereas five others have not been collected since the original specimens were taken (+*Ameiva cineracea*, *Ameiva major*, *Gonatodes albogularis*, *Diploglossus montserrati*, and +*Leiocephalus herminieri*), and two species are known only from the fossil record (*Leiocephalus cuneus*, *Cyclura* sp.). Many of the genera are widespread (Fig. 6.2) in the Lesser Antilles (*Anolis*, *Ameiva*, *Iguana*, *Thecadactylus*, *Hemidactylus*, *Sphaerodactylus*, and *Mabuya*), whereas a few are restricted to one or a few islands (*Kentropyx*, *Gymnophthalmus*, *Bachia*, *Phyllodactylus*).

There are two genera of extant Iguania in the Lesser Antilles, *Anolis* (Polychridae) and *Iguana* (Iguanidae), and two that are extinct, *Leiocephalus* (Tropiduridae) and *Cyclura* (Iguanidae). Whereas there are only two species of *Iguana* in the Lesser Antilles, *Anolis* is represented by 16 species, most of which are bank endemics. By 1900, no fewer than 33 species of *Anolis* had been described by such noted men as Samuel Garman (1887b), Edward Drinker Cope (1864), John Edward Gray (1840), and A. -M. -C. Duméril and G. Bibron (1837). However, Boulenger (1887) synonymized 11 of Garman’s (1887b) 13 new species into four species (*virgatus* = *gingivimus*; *asper*, *nubilus*, *speciosus*, *sabanus*, *lividus* = *leachii*; *gentilis*, *cinereus*, *extremus*, *vincenti*, *luciae* = *alligator*; and *griseus*, *trossulus* = *richardi*). One year later, Günther

(1888) split *Anolis leachii* and placed *lividus* and *sabanus* into *alliceus*. Barbour (1914) disputed both Boulenger's and Günther's taxonomy and elevated many of these variants to full species (*asper*, *speciosus*, *lividus*, *sabanus*, *nubilus*, *griseus*, *vincenti*, *luciae*, *extremus*, *trossulus*). Barbour (1915, 1923) also described four new species (*antigua*, *barbudensis*, *forresti*, *terraealtae*), and Fowler (1918) described one new species (*mayeri*).

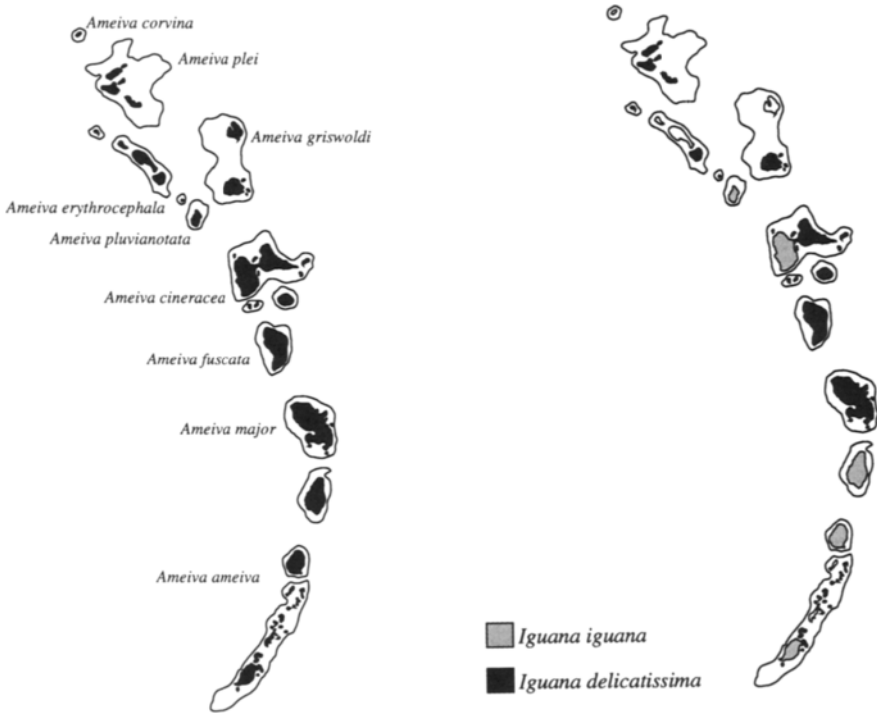
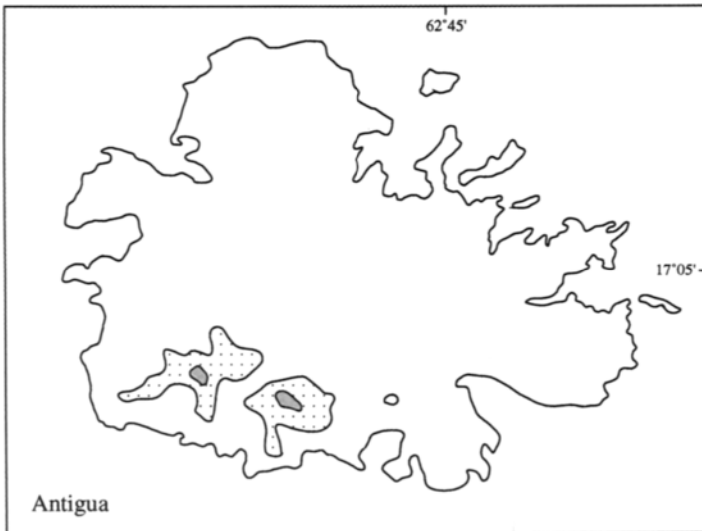


Figure 6.2. Examples of two types of widespread taxa. The two species of *Iguana* are distributed throughout the islands and do not exhibit single-island or island bank endemism. The genus *Ameiva* is also widely distributed in the Lesser Antilles, but is represented by nine endemic species, none widespread like the *Iguana* taxa.

With new material that included live “colorful” specimens, Underwood (1959b) revised the Lesser Antillean *Anolis*. He recognized three groups: the *wattsi* group from the Anguilla Bank, the Antigua Bank and the St. Kitts Bank; the *bimaculatus* group from the islands extending from Dominica northward; and the *roquet* group ranging from

Martinique southward. Once again, many of the full species were relegated to subspecific status or synonymized, reducing the number of species in the Lesser Antilles to nine: group: *bimaculatus* (six subspecies), *marmoratus*, *ferreus*, *speciosus* (two subspecies), *occulatus*, and one undescribed species from Desirade; *roquet* group: *roquet* (three subspecies), *trinitatus* (four subspecies), and *richardii* (two subspecies); *wattsi* group: *Anolis foresti* was placed in the synonymy of *wattsi*.



Although he doubted the conspecific relationship of *gingivinus* to *bimaculatus*, Williams (1962c) nonetheless reviewed the *bimaculatus*

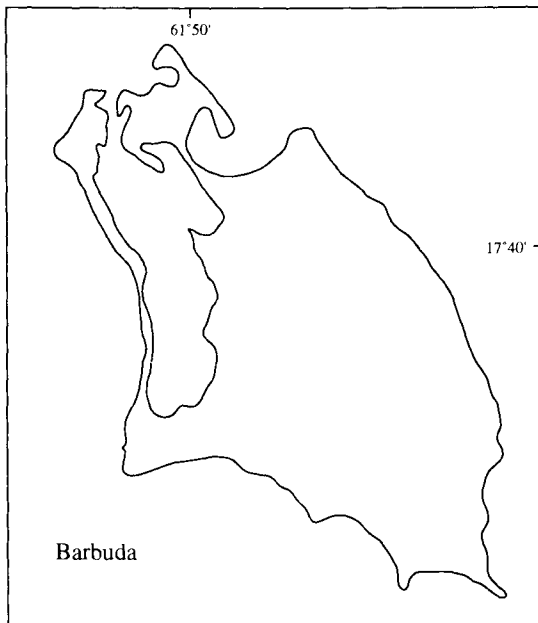
group and retained all trinomials. In addition, he described a new species from Barbuda, *Anolis alter*, and placed it in the *wattsi* group. Lazell (1962, 1964a) subdivided *oculatus* into four subspecies and *marmoratus* into 12 subspecies, including six forms that Underwood (1959b) listed as members of different species.



Lazell (1972) retained only two taxa unchanged nomenclaturally from Underwood (1959b), *bimaculatus leachii* and *oculatus*. Many of Underwood's subspecies were once again elevated to full species (*gingivinus*, *lividus*, *sabanus*, *luciae*, *richardi*, *griseus*), some of the species that Underwood had synonymized were taken out of synonymy (*extremus*, *aeneus*, *nubilus*), and some subspecies were placed as subspecies of other species. Lazell also synonymized *Anolis alter* with *wattsi*. He followed Etheridge's (1959) arrangement of species groups which recognized only two in the Lesser Antilles, *bimaculatus* with Greater Antillean affinities and *roquet* with South American affinities. Lazell's work substantiated the conclusions on the *roquet* species group by Gorman and colleagues (Gorman and Dessauer 1965, 1966; Gorman and Atkins 1968a,b) which examined karyology and blood proteins. Work by Burnell and Hedges (1990) has verified these conclusions. Starch gel electrophoretic studies (Gorman and Kim, 1976) have shown that the genetic divergence of the northern Lesser Antillean *bimaculatus* group is less than that of the southern roquet group, suggesting that *bimaculatus* is evolutionarily younger.

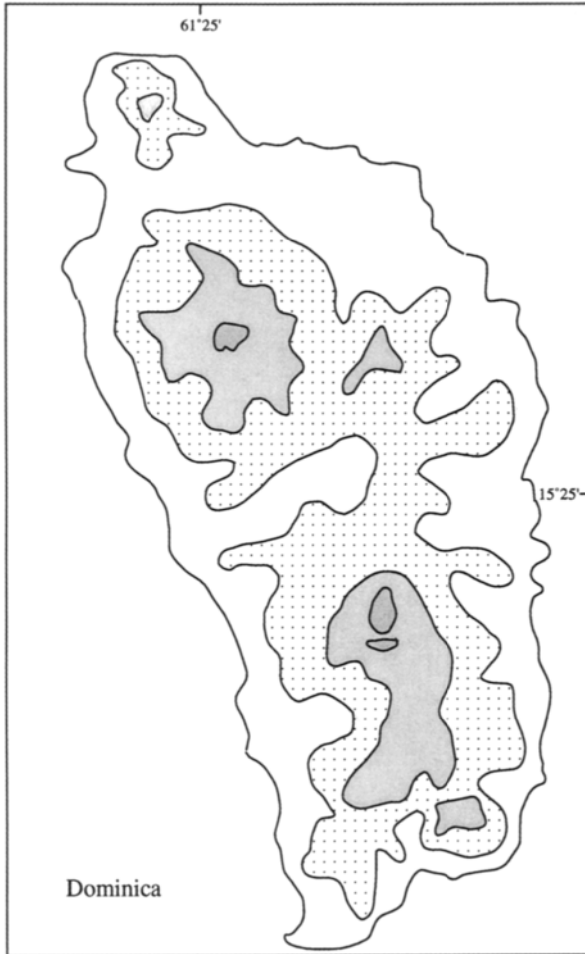
Much of the confusion in Lesser Antillean *Anolis* systematics lies in the fact that these species are extremely variable in morphological characters and coloration. Williams (1959) stated "it is probable that the rather subtle scale characters that distinguish them would have been dismissed...had these species not first been recognized by color or behavior or both." Today, 16 species and 27 subspecies are recognized in the Lesser Antilles, though it should be noted that Malhotra and Thorpe (1991) challenged the validity of subspecies, specifically *oculatus* subspecies. Their research demonstrated that the pattern of variation in characters systems was incongruent, indicating an ecogenetic rather than phylogenetic origin. Each bank of islands has its own endemic *Anolis* except the St. Kitts and Antigua Banks which share two species (*bimaculatus* and *wattsi*; *wattsi* also occurs on the Anguilla Bank), and the Grenada Bank which shares its two species with Trinidad and South America (*richardi*, *aeneus*; Table 6.2). In addition, St. Vincent (and Young Island; Lazell and Sinclair, 1990a) has a population of a species known from Trinidad (*trinitatus*) and two species of *Anolis* (*extremus* and *wattsi*) have been introduced to St. Lucia. Late quaternary fossils of *Anolis* have been found on the islands of Anguilla, Antigua, Barbuda, Guadeloupe, Barbados, St. Eustatius, and St. Kitts. *Anolis* from the latter two islands were from cultural remains (Pregill *et al.*, 1994).

Both species of *Iguana* occur in the Lesser Antilles. *Iguana delicatissima* is a Lesser Antillean endemic, occurring on a number of islands from Martinique northward (Table 6.2). *Iguana iguana* has a more widespread distribution occurring from northern Mexico through Central America to the Tropic of Capricorn in South America and into the West Indies. Barring a few exceptions in the Lesser Antilles, *Iguana iguana* occurs on all islands where *I. delicatissima* does not occur (Table 6.2). Those exceptions are Barbados, Barbuda and Redonda where no *Iguana* have been recorded, and Guadeloupe where both species occur.



Lacerta iguana (= *Iguana*) was one of the first species described from the Lesser Antilles (Linnaeus, 1758). Only ten years later *Iguana delicatissima* was described, along with *Iguana tuberculata* (Laurenti, 1768). Boulenger (1885) recognized Laurenti's two species of *Iguana*, but also placed *rhinolopha* Weigmann as a subspecies of *tuberculata* and gave its distribution as Central America and the West

Indies. Dunn (1934) synonymized *tuberculata* with *Iguana iguana*, and supported Boulenger's assignment of *rhinolopha*. Lazell (1973) retained two species in his review of Lesser Antillean *Iguana*, but did not recognize *rhinolopha* as a subspecies.



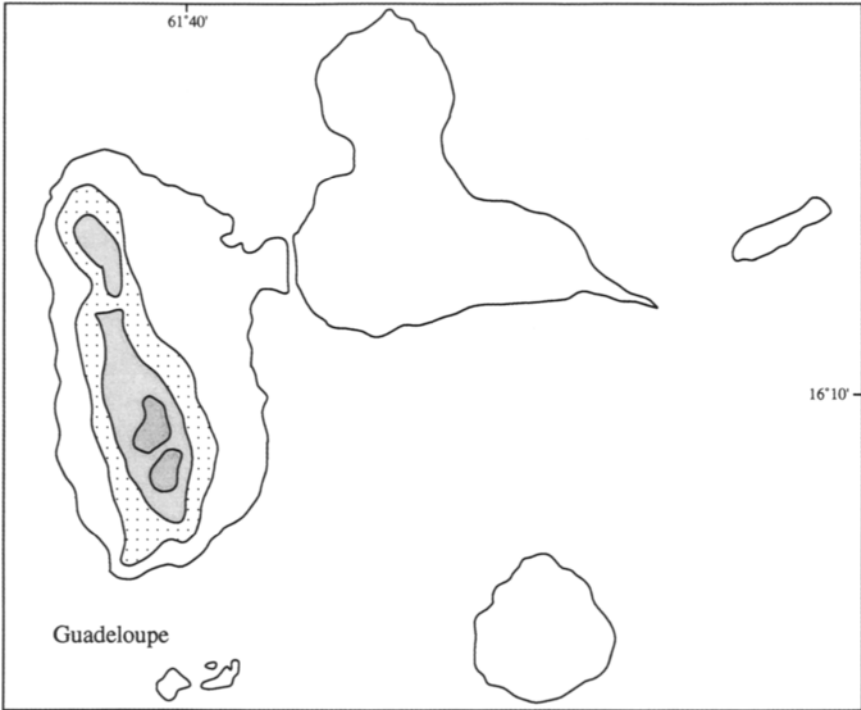
Late quaternary fossils of *Iguana delicatissima* have been discovered in cultural remains on St. Eustatius, St. Kitts and Antigua. Noncultural remains of *Iguana iguana* have been found on Barbados and ? *Cyclura* noncultural remains have been found on Barbuda (Pregill *et al.*, 1994).

Because it is likely that Amerindians transported *Iguanas* between islands, the present distribution of these two species is probably not an accurate reflection of their prehistoric range (Steadman *et al.*, 1984).



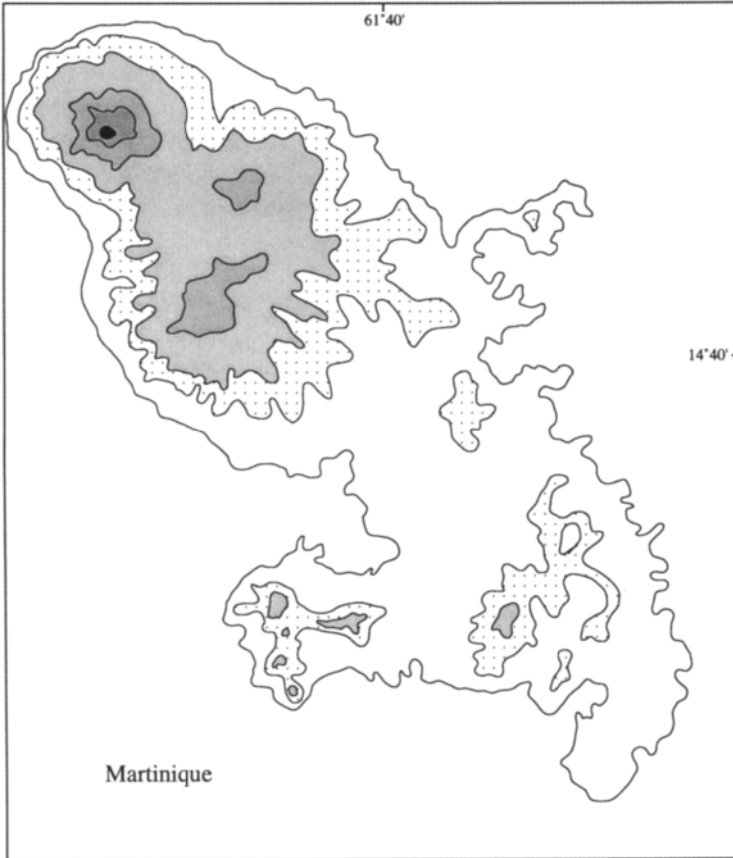
The only tropidurid recorded in the Lesser Antilles is *Leiocephalus*. Duméril and Bibron (1837) described *Leiocephalus herminieri* (three specimens) and Boulenger (1885) added a fourth specimen to the series. Although no specimens have been taken since the original series, Pregill (1992) reported a fifth specimen, collected at the same time, in Rijksmuseum van Natuurlijke Historie (RMNH 2888). There has always

been doubt about the type locality, which has been listed as Martinique. Pregill (1992) stated that “there is nothing that assures us that *Leiocephalus herminieri* ...is endemic to that island.”



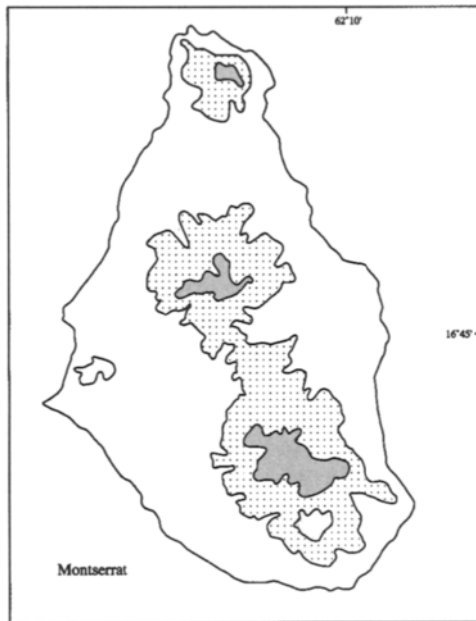
The other species of *Leiocephalus* from the Lesser Antilles is the extinct *L. cuneus*, which is known from fossils taken on Barbuda and Antigua and possibly Anguilla and Guadeloupe (*Leiocephalus* cf. *cuneus*; Pregill, 1992). The occurrence of *Leiocephalus* specimens (fossil

and otherwise) on a number of banks in the Lesser Antilles led Pregill (1992) to state that *Leiocephalus* probably was distributed throughout a good portion of the Lesser Antilles.



There are three genera of medium-sized teiids in the Lesser Antilles. *Ameiva* is widespread and diverse, occurring throughout the Lesser Antilles. The other two (*Kentropyx*, *Cnemidophorus*) are restricted to a single island each. As with *Anolis*, early workers on *Ameiva* described a different species from each of the islands in the Lesser Antilles (Garman, 1887d; Gray, 1838a,b; Cope, 1869), and of the 19 described species, only

11 appear valid today. Barbour and Noble (1915) were the first to do a revision of the genus *Ameiva* which also included the Lesser Antillean species. For the most part, they retained one species per island, though they did report *Ameiva plei* from two islands, St. Martin and St. Barthélemy, and *Ameiva aquilina* from St. Vincent, Grenada and possibly the Grenadines. They also synonymized *scutata* and *analifera* with *plei* and described a new species (*cineracea*) from Guadeloupe. Subsequent to this review, Barbour (1916) described a new species from Antigua (*griswoldi*) and Schmidt (1920) described a new species from Nevis (*nevisana*). This scheme of Lesser Antillean *Ameiva* systematics stood until Baskin and Williams (1966) undertook a reevaluation of the group. They recognized one species for each bank of islands in the Lesser Antilles, except for *Ameiva ameiva* which occurs on two banks, St. Vincent and Grenada. As a result, they synonymized a number of named forms. They also named a new species, *Ameiva vanzoi* from Maria Islands, off St. Lucia. This was later placed in the genus *Cnemidophorus* based on the tongue structure (Presch, 1971). This is the only species of *Cnemidophorus* known from the Lesser Antilles and is currently listed as an endangered species (USFWS 50 CFR 17.11).

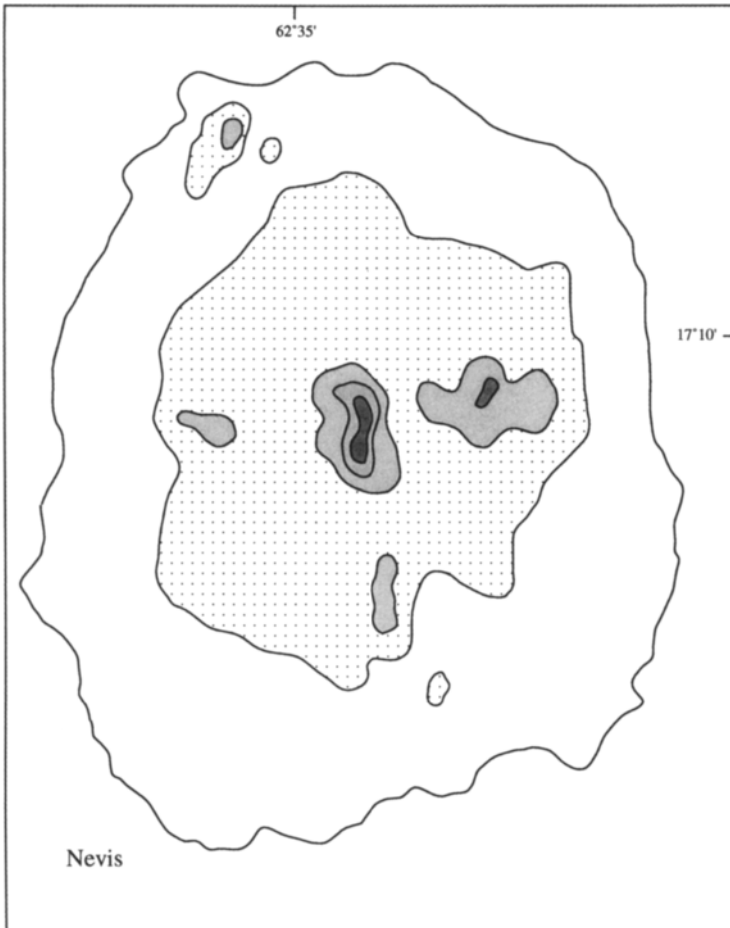


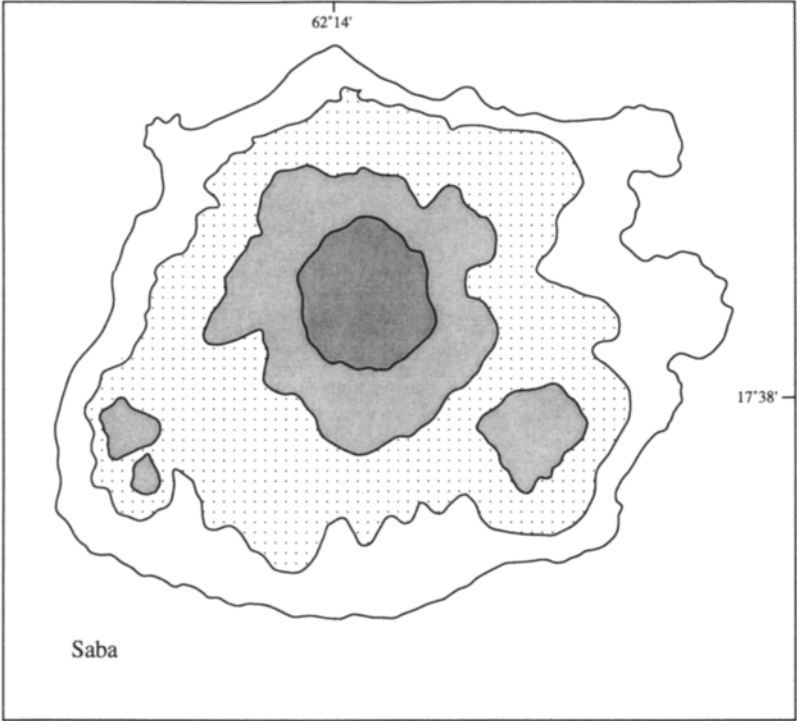
Tuck and Hardy (1973) reexamined the type of *Ameiva surinamensis tobaganus* Cope and concluded that it could not have come from Tobago, but probably was from St. Vincent or Grenada. Therefore, *Ameiva ameiva aquilina* became a junior synonym of *A. a. tobagana*. In 1992, Censky and Paulson revised the Anguilla Bank *Ameiva* (*Ameiva plei*), and described a second species, *Ameiva corax*, from the tiny island of Little Scrub, off northern Anguilla. This species is one of three melanistic species of *Ameiva* found in the Lesser Antilles. The other two are *corvina* on Sombrero and *pluvianotata atrata* on Redonda. All three species are found on small, barren islands with dark substrate. It has been suggested these species exhibit cryptic coloration due to selective predation (Censky and Paulson, 1992). Today, there are 9 extant species of *Ameiva* in the Lesser Antilles (Table 6.2). One species, *major*, is known only from the type series and a second species, *cineracea*, is known from the type series and the fossil record. Both are apparently extinct. *Ameiva ameiva tobagana*, while apparently common in the Grenadines, is considered extinct on St. Vincent (Corke, 1992). Noncultural fossil remains of *Ameiva* have been found on Anguilla, Barbuda, Antigua, and Guadeloupe, whereas cultural fossil remains have been found on St. Kitts, Statia, and Antigua (Pregill *et al.*, 1994). All remains are late Quaternary.

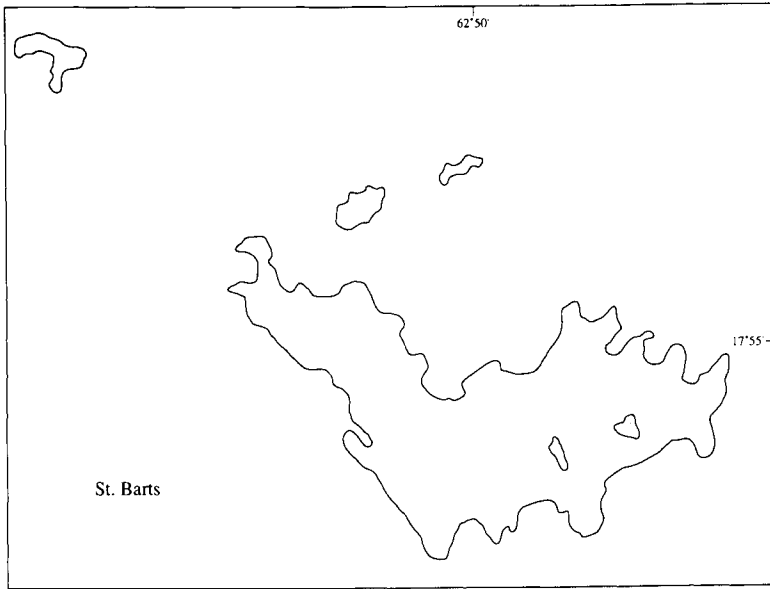
The other medium-sized teiid found in the Lesser Antilles is *Kentropyx*. *Kentropyx* is a South American lizard with only one species reaching the Lesser Antilles (*Kentropyx borckiana*). Cope (1861) was the first to report *Kentropyx* from Barbados. He identified the species as *Centropyx intermedius* Gray. Later, Peters (1869) noted that a species had been described by Merrem (1809) and named "Die Borckische Eidechse", but Peters (1869) was the first to use the binomial *C. Borckiana*. Garman (1887d) described a species, *Centropyx copii*, from Barbados which was later synonymized with *borckiana* (Gallagher and Dixon, 1980). This species was thought to be extinct (Westermann, 1953); however, Underwood (1962) disputed this based on visual records.

There are two genera of microteiids in the Lesser Antilles, *Gymnophthalmus* and *Bachia*. Both occur in the southern Lesser Antilles with *Gymnophthalmus* occurring on all major islands from Guadeloupe southward, whereas *Bachia* occurs only on Grenada and some of the Grenadines. Bocourt (1881) described two species of *Gymnophthalmus* from the Lesser Antilles, *plei* from Martinique and *luetkenii* from St.

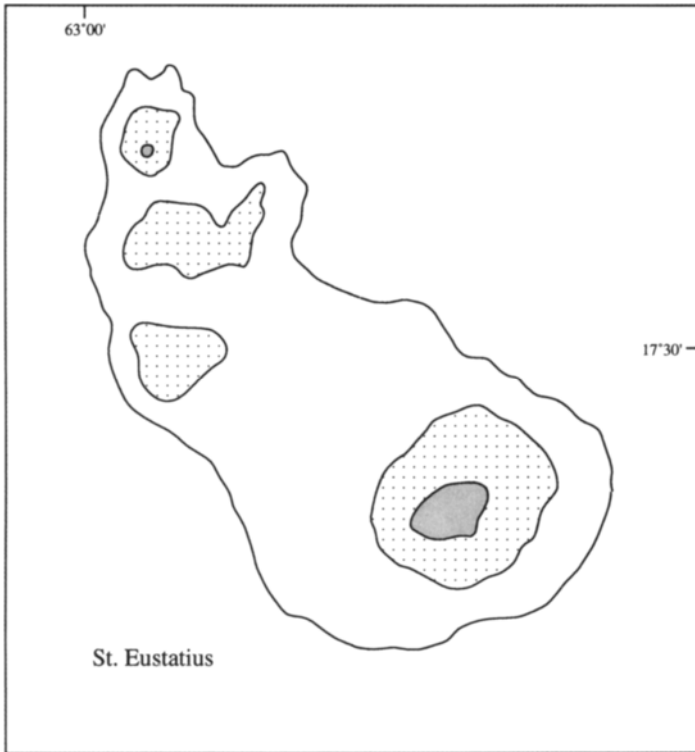
Lucia. Later collectors (Garman, 1887d; Boulenger, 1891), failed to find *G. luetkenii* on St. Lucia, but did find *G. pleei*. As a result, Barbour (1914) stated that the locality for the type of *luetkenii* was undoubtedly incorrect. With fresh materials collected by Schwartz and colleagues, Thomas (1965d) noted some distinctions between island populations and described three subspecies of *G. pleei*: the nominate subspecies from Martinique, *luetkenii* from St. Lucia, and *nesydriion* from Maria Islands off St. Lucia.





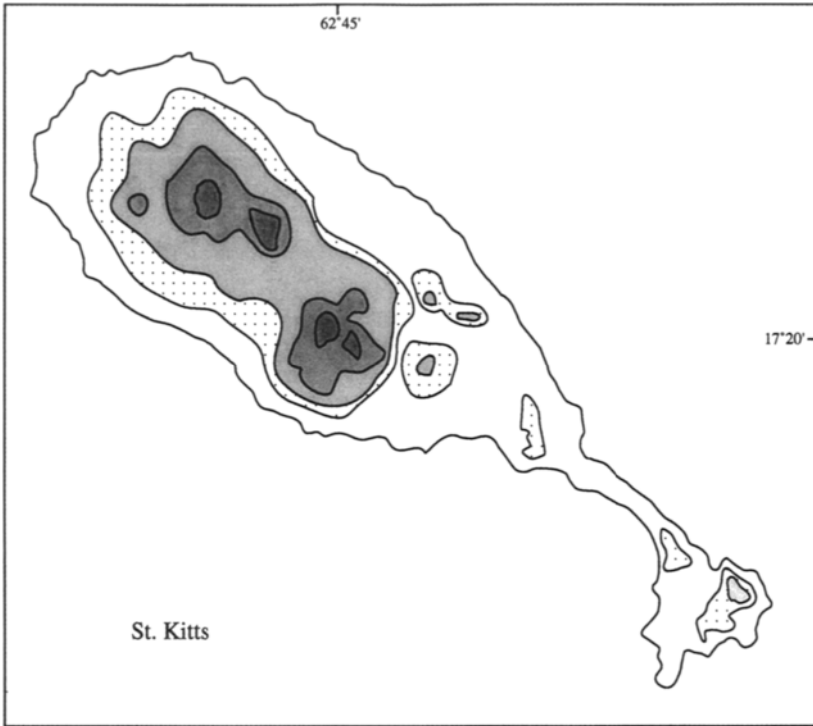


Gymnophthalmus underwoodi was described by Grant (1958) from Barbados. Thomas (1965d) found very little morphological differentiation between specimens from Barbados and Trinidad and the adjacent mainland, but chose to retain the species status of *underwoodi*, and Hoogmoed (1973) concurred by assigning Suriname specimens to the species *underwoodi*. Recently, *G. underwoodi* has been found on St. Vincent, Guadeloupe, and Dominica (Brooks, 1983; tentatively identified as *pleei*, but redetermined by Vanzolini, 1990), Bequia Island in the Grenadines (Lazell and Sinclair, 1990b), and Barbuda (Censky and Lindsay, 1997). While Schwartz and Henderson (1988) considered the Guadeloupe and St. Vincent populations as probable introductions, Corke (1990) disputed this because of the presence of a well-established population on an uninhabited islet off St. Vincent, which he considered an unlikely port for introduction. Thomas (1965d) considered this species to be parthenogenic because no males had, as yet, been found. Interestingly, Brooks (1983) reported one reproductively mature male from Dominica. A South American species of *Gymnophthalmus*, *G. speciosus*, has recently been reported from the southern Lesser Antillean island of Grenada (Vanzolini, 1990). This species occurs from northern South America to Central America.



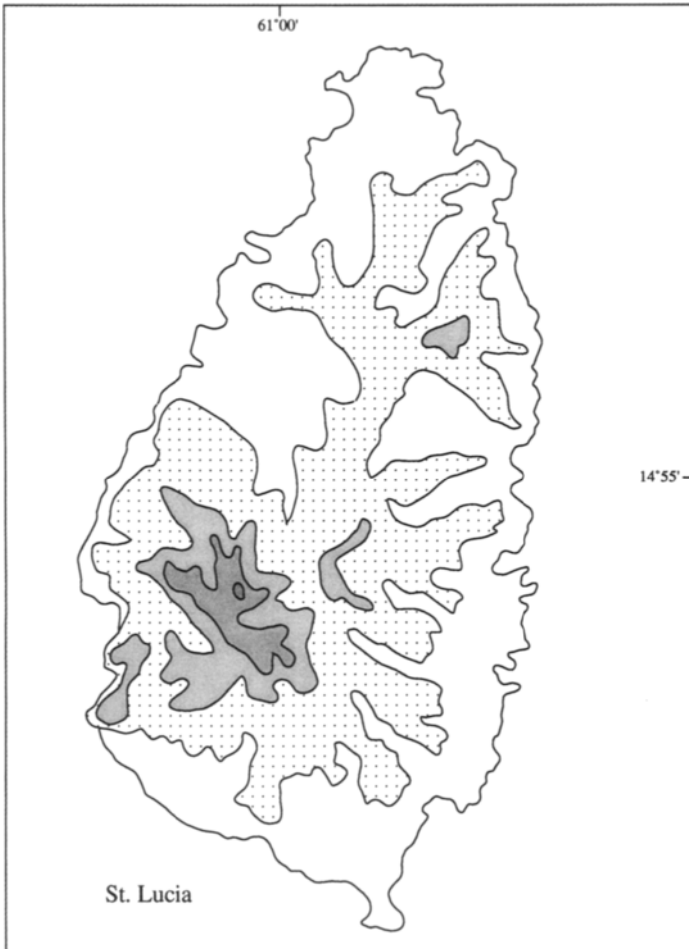
Garman (1887d) was the first to report *Bachia* (= *Scolecosaurus*) from the Lesser Antillean island of Grenada, and he referred his single specimen to *Scolecosaurus cuvieri*. Barbour (1914), with additional specimens from Grenada, noted their distinctiveness and described them as *Scolecosa[ur]us alleni*. Later, Barbour (1933) described a subspecies of *alleni* from the Grenadines, *Scolecosaurus alleni parviceps*. Vanzolini (1961) considered the distinctions between *Scolecosaurus* and *Bachia* to be inconsistent and placed *Scolecosaurus* in the synonymy of *Bachia*. In a review of southeastern Caribbean microteiidids, Thomas (1965d) placed *Bachia alleni parviceps* in synonymy with *B. alleni alleni*. Dixon (1973b) then revised the genus *Bachia* placing *alleni*, along with 5 South American species, in synonymy with *heteropa*, based on the presence of lateral hexagonal scales, 4 digits on forelimbs, and the presence of interparietal and frontonasals. The specimens from Grenada, Grenadines,

and Tobago were placed in the subspecies *Bachia heteropa alleni*. No fossil remains of microteiidids have been found.



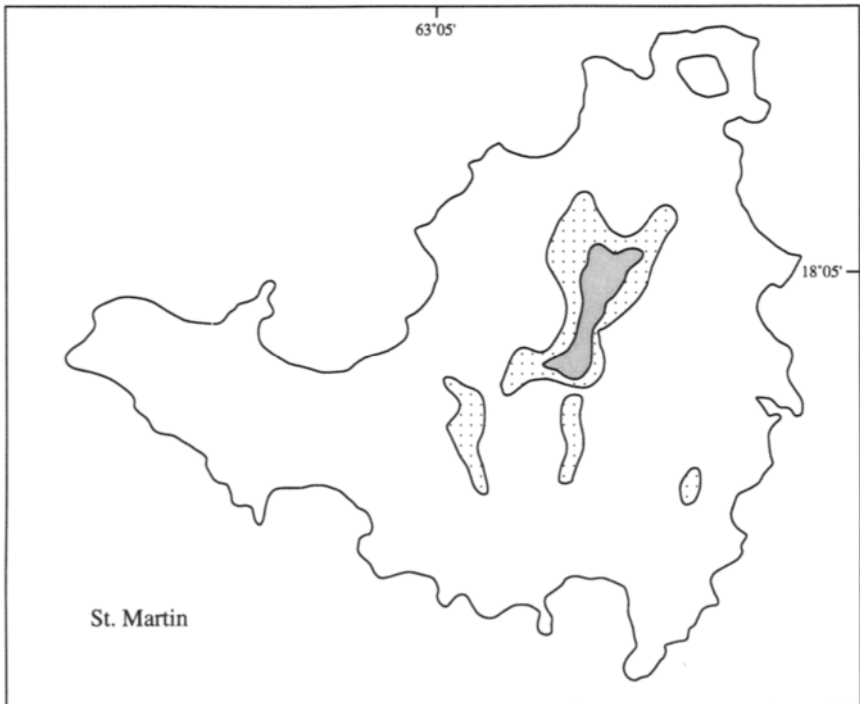
There are four extant genera of geckos in the Lesser Antilles (*Hemidactylus*, *Thecadactylus*, *Sphaerodactylus*, *Phyllodactylus*), one recently introduced species (*Gecko*), and one extinct species (*Gonatodes*). Two species of *Hemidactylus* are found in the Lesser Antilles, the widespread (*H. mabouia*) and the less widely distributed *H. palaichthus* from the southern Lesser Antillean islands. *Hemidactylus mabouia* is found on all major islands, with the exception of St. Barthélemy and Barbuda; a specimen from Redonda was photographed by K. Lindsay and the photo is deposited at Carnegie Museum. Though Du Tertre first reported "*Hemidactylus*" from the Lesser Antilles in 1654 (Kluge, 1969), the species was not described until 1818 by Moreau de Jonnés as *Gecko mabouia*. However, Duméril and Bibron (1836) were the first to use the name *Hemidactylus* for this species. Moreau de Jonnés noted that this species was distributed widely in the Greater Antilles,

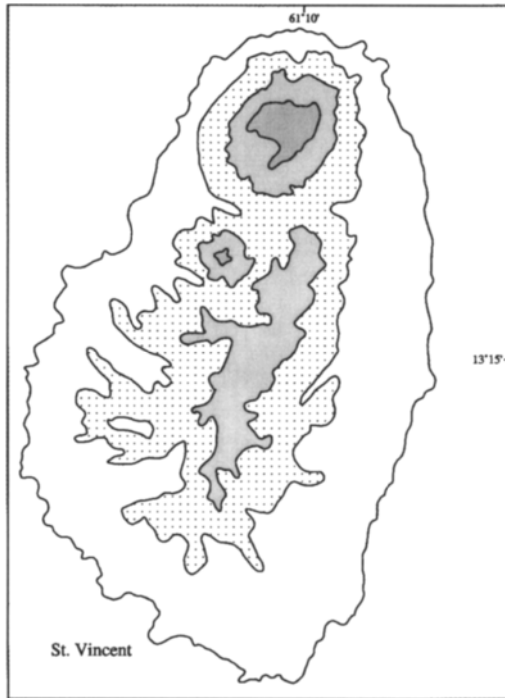
Lesser Antilles, and South America. Stejneger (1904) later restricted the type locality to St. Vincent. Kluge (1969), in a review of the New World *mabouia-brooki* complex, reexamined the type and though it was clear to him that the type specimen of *H. mabouia* was actually a *brooki*, he chose, for nomenclatural stability, to retain the name *mabouia* for the Lesser Antillean radiation. This species is of African origin, and it has been inferred that its presence in the New World is due to transport aboard slaving ships (Grant, 1959). However, Kluge (1969) believed its presence more likely to be due to natural dispersal across an ocean barrier, with subsequent further colonization. He concluded this because of the significant differences between Old World and New World populations and the nonrandom differences in New World populations.



The second species of *Hemidactylus* which occurs in the Lesser Antilles was described as *palaichthus* by Kluge in 1969. The species was known only from Maria Major Island off of St. Lucia until Corke (1987) reported it from the main island of St. Lucia and additional islets. This species is also found on Trinidad and in north central South America. It is an endemic New World species (Kluge, 1969). Noncultural, late Quaternary fossils of *Hemidactylus* cf. *mabouia* have been found on Guadeloupe (Pregill *et al.*, 1994).

Thecadactylus rapicauda is the only species assigned to this genus. It occurs throughout the Lesser Antilles, as well as in South America, and in Central America north to Mexico. The only Lesser Antillean islands that it has not been recorded from are Redonda and Barbados (Garman, 1887a; Barbour, 1914; Schwartz and Thomas, 1975). Non-cultural, late Quaternary fossil remains of *Thecadactylus* have been found on Anguilla, Barbuda, Antigua, and Guadeloupe (Pregill *et al.*, 1994).





Sphaerodactylus is the most speciose genus of geckos in the Lesser Antilles. Eight species and 20 subspecies have been reported thus far (Schwartz and Thomas, 1975; Schwartz and Henderson, 1988, Lazell, 1994). As with *Anolis* and *Ameiva*, the historical tendency to name one species per island has resulted in numerous names, many of which have been relegated to synonymy. Sparrman (1784) was the first to describe this lizard in the Caribbean, with his description of *Lacerta sputator* (= *Sphaerodactylus sputator*) from St. Eustatius. Barbour (1923) considered *sputator* a dichromatic species; however, King (1962a), showed that the purported dichromatism was actually due to the fact that two species, *sputator* and *sabanus* were represented in the syntypes. *Sphaerodactylus sabanus* was described from a series of specimens taken on the island of Saba (Cochran, 1938b). Garman (1887a) described *pictus* from St. Kitts, which King (1962a) placed in the synonymy of *sputator*. Today, *sabanus* occurs on the St. Kitts Bank and Saba, while *sputator* is known from the Anguilla Bank and the St. Kitts Bank. Another species found on the Anguilla Bank is *Sphaerodactylus macrolepis*. Nine subspecies of

macrolepis have been described from the greater Puerto Rico region (Thomas and Schwartz, 1966), but only one subspecies (*macrolepis parvus*) occurs in the Lesser Antilles (King, 1962a).

In 1836, Duméril and Bibron described *Sphaerodactylus fantasticus* from Martinique. However, Barbour (1915) questioned whether the specimens described actually came from Martinique because it was one of those received from Plée whose localities have been questioned (Censky and Paulson, 1992; Pregill, 1992). Barbour restricted the range of *fantasticus* to Guadeloupe based on scale characters and described the species from Martinique as *Sphaerodactylus festus* (Barbour, 1915). King (1962a) agreed with Barbour's locality designation for *fantasticus* and described a new subspecies from Montserrat, *Sphaerodactylus fantasticus ligniservulus*. Thomas (1964) reviewed the group and described six more subspecies, confined to the Guadeloupe complex of islands, and one subspecies on Dominica, though Evans (1989) stated that the subspecies *fuga* was introduced. Boulenger (1885) described a specimen from Antigua as *fantasticus*, but it was later put in synonymy with *Sphaerodactylus elegantulus*. *Sphaerodactylus fantasticus* is known to occur on Montserrat, Dominica, and Guadeloupe and its associated islands and cays.

Sphaerodactylus elegantulus was first described in 1917 by Barbour from one specimen collected on Antigua. Parker (1933), with little reservation, placed a female specimen from St. Lucia in this species, noting that there was little doubt about the locality, and that *elegantulus* must either have recently been introduced onto St. Lucia or it naturally cooccurred with *microlepis*. Because no *S. elegantulus* had been reported from St. Lucia since that time, King (1962a) suggested that it was more reasonable to assume that it belonged to the species *S. vincenti* because of the proximity of St. Lucia to Dominica, and because of the superficial similarity between *elegantulus* and *vincenti*.

Sphaerodactylus vincenti was described by Boulenger (1891) from several specimens from St. Vincent. In his revision of the Lesser Antillean *Sphaerodactylus*, King (1962a) placed *festus* as a subspecies of *vincenti* due to the overlap of many characters. For the same reason, he placed *monilifer* from Dominica, a species originally part of a series listed by Günther (1888) as *S. copii* and later described by Barbour (1921) as *monilifer*, in synonymy with *vincenti festus*. In a review of *S. vincenti*, Schwartz (1964) named an additional five subspecies from

Martinique. He also named a new subspecies of *vincenti* from St. Lucia and resurrected *monilifer* as a subspecies of *vincenti* on Dominica.

Sphaerodactylus microlepis was described by Reinhardt and Lütken (1862) and the type locality was listed as St. Croix. However, Barbour (1921) indicated that the type locality was in error, stating that the species was from St. Lucia. Boulenger (1891) added Dominica as a locality for *microlepis*, and King (1962a) followed suit noting that "Martinique lies between the islands of St. Lucia and Dominica and that future collecting may reveal this species on that island." However, in 1965, Schwartz considered the species to occur on only St. Lucia, stating that there was only one record of one specimen for Dominica and until further specimens were found, it should not be considered to occur on this island. In the same paper, Schwartz described a subspecies *S. microlepis thomasi* from the small Maria Islands, off the southern coast of St. Lucia. Interestingly, the first specimens of *Sphaerodactylus* taken from the Grenadines (Bequia) were as recent as 1989 (Lazell and Sinclair, 1990c) and were described as *Sphaerodactylus kirbyi* (Lazell, 1994).

Sphaerodactylus is widespread in the Lesser Antilles (Table 6.2), with at least one species occurring on each bank of islands except Barbados (a photograph of a *Sphaerodactylus* sp. from Redonda is deposited in the Carnegie Museum and a series of specimens were recently collected; H. Kaiser, personal observation). The only *Sphaerodactylus* that is endemic to a single bank of islands is *S. elegantulus* on the Antigua Bank. *Sphaerodactylus microlepis* may also be a single-bank endemic, occurring on St. Lucia and satellites, with a disputed distribution on Dominica. *Sphaerodactylus sputator* and *S. sabanus* occur on two banks each (Anguilla and St. Kitts, and Saba and St. Kitts, respectively). *Sphaerodactylus vincenti* and *S. fantasticus* occur on a number of banks. The only fossil remains of *Sphaerodactylus* in the Lesser Antilles were found on Anguilla (Pregill *et al.*, 1994). They have not been identified to species.

The relationships among Lesser Antillean *Sphaerodactylus* are based primarily on similarity and proximity; entry into the Lesser Antilles is presumed to have been from South America via stepping stones to the Greater Antilles (King, 1962b). Hass's (1991) molecular study of West Indian *Sphaerodactylus* included a few of the species found in the Lesser Antilles. Her data suggested a single mainland dispersal to Hispaniola, then outward migration from there, including two separate events into

the Lesser Antilles. Her data supported a monophyletic West Indian group. Though she included only three Lesser Antillean species (*sputator*, *sabanus*, and *fantasticus*), she was able to demonstrate a relationship between *sputator* and Greater Antillean species. *Sphaerodactylus fantasticus* and *sabanus* were most divergent and she suggested that these two species along with *elegantulus*, *microlepis* and *vincenti* form a group. Hass' recent study (1996) determined that *sputator* was as different from the Greater Antillean group as are *sabanus* and *fantasticus*. She concluded that all Lesser Antillean species form a group, but cautioned that further study is needed.

There is only one species of *Phyllodactylus* extant in the Lesser Antilles, although several names appear in the literature. Gray described *Phyllodactylus pulcher* from a specimen of unknown origin (= Tropical America; Gray, 1845). Later, Cope (1862a) described a species from Barbados as *Phyllodactylus spatulatus*. Parker (1935), upon examination of type material of both species, failed to find any differences between them and synonymized *spatulatus* with *pulcher*. Dixon (1962) confirmed this decision. Dixon also described a new species of *Phyllodactylus* from Grenada, naming it *Phyllodactylus underwoodi*. Later, Dixon and Huey (1970) synonymized *underwoodi* with *P. ventralis*, stating that locality data of the label on the type was in error [Grenada = New Grenada (Colombia)]. *Phyllodactylus pulcher* occurs only on Barbados.

Gonatodes albogularis was described by Duméril and Bibron in 1836 from a specimen presumably collected by Plée in Martinique. It is the only record of the genus in the Lesser Antilles and, as such, has been doubted (Barbour and Ramsden, 1919). Vanzolini and Williams (1962) stated that collecting had not been intensive enough to claim this unequivocally. To date, no *Gonatodes* have been collected in the Lesser Antilles.

Gecko gecko was introduced on to the island of Martinique about 20 years ago. Henderson *et al.* (1993) report that it is limited to areas of human dwellings.

Anguid lizards are represented in the Lesser Antilles by only one specimen of a single species, *Diploglossus montisserrati* from Montserrat (Underwood, 1964). Although many of the islands of the Lesser Antilles have experienced extensive collecting over the past three decades, no further specimens have been found.

There is only one representative of the family Scincidae in the Lesser Antilles, *Mabuya mabouya* (= "*sloanei* ", R. Crombie, personal

communication), though many more species have been described. Between 1831 and 1887, six species of *Mabuya* were described from the Lesser Antilles, each from a different island or islands [Gray (1831); *aeneus*; Duméril and Bibron (1839); *mabouia*; Cope (1862a); *lanceolata*; Bocourt (1879); *metallica*; Garman (1887d); *dominicana* and *luciae*]. These were all synonymized with *M. mabouya* by Dunn (1936). In this same paper he also reported the species for the first time from St. Martin, Redonda, and Marie Galante, Montserrat. It has since been reported from Anguilla and St. Barthélemy (Schwartz and Thomas, 1975). Barbour (1930b), however, listed the *Mabuya* on Martinique, St. Lucia, St. Vincent, and Barbados as extinct and linked their extinction to the introduction of the mongoose. Corke (1992) stated that they were extinct on Martinique and that he failed to find them on St. Lucia and St. Vincent, suggesting that they are either very rare or extinct. However, there are recent reports of the species from Barbados (Fraser *et al.*, 1990). There have been no fossil remains of *Mabuya* found to date in the Lesser Antilles.

Snakes

The taxonomy of Lesser Antillean snakes has had a very interesting history that is intimately related to human settlement of the islands and the importation of exotic animals. Slave laborers working in sugarcane fields were inexperienced with the fauna on their new island homes, and they would usually kill and mangle a snake on sight for fear of their lives. Bad experiences in mainland colonies made amateur naturalists, such as missionaries, equally wary. Thus, scientific collecting of snakes was not of sufficient volume until the early 1960s (Lazell, 1964b), and a consolidation of the taxonomy was achieved only after that time. Considering the sizes of islands in the Lesser Antilles, however, there exists a fairly high level of snake diversity and endemism. The total of 32 species contains 13 subspecies, and their description spans our entire taxonomic history, beginning with Carolus Linnaeus.

Three taxa (*Boa constrictor*, *B. c. orophias*, and *Corallus hortulanus*) were described by Linnaeus (1758), and these are the only members of the family Boidae found in the Lesser Antilles. Lesser Antillean boids are limited in distribution. Whereas *Boa* occurs on Dominica (subspecies *nebulosa*) and St. Lucia (subspecies *orophias*), *Corallus hortulanus cooki* is limited to St. Vincent, Grenada, and some of the Grenadines. In a review of Lesser Antillean *Boa*, Lazell (1964b)

clarified some of the taxonomy and provided a biogeographical background. He considered the Dominican form *nebulosa* the terminus of a "stepped-cline series," which, by his definition of a subspecies, is a sharply defined peculiar geographic variant that is diagnostically homogeneous (Lazell, 1964b). The subspecies *B. c. nebulosus* was described by Lazell (1964b) in the genus *Constrictor* but was transferred to *Boa* by Peters and Orejas-Miranda (1970). *Corallus cooki* was described by Gray (1842), but later reduced to the rank of subspecies of *Corallus enydris* by Forcart (1951) who also synonymized Barbour's (1914) *Boa grenadensis* with *C. e. cooki*. McDiarmid *et al.* (1996) addressed the confusion surrounding the proper scientific name for this species and placed *B. enydris* in synonymy of *Corallus hortulanus*. As with *B. c. nebulosus*, there are indications that the island populations of *C. hortulanus* are morphologically distinguishable (R. W. Henderson, personal communication). *Boa constrictor* was recovered from the Indian Creek archaeological site on Antigua (Pregill *et al.*, 1988).

Lazell (1964b) removed both Caribbean members of *Bothrops* from the synonymy of *B. atrox* and reinstated them as full species, *B. lanceolata* restricted to Martinique, and *B. caribbaea* confined to St. Lucia. As with his investigations of *Boa*, he capitalized on his own recent collections and the availability of increased numbers of specimens from what might best be considered the "*B. atrox* complex" of South and Central America (Villa, 1984). Garman's (1887e) taxon *Trigonocephalus* is a junior synonym of *Bothrops*, and Lazell (1964b) designated a lectotype to stabilize the taxonomy. Lazell (1964b) also designated Martinique as the type locality for Lacépède's (1789) species *Coluber lanceolatus* and validated the name of the taxon as given by Hoge (1952). Interestingly, the distributions of *B. caribbaea* and *Boa constrictor orophias* are largely coastal and overlap completely, with *B. c. orophias* ranging to slightly higher elevations. Such a distribution may be indicative of relatively recent colonization combined with a slow spread through the inhabited areas that provide sufficient rodent abundance.

Lesser Antillean snakes of the genus *Typhlops* are small, secretive burrowers and consequently relatively rare in museum collections. They have therefore posed a taxonomic challenge that has only recently been addressed in the research of Thomas (1966c, 1974b, 1989) and Richmond (1966). *Typhlops dominicana* was apparently described by Stejneger (1904) on the basis of Boulenger's (1893) description of some

T. richardi (Schwartz and Thomas, 1975). Its nominate subspecies is limited to Dominica (Schwartz and Henderson, 1991). Richmond (1966) described *T. guadeloupensis* as an endemic species from Guadeloupe, but after being listed by Schwartz and Thomas (1975), this form was reduced to subspecies rank within *T. dominicana* (Thomas, 1976). Subsequent check lists use the subspecific form without further comment (Hedges and Thomas, 1989a; Schwartz and Henderson, 1991; Schwartz *et al.*, 1978). Thomas (1966c) described *T. monastus* from Montserrat as well as the subspecies *T. m. geotomus* (occurring on Antigua, Barbuda, Great Bird, Nevis, St. Kitts), differentiating the two taxa by middorsal scale counts. He considered these forms most closely related to the Jamaican *T. jamaicensis* (Thomas, 1989). *Typhlops tasymicris* is restricted to Grenada, and thought to be a relic of waif dispersal from northern South America (Thomas, 1989). Given the difficulty of obtaining series of specimens sufficient for detailed comparisons, a further refinement of the taxonomy of Lesser Antillean *Typhlops* cannot be excluded, especially in view of the as yet uncertain relationships of some key taxa (Thomas, 1989).

Another genus in the family Typhlopidae was recently recorded from the Lesser Antilles. Censky and Hodge (1997) reported *Ramphotyphlops braminus* from the island of Anguilla. Two specimens were collected and were undoubtedly introductions.

Two members of the family Leptotyphlopidae have been recorded from the Lesser Antilles, both in the genus *Leptotyphlops*. The range of the more widely distributed *L. bilineata* was at one point thought to include Barbados, Guadeloupe, Martinique, and St. Lucia (e.g., Barbour, 1937). Its presence on Guadeloupe, as indicated by Duméril and Bibron (1844), is doubtful since there are no recent specimens from that island (Schwartz and Thomas, 1975). Its name also represents a taxonomic curiosity: named *Typhlops bilineatus* and figured in Schlegel (1844), it was described by Duméril and Bibron (1844) and placed into the genus *Leptotyphlops* by Barbour (1914). The occurrence of the second species, *L. tenella*, on Antigua is highly questionable and based only on a single record. Whereas Schwartz and Thomas (1975) and Schwartz and Henderson (1991) list the species for Antigua, Schwartz and Henderson (1985) omitted it from their checklist.

The remaining Lesser Antillean snakes are members of the family Colubridae. Of the four extant species of *Alsophis*, *A. antillensis* has the widest range. Although Schlegel (1837) gave the range for his taxon

Psammophis antillensis simply as "Antilles", its range was restricted to Guadeloupe by Brongersma (1937). The nominate subspecies is now known to occur on Guadeloupe and Marie-Galante (Schwartz and Thomas, 1975), with the subspecies *danforthi*, *manselli*, *sanctonum*, and *sibonius* endemic to Terre-de-Bas, Montserrat, Terre-de-Haut, and Dominica, respectively. Schwartz (1966c) revised the taxonomy of *A. antillensis* by transferring both Parker's (1933) *A. leucomelas manselli* and Cope's (1879) *A. sibonius* to subspecific status under *A. antillensis*. Lazell (1967) placed Barbour's (1915) taxon *sanctonum* into the synonymy of *A. antillensis* at the rank of subspecies. Two other species of *Alsophis* have regionally restricted distributions. *Alsophis rijersmai* is native to the Anguilla Bank (Anguilla, St. Martin, St. Barthélemy), and *A. rufiventris* occurs on Saba and the St. Kitts Bank (Schwartz and Henderson, 1991). *Alsophis rijersmai* was only recently seen on mongoose-infested St. Maarten after a 40-year hiatus (Powell *et al.*, 1992). The former includes in its synonymy Garman's (1887e) *cinereus* (Schwartz and Thomas, 1975). The Antiguan *Alsophis* has had a difficult nomenclatorial history which is still unresolved. Schwartz and Henderson (1991) revived Parker's (1933) name *antiguae* as the specific name for Antiguan *Alsophis* and provided some largely historical evidence (see also Pregill *et al.*, 1988). Parker (1933) described *A. leucomelas antiguae* and subsequently elevated it to full species (Parker, 1936). With no discussion, Schwartz (1966c) reduced it in rank to subspecies. Schwartz *et al.* (1978) remarked that an earlier taxon than Parker's (1933), *punctifer*, described by Martin (1838), might be the appropriate subspecific name for the Antiguan taxon, and that further study was necessary to investigate this question. Henderson (1990) placed the only extant population of *A. antiguae*, on Great Bird Island, into its own subspecies, *A. a. sajdaki*.

The genus *Liophis* is represented by five species and displays an extreme degree of single-island endemism. Each described species occurs basically on a single island, and the distribution of the genus in the Lesser Antilles is nearly continuous along the island chain, reaching from Grenada in the south to Guadeloupe in the north. The detailed work by Dixon (1980, 1981) provided the basis for the transfer of Lesser Antillean *Dromicus* to *Liophis*, reviving Cope's (1862b) genus. *Liophis cursor* is a very rare species that used to occur abundantly on Martinique before the introduction of the mongoose (Lazell, 1967; Henderson and Sajdak, 1986). It was previously thought to be extinct and is now

possibly restricted to a few dozen specimens on Rocher de Diamant, a small satellite island off Martinique (Lazell, 1967). Cope's (1862b) *L. putnami* is a junior synonym of this species. To the north, *L. juliae* occurs on Dominica in its nominate subspecific form, with *L. j. copeae* and *L. j. mariae* native to Guadeloupe and Marie-Galante, respectively. Parker (1933) synonymized Barbour's (1914) *Leimadophis mariae* with *L. mariae*. Three species of *Liophis* occur on southern Lesser Antillean islands. The situation of *L. ornatus* on St. Lucia is similar to that of *L. cursor* on Martinique: the species is now only extant on one small, mongoose free satellite island, Maria Major (Corke, 1987). *Liophis melanotus* exists on Grenada in the Lesser Antilles, but is more widely distributed on the continent (e.g. Trinidad, Tobago, northern South America; Schwartz and Henderson, 1991). The Barbadian populations of *Liophis*, *L. perfuscus*, have a very restricted distribution in the hilly, extreme east central portion of the island which is relatively mongoose free and not affected by sugar cane monoculture.

Four other snakes occur in the Lesser Antilles, and all are scarce in collections as well as in the wild. *Chironius vincenti* is endemic to St. Vincent, where it had been considered extinct until 1988 (Henderson *et al.*, 1988). Taxonomically, Schwartz and Thomas (1975) combined Boulenger's (1891) *Herpetodryas carinatus* var. *vincenti* to the current name to distinguish the St. Vincent and mainland populations. Although the name *C. carinatus* has also been applied to these snakes, only a questionable record from Guadeloupe (Boulenger, 1894) actually places *C. carinatus*, a species widespread on the mainland, on a Lesser Antillean island. The widespread snake *Clelia clelia* occurs in its nominate subspecific form throughout much of the neotropics and until recently was regarded as part of the fauna on Dominica and St. Lucia. Greer (1965) taxonomically flagged the populations on Grenada by placing them into the subspecies *groomei*. However, this subspecific designation was not recognized by Peters and Orejas-Miranda (1970), and has not been further validated. The populations on Dominica and St. Lucia were described by Underwood (1993) as a new species, *Clelia errabunda*. Garman collected three specimens of *Pseudoboia newwiedi* from Grenada in 1883. It has not been taken since and has been considered extirpated (Underwood, 1962). Lastly, *Mastigodryas bruesi* is a native to the southern Lesser Antilles that was described by Barbour (1914) in the genus *Alsophis*. Stuart (1941) placed Barbour's taxon into the genus *Mastigodryas*.

Turtles

There are four species of turtles on the Lesser Antillean islands, though three (*Pelusios subniger*, *Trachemys scripta*, *Trachemys stejnegeri*) and possibly the fourth (*Geochelone carbonaria*) have been introduced. *Pelusios subniger* is an African species that was introduced onto Guadeloupe where it is moderately common (Schwartz and Thomas, 1975). *Trachemys stejnegeri stejnegeri* occurs throughout low elevations in Puerto Rico and it has been introduced to the island of Marie Galante, off of Guadeloupe (Seidel, 1988a; Schwartz and Thomas, 1975). Likewise, *Trachemys scripta* has been introduced onto Guadeloupe (Schwartz and Henderson, 1991).

Populations of *Geochelone* are known from every major island and many smaller islands in the Lesser Antilles, except Barbados and Sombbrero where other extinct tortoises have been reported (Censky, 1988). Most of these populations have been identified as *Geochelone carbonaria*, and those that have been reported as *G. denticulata* were listed before a distinction was made between the two species and may well be *carbonaria*. There have been four hypotheses proposed for *Geochelone* distribution in the Caribbean: (1) natural dispersal, (2) introduction by prehistoric indians, (3) introduction by early European settlers, and (4) recent introduction as escaped pets. None of these can be irrefutably supported by the distributional data, and the current distribution may be the result of a combination of dispersal modes (Censky, 1988). The extinct tortoise, *Geochelone sombreroensis*, was described from remains found on Sombbrero island in the northern Lesser Antilles (Leidy, 1868). Fossil bones of *Geochelone* sp. were found on Barbados (Ray, 1964) and *Testudines* sp. from cultural remains on Barbuda (Pregill *et al.* 1994).

Crocodiles

Crocodiles are not native to the Lesser Antilles. Two species have been recorded in the southern Lesser Antilles. *Caiman crocodylus*, a Central and South American species, is apparently an occasional vagrant to the Grenadines (Schwartz and Henderson, 1991). In addition, in September 1910, a *Crocodylus intermedius* washed ashore in Grenada (Underwood, 1962).

Biogeography

Although there is considerable diversity and high endemism in a variety of genera (e.g., *Anolis*, *Ameiva*, *Eleutherodactylus*, and *Liophis*), a fairly clear picture of the biogeographic origins for some of the Lesser Antillean herpetofauna can be constructed. In general terms, there seem to be two faunal stop gaps, one in the north between the Anguilla Bank and the Virgin Islands and the other in the south between St. Lucia and St. Vincent. The southern gap was already considered important by Lescure (1987), when he observed a progressive reduction of species diversity for amphibians and reptiles between South America, Trinidad, Tobago, and Grenada and St. Vincent in the southern Lesser Antilles. However, as gross diversity decreases when moving north into the island chain, single-island endemism increases, with some of its highest levels reached in the center of the island chain (Table 6.1), at the greatest distance from the diverse herpetofaunas in the Greater Antilles or South America.

All conceivable variations on distributional motif occur for amphibians and reptiles in the Lesser Antilles. Some taxa are endemic only on tiny specks of land (e.g., *Anolis nubilus* or *Ameiva atrata* on Redonda, *Ameiva corax* on Little Scrub Island, *Alsophis antiguae sajdaki* on Great Bird Island), some are native to small islands (e.g., *Anolis sabanus* on Saba, *Diploglossus montisserrati* on Montserrat), some to larger islands (e.g., *Eleutherodactylus euphronides* on Grenada, *Liophis perfuscus* on Barbados), others are Bank endemics (e.g. *Alsophis rijersmai* on the Anguilla Bank, *Ameiva erythrocephala* on the St. Kitts Bank) or regional endemics (both *Eleutherodactylus martinicensis* and *Gymnophthalmus pleei* occur in the central Lesser Antilles), and only *Eleutherodactylus johnstonei* and an *Iguana* species occur on all major islands. Multiple waif dispersal, with subsequent in situ speciation under a variety of selective pressures and extinctions on different islands, is the most encompassing and most easily invoked mechanism to populate the islands with the species we observe today. However, in some cases (e.g., *Eleutherodactylus*, *Iguana*) human-mitigated introductions, either cultural or accidental, are very important to consider because of the nonrandom distribution of some taxa (Pregill *et al.*, 1994; see Kaiser, 1992, for *Eleutherodactylus*).

As a major faunal division in the southern Lesser Antilles, the St. Vincent passage is of unrivaled importance. There are 11 species (one frog, two crocodylians, four lizards, four snakes) that exist in northern

South America and on Grenada and/or St. Vincent whose distribution terminus is the St. Vincent passage. This is the major influx from a more highly diverse fauna. In contrast, at the northern extreme of the Lesser Antilles only *Sphaerodactylus macrolepis* occurs in both the Greater and Lesser Antilles. However, the assumption that the herpetofauna of the Lesser Antilles has its ancestry in both the Greater Antilles and South America is borne out by studies of specific relationships and the Quaternary fossil record (e.g., Hass, 1991; Kaiser *et al.*, 1994b). Given that the occasional and random dispersal event from island to island within the Lesser Antilles still occurs, either by natural means or by human introduction, we can assume that the present-day distribution of taxa is only a vignette in time, and that species ranges in this unique island system may be in continuous flux.

Conservation

The small sizes and limited resources of the islands of the Lesser Antilles, in relation to the human populations, have created serious problems of resource depletion and exploitation. Many of the islands have had a long history of environmental degradation due to numerous factors. Chief among them was the alteration of habitat which began shortly after Europeans arrived with tobacco and sugar-cane. Recently, the tourism industry has taken its toll on a number of islands, with hotel complexes constructed at the expense of natural habitat. While some species appear to do well in altered habitats (*Anolis*, *Hemidactylus*, *Eleutherodactylus*), other species do not (*Iguana*). Seventy-nine populations and/or species of vertebrates no longer occur in the Lesser Antilles since the late Quaternary, and most have been extirpated since European/African settlement (G. Pregill, personal communication).

Another factor that has had an impact on the herpetofauna of the islands is the introduction of exotic animals, particularly mongoose and goat. Barbour (1930) suggested that the mongoose was responsible for the extirpation of a number of populations of reptiles. However, Baskin and Williams (1966) disputed this, stating that there was no simple relationship between reptile population declines and the presence of mongoose. Corke (1992) found that ground lizards and skinks are abundant *only* on mongoose-free islands in the windwards. He did not unequivocally argue that the mongoose is the culprit but stated that more research into anthropogenic causes needs to be done. However, the

mongoose is probably responsible for the extirpation of a number of populations of *Alsophis* (Sajdak and Henderson, 1991). Direct killing of snakes has led to the decline of other species, especially on islands where venomous species occur (Sajdak and Henderson, 1991; Corke, 1992). Goats and other herbivores have had a decimating impact on at least one *Iguana* population (Anguilla, E. Censky, personal observation), while other *Iguana* populations have been impacted by hunting (St. Martin, Nevis, St. Lucia, St. Vincent; Corke, 1992; M. Day, personal communication). *Iguana delicatissima* on Dominica, however, remains healthy even though hunting occurs (M. Day, personal communication).

The loss of biodiversity has not gone unnoticed by many governments in the Lesser Antilles. In the early 1980s, a cooperative effort between the Caribbean Conservation Association and the University of Michigan Wildlife Management Center established ENCAMP. ENCAMP produced a series of technical reports with general guidelines for sustainable management of natural resources. Since then, the Island Resources Foundation, with support from the United Nations Development Program and in collaboration with a number of non-government organizations, has produced in-depth summaries and specific guidelines (Country Environmental Profile) for many of these islands. These reports are based on the information available, scant as it is. A general lack of information on population biology and ecology impedes the efforts put forth by these governments toward recovery and management programs. It has become clear that general systematic, ecology, and survey work (especially on the cays) is needed on many of these islands.

This Page Intentionally Left Blank

West Indian Herpetoecology

Robert W. Henderson¹ and Robert Powell²

¹ Section of Vertebrate Zoology, Milwaukee Public Museum, 800 W. Wells Street, Milwaukee, Wisconsin 53233-1478

² Department of Natural Sciences, Avila College, 11901 Wornall Road, Kansas City, Missouri 64145

The islands, especially the shores, are teeming with lizards of every color, of every variety of marking, and of all sizes.

Ober, 1880

Another individual of this species [Leiocephalus carinatus] seemed to exist in part on food scraps dropped by diners at a hotel in South Andros; this lizard would lurk behind a large curtain, darting out to the appropriate table and back as the occasion arose.

Schoener *et al.*, 1982

Introduction

Aside from the blatantly obvious physical attributes of West Indian islands, the amphibians and reptiles of the region have proven to be ideal tools for biologists with an interest in ecology. Among the alluring qualities of the herpetofauna are its intriguing distribution, wonderful diversity, and the frequently high population densities. New species of West Indian amphibians and reptiles are still being described on a routine basis (especially from Cuba and Hispaniola), the evolutionary species concept (*sensu* Frost and Hillis, 1990) as applied to the West Indian herpetofauna (e.g., Powell, 1993) has provocative implications (e.g., as many as 25 species of *Ameiva* may occur on Hispaniola instead of the currently recognized four species), and West Indian biogeography remains controversial (e.g., Williams, 1989b; Hedges *et al.*, 1992b; Crother and Guyer, 1996). However, research publications that focus on aspects of the ecology of the West Indian herpetofauna are now much more prevalent than taxonomic and systematic publications. For example, Powell and Henderson (1996b)

determined that, through the 1960s, taxonomic publications far outnumbered nonsystematic publications in West Indian herpetology. The 1970s was a transitional decade, and during the 1980s the emphasis had shifted dramatically to nontaxonomic subjects; this trend has continued into the 1990s. The approaches to collecting ecological data and the kinds of ecological data collected also have changed dramatically, from largely anecdotal accounts with varying degrees of reliability to quantification of structural aspects of the habitat and problem-oriented research addressing a wonderfully diverse array of topics ranging from habitat selection (e.g., Chandler and Tolson, 1990), foraging behavior (e.g., Lewis, 1989), dietary analyses (e.g., Floyd and Jenssen, 1983), population densities (e.g., Schoener and Schoener, 1980), and social behavior (e.g., Stamps, 1976) to the ecological context of parasite communities in *Anolis* (Dobson *et al.*, 1992), ultraviolet vision and anole communication (Fleishman *et al.*, 1993), and genetic programming in behavioral ecology (Koza *et al.*, 1992).

Here we briefly review topics that have been important in West Indian herpetoecology (species diversity and distribution), citing relevant literature as sources of additional, more detailed information. Additionally, we have concentrated on several topics (population densities, trophic relations, and ecological versatility) that until now have been addressed only casually (or in reference to only one or two species) and that we view as focal to the ecology of West Indian amphibians and reptiles. Throughout, we have tried to make comparisons between the herpetofaunas of the West Indies and the Neotropical mainland. The taxonomy used herein follows the recent checklist of West Indian amphibians and reptiles by Powell *et al.* (1996a).

History

Species by species summaries of the current state of our ecological knowledge of the West Indian herpetofauna appear in Schwartz and Henderson (1991). They estimated that ecological knowledge was limited to about 5% of the herpetofauna. In retrospect, this estimate is low; about 12% of the herpetofauna has received active attention (i.e., has met one or more of the following criteria: animals were marked and released, portions of the habitat were quantified, thermal responses to the habitat were determined, population densities were determined, prolonged observations of social behavior were made, and/or activity ranges were determined; it precludes fortuitous

observations associated with collecting, stomach content analyses devoid of fieldwork, and/or observations made of strictly captive animals). Not surprisingly (because of their often high densities, diurnal activity, and ubiquity), more lizard species have received attention (about 19% of the lizard fauna) than either frogs (ca. 4%) or snakes (ca. 5%), and 72% of the lizards that met the criteria were *Anolis*.

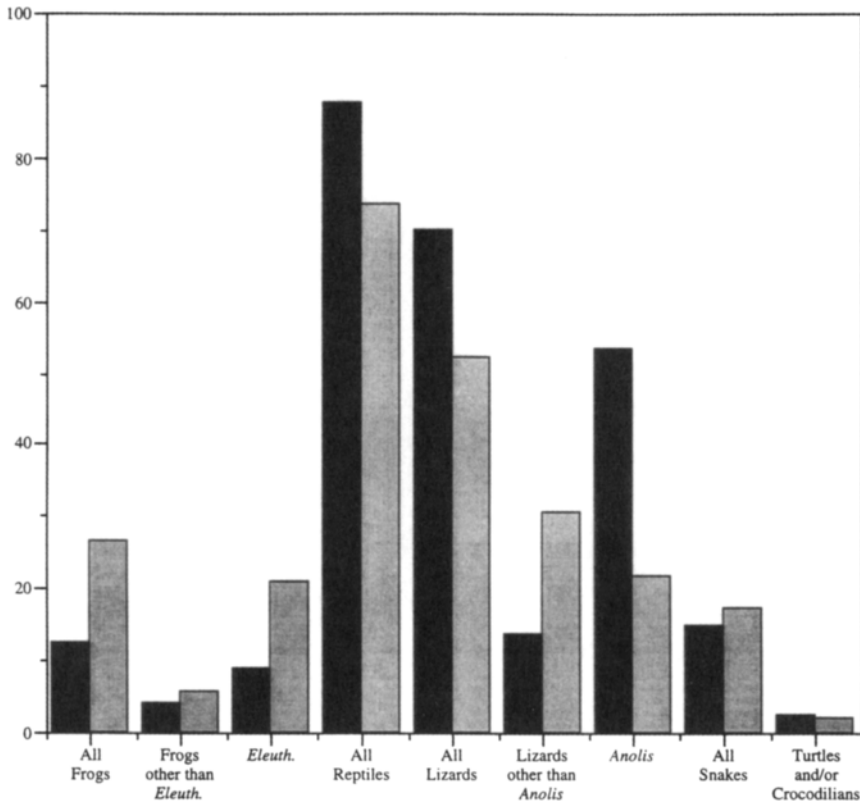


Figure 7.1. The relationship between taxonomic group and the amount of published research in West Indian herpetoecology. Solid bars indicate percentage of publications and gray bars indicate proportion of the herpetofauna the taxonomic group represents. For example, there have been a disproportionate number of publications concerned with *Anolis*, whereas lizards other than *Anolis* have been relatively neglected. *Eleuth.*, *Eleutherodactylus*.

An exhaustive literature analysis is beyond the scope of this chapter. Natural history summaries and species by species reviews of the literature are available in Schwartz and Henderson (1991). Of the ecologically oriented papers published between 1960 and 1990 and

cited by Schwartz and Henderson (1991) in either their Bibliography or Literature Cited sections, 12.6% dealt with frogs and 87.4% with reptiles. Of the frog papers, 68.4% focused on *Eleutherodactylus*. The papers on reptiles included a very high percentage that addressed lizard ecology (79.9%) compared to 17% for snakes and 3% for turtles and crocodylians combined. Figure 7.1 summarizes the taxonomic distribution of the published papers. The number of papers that used anoles as their subject species is highly disproportionate to the rest of the herpetofauna but hardly surprising in light of their ubiquity, density, and visibility. They comprised 64.4% of all papers on reptiles and 53.3% of all papers on frogs or reptiles. Papers with frogs other than *Eleutherodactylus* or with snakes, crocodylians, and/or turtles as their focal organisms have appeared in about the same proportion as reflected by their diversity within the West Indian herpetofauna.

Early Contributions

Although more comprehensive histories of West Indian herpetology appear elsewhere (Powell and Henderson, 1996b; see Chapter 1), here we want to broaden the ecological scope presented therein and to specifically put the study of ecological topics into an historical context. By doing so, we can gain a vantage point for the developmental chronology of an ecological perspective toward the West Indian herpetofauna and see how several key individuals have influenced the advancement of Antillean herpetoecology.

In reviewing the earliest literature that might contain potentially useful observations regarding the West Indian herpetofauna, we determined that the best to be hoped for was accurate anecdotal observations [and we, like Fitch (1987), are advocates of anecdotal observations, excluding them from criteria set earlier only because they do not indicate a concentrated effort to do research on a particular species or problem]. Most early contributions originated primarily from descriptions of habitats and, less frequently, from accounts of the herpetofauna. The letters and diaries of Christopher Columbus, for example, provided descriptions of, in many instances, a West Indies that no longer exists. In 1493, on Hispaniola, he wrote of “trees of a thousand kinds and tall, and they seem to touch the sky” (Major, 1870), describing forests that were long ago sacrificed for ship timbers.

By the early to mid-nineteenth century, natural history exploration in the neotropics produced a number of detailed accounts in the “A Naturalist in ...” genre (e.g., Gosse, 1851; Ober, 1880; Bell,

1889). Certainly the single best example for the West Indies is Gosse's (1851) *A Naturalist's Sojourn in Jamaica*. Gosse's most detailed and appealing accounts deal with Jamaica's rich lizard fauna:

One feature with which a stranger [to Jamaica] cannot fail to be struck on his arrival in the island, and which is essentially tropical, is the abundance of the Lizards that everywhere meet his eye. As soon as he ever sets foot on the beach...he sees on every hand the beautifully-coloured and meek-faced Ground Lizard (*Ameiva dorsalis*) scratching like a bird among the sand.

If he looks into the outbuildings of the estates, the mill-house, or the boiling house, or the cattle sheds, a singular croaking sound above his head causes him to look up...It is the gecko, or Croaking Lizard [*Aristelliger praesignis*], a nocturnal animal in its chief activity, but alwas to be seen in these places, or in hollow trees, even by day.

"The stranger walks into the dwelling-house. Lizards, lizards, still meet his eye. The little anoles ([*A. grahami*, *A. opalinus*] &c) are chasing each other in and out between the jalousies, now stopping to protrude from the throat a broad disk of brilliant colour, crimson or orange, like the petal of a flower, then withdrawing it, and again displaying it in coquettish play...Another is running up and down on the plastered wall, catching the ants as they roam in black lines over its whited surface; and another leaps from the top of some piece of furniture upon the back of the visitor's chair, and scampers nimbly along the collar of his coat.

He lifts the window sash; and instantly there run out on the sill two or three minute Lizards of a new kind, allied to the Gecko, the common Palette-tip (*Sphaeriodactylus* [sic] *argus*).

In the woods he would meet with other kinds. On the trunks of the trees he might frequently see the Venus [*Anolis garmani*], as it is provincially called; a Lizard much like the Anoles of the houses, of a rich grass-green colour, with orange throat disk, but much larger and fiercer: or in the eastern parts of the island the great Iguana [*Cyclura collei*], with its dorsal crest like the teeth of a saw running all down its back.

Gosse (1851) devoted an impressive number of pages to describing *Anolis* behavior (especially of *A. grahami* and *A. opalinus*: "They are particularly numerous in the lieux."), habitat, and distribution; little did he know how important these ubiquitous lizards would become in the study of many aspects of vertebrate ecology and evolution.

Other observations provide an historical context; for example, his premongoose observations on the abundance of the probably now-extinct racer *Alsophis ater*. Gosse described the "Gray Snake," *Natrix capistrata* (= *A. ater*), and gave maximum snout-vent length (SVL) as 16 in.; he described the diet as including *Anolis* and gekkonids; he observed that "though not rare, it can scarcely be called abundant...I have found it scattered in distant and varying localities, in the lowlands and on the mountain-tops." In 1872, the mongoose was introduced to Jamaica (Espeut, 1882); after 1930, specimens of *A. ater* were no longer added to museum collections (Henderson, 1992); by 1940, Grant (cited in Lynn and Grant, 1940) reported that he had not met "anyone who had seen a live specimen in years;" and, as of this writing (April 1996), despite intensive herpetological fieldwork in

Jamaica, no specimen of *A. ater* has been cataloged into a collection in over 60 years.

Toward the end of the nineteenth century, Gundlach (1894) published his *Erpetologia Cubana* which presented interesting and useful anecdotes regarding distribution, abundance, and behavior of some members of the Cuban herpetofauna. Garman, who initiated the long tradition of West Indian fieldwork by herpetologists at Harvard's Museum of Comparative Zoology, produced a series of papers on West Indian reptiles and included field observations in his accounts (e.g., Garman, 1887e).

The Twentieth Century

In the first half of the Twentieth Century, summary papers on various West Indian herpetofaunas were published (e.g., Stejneger, 1904; Barbour, 1914; Barbour and Ramsden, 1919; Schmidt, 1928; Lynn and Grant, 1940; Mertens, 1939, 1940; Cochran, 1941). Each contained interesting observations on aspects of natural history, but many of the observations were not firsthand. Barbour, especially, relied on secondhand accounts or made offhand judgments on the abundance of many species and subsequently has been chastised by several authors (e.g., Curtis, 1946; Lazell, 1972). Notable exceptions to the anecdotal "ecology" of the first six decades of the twentieth century are the publications of Wolcott (1923) on the diets of Puerto Rican lizards, Noble (e.g., 1933) on lizard mating behavior, and Evans (1938) on territoriality in Cuban *Anolis*.

Not until the 1960s did the approach to studying the ecology of West Indian reptiles take several innovative turns. We believe that three catalysts were primarily responsible: (1) Ecology in general was coming into its own as a scientific discipline and no reason existed to preclude the West Indies as a geographic arena, especially considering their proximity to the United States; (2) Albert Schwartz had just started his remarkable inventory of the West Indian herpetofauna and was clearly demonstrating the remarkable species diversity of the area; and (3) Ernest Williams began his multidisciplinary analyses of anole biology.

Indeed, that the animals involved in these ecological innovations were Gosse's "exceedingly interesting" anoles seems altogether appropriate. Preliminarily, Oliver (1948) characterized habitat preferences in four species of *Anolis* on Bimini by perch height. Subsequently, Ruibal (1961) used body temperature and "thermal habitats" to describe ecological differences in five species of Cuban anoles. In 1961, Collette investigated relationships between aspects of

ecology and morphology in several species of Cuban *Anolis*, and he presented a stylized habitat diagram indicating niches of five species. Rand (1962) used perch height to characterize microhabitat in three species of Hispaniolan *Anolis* under captive conditions. Rand (1964b) used morphology, climatic habitat (including body temperatures), and structural habitat (adding the critical dimension of perch diameter to the by now standard perch height dimension) to define the ecological distribution of *Anolis* in Puerto Rico, and he used the term "structural habitat" to define the perch characteristics. Rand (1967a) refined his methods and did a similar analysis of anoles in the Kingston (Jamaica) area, and Schoener (1968) and Schoener and Gorman (1968) took the analyses a logical step farther by adding an examination and quantification of diets. However, the common thread connecting Ruibal, Collette, Rand, Schoener, and Gorman was Ernest E. Williams of Harvard University: all came under his influence early in their research careers.

The Past 30 Years

Ecological research on the West Indian herpetofauna during the past 30 years has been largely problem oriented. Few autecological studies have been produced, but excellent exceptions to this trend include detailed accounts for *Cyclura carinata* (Iverson, 1979), *Anolis lineatopus* (Rand, 1967b), and a Haitian population of *Crocodylus acutus* (Thorbjarnarson, 1988). In addition, via series of papers, several species have been studied more extensively and intensively than others. One frog, *Eleutherodactylus coqui* on Puerto Rico, has been more thoroughly investigated than any other, primarily through the efforts of Margaret M. Stewart and her co-workers (e.g., Stewart, 1985; Stewart and Rand, 1991; Stewart and Woolbright, 1996; Taigen *et al.*, 1984; Townsend and Stewart, 1986; Woolbright, 1985; Woolbright and Stewart, 1987). Similarly, Judy Stamps chose *Anolis aeneus* on Grenada for intensive socioecological research (e.g., Stamps, 1976, 1983, 1987, 1988; Stamps and Tanaka, 1981), and Jonathan Roughgarden and colleagues (summarized in Roughgarden, 1995) addressed the ecology of Lesser Antillean *Anolis*, and various authors have contributed to an understanding of the ecology of *Anolis sagrei*. Much of the content of these papers has been summarized in Schwartz and Henderson (1991). More recently, Ellen Censky (1995a,b, 1996) has conducted detailed field studies with *Ameiva plei*, and Allen Lewis and coworkers have examined intriguing aspects of the natural history of *Ameiva exsul* (Lewis, 1986, 1989; Lewis and Saliva, 1987; Lewis *et al.*, 1996). No snake species to date has

received comparable attention, but Tolson's work (1988, 1996; Chandler and Tolson, 1990) on *Epicrates monensis*, and Henderson's work on *Corallus* in the West Indies (1993a; Henderson and Winstel, 1995; Henderson and Henderson, 1995; Henderson *et al.*, 1996) and field and laboratory research with *Alsophis portoricensis* (e.g., Thomas and Prieto-Hernández, 1985; Rodríguez-Robles and Leal, 1993; J. C. Gillingham, unpublished data), come closest.

The concept of the structural habitat has continued to be an important one in West Indian herpetology. It has provided a convenient means of defining and quantifying strategic habitat variables in arboreal (or scansorial) snakes and lizards, especially in situations in which congenetics occur syntopically. Studies of structural aspects of anole habitats have subsequently proliferated (e.g., Schoener, 1968; Schoener and Gorman, 1968; Rand and Williams, 1969; Schoener and Schoener, 1971a,b; Roughgarden *et al.*, 1981), and the concept has recently been applied to West Indian snakes by Henderson *et al.* (1982) and Henderson and Winstel (1995). Chandler and Tolson (1990) have examined this concept in the context of predator (*E. monensis*) and prey (*Anolis cristatellus*) relations. An important concern in the study of structural habitats, and one that is usually overlooked, is that of habitat availability (Schoener, 1975); species may differ in structural habitat only because the habitats in which they live differ in what is available.

Rand noted that anoles with similar structural habitats also shared morphological similarities; he suggested that these similarities could be the result of being closely related or the result of convergence due to environmental similarities (= ecomorph). Williams (1972, 1983) refined and expanded the ecomorph concept. Ecomorphs are species of different phyletic origin but with similar morphological adaptations to similar niches. Moermond (1979) carried the ecomorph concept farther, including orientation and movements of anoles within a structural habitat matrix. He found that the perch matrix affects the rapidity of movements on surfaces and the frequency of jumps and distances jumped between surfaces; also, species can be separated into jumpers, runners, and crawlers based on movement type and limb and body proportions. Furthermore, Moermond (1979) suggested that availability of structural habitats may help explain the presence or absence of various ecomorphs, and that the local spatial distribution of these microhabitats influences the within-habitat distribution of each species. These data have been independently confirmed in applications to the anoles of other Greater Antillean communities (e.g., Estrada and Rodríguez, 1984; Losos, 1990a,b; Powell and

Russell, 1992) and on the Neotropical mainland (e.g., Pounds, 1988). Losos (1990a,b,c) used the ecomorph concept and found evidence to support the hypothesis that morphology, performance capability, ecology, and behavior have evolved synchronously. In discussing the application of the ecomorph concept and ecomorphology to Neotropical snake assemblages, Cadle and Greene (1993) stressed the need for maintaining an historical perspective when analyzing ecomorphs and ecomorphology. The ecomorph concept has been subsequently applied to West Indian species of *Eleutherodactylus* (Hedges, 1989a) and *Sphaerodactylus* (Thomas *et al.*, 1992) but not nearly as intensively or thoroughly as for *Anolis*. These remain fruitful areas for future research.

Like the structural habitat concept, studies of other aspects of West Indian herpetoecology have undergone a gradual refinement. Research on thermal biology focused on *Anolis* and, again, began with Ruibal (1961), Rand (1967b), and Schoener and Gorman (1968). However, new students, again with ties to Harvard and E. E. Williams (Huey and Webster, 1975, 1976; Hertz, 1981, 1983; Hertz *et al.*, 1993), established ambitious research protocols. Similarly, analyses of diets in frogs (Woolbright and Stewart, 1987; Duer *et al.*, 1992), anoles (e.g., Floyd and Jenssen, 1983; Schoener and Toft, 1983; Pacala and Roughgarden, 1985; Schoener and Spiller, 1987; Bullock *et al.*, 1993; Cullen and Powell, 1994), other lizards (Powell *et al.*, 1990a; Cunningham *et al.*, 1993), snakes (Henderson *et al.*, 1987a, 1988a; Henderson and Crother, 1989), and mixed reptile communities (White *et al.*, 1992) examined seasonal effects, sexual differences, and/or variation in diets by habitat differences.

Some Aspects of the Ecology of the West Indian Herpetofauna

Several aspects of the ecology of the West Indian herpetofauna that we believe have made it especially attractive include: (1) species diversity, (2) distributions, (3) population densities, (4) trophic relations, and (5) versatility. In brief discussions on each of these five topics, we have provided data, compared these with herpetofaunas on the Neotropical mainland, and offered explanations as to why things are as they are in the West Indies. Our treatment is not exhaustive, but the literature cited provides an overview and a starting point for those interested in pursuing particular facets of our discussion.

Origin and Diversity

West Indian islands collectively support one of the richest terrestrial vertebrate faunas in the Western Hemisphere. Considering that the mammalian fauna is relatively small [145 species (80% endemic) historically and about 63 extant species; Hedges, 1996a] and the avian fauna is modest (about 425 species of which only 35% are endemic; Hedges, 1996a), the herpetofauna obviously represents, in respect to number of species, the dominant vertebrate group in the West Indies [622 total described species (169 amphibians and 453 reptiles) of which 572 (92%) are endemic; Hedges, 1996a,b]. Among amphibians, 99% are endemic to the West Indies, and 93% of reptiles are endemic to the area. Seventy-seven independent lineages were identified by Hedges (1996b) and, where a source area within the New World could be determined (42 of 53 lineages), 79% show a South American origin [with smaller contributions from Central America (8/53, 15%) and North America (3/53, 6%)]. Although *Eleutherodactylus* originated by either vicariance or dispersal, and the xantusiid lizard *Cricosaura typica* may have originated by vicariance, "all other lineages appear to have arrived by dispersal during the Cenozoic, and all but nine lineages in the last half of the Cenozoic (30-0 mya)" (Hedges, 1996; but see Page and Lydeard, 1994). Three genera dominate in terms of numbers of species: *Eleutherodactylus* (139 species), *Sphaerodactylus* (79), and *Anolis* (138). Combined, they represent about 57% of the entire herpetofauna. The next most diverse genera are *Typhlops* (represented by 23 species), *Leiocephalus* (23 species), *Celestus* (19 species), *Ameiva* (19 species), *Tropidophis* (13 species), *Amphisbaena* (16 species), *Bufo* (12 species), *Arrhyton* (12 species), and *Alsophis* (11 species). All other genera are represented by fewer than 10 species. Roughgarden (1995) offered an explanation for the reptile-dominated faunas of the Caribbean (which contrast markedly with the avian-dominated faunas of Pacific islands). He believes that

The Caribbean islands are old enough to have accumulated a reptile fauna before birds and mammals diversified. Because the Caribbean Islands remained at warm latitudes, and because the islands are small enough to lead to relatively little predation on lizards, the islands today may show communities that resemble those of the Cretaceous.

Table 7.1. The Number of Genera and Species of Native Frogs, Lizards, and Snakes Occurring at West Indian and Mainland Localities of Varying Area and the Densities of Frogs + Lizards + Snakes at These Localities.

Location	Genera/Species			Species/km ²
	Frogs	Lizards	Snakes	
West Indies (ca 214,305/ km ²)	6/166	22/321	23/105	0.003
Hispaniola (73,147 km ²)	5/64	11/107	10/28	0.003
Dominica (790 km ²)	2/3	6/7	4/4	0.018
Guatemala and Belize (131,852 km ²) ¹	7/77	27/75	56/119	0.002
Costa Rica (50894 km ²) ²	21/120	30/68	59/128	0.006
La Selva (15 km ² , Costa Rica) ³	14/49	15/27	37/47	8.2
Barro Colorado Is. (15 km ² , Panama Canal Zone) ³	13/49	14/26	32/47	8.1
Santa Cecilia (3 km ² , Amazonian Ecuador) ³	30/86	21/30	34/53	56.3
Manu (10 km ² , Cocha Cashu, Amazonian Perú) ³	22/75	19/16	31/31	7.2

¹From Campbell and Vannini (1989).

²From Savage and Villa (1986).

³From Duellman, (1990).

Drawing comparisons of species diversity between the West Indies and the Neotropical mainland is difficult since so many variables can affect the numbers (e.g., area, topography, latitude, vegetation). The larger the areas compared, the more similar the numbers. For example, Guatemala + Belize have 0.002 species/km², and Hispaniola has 0.003 species/km²; Costa Rica has 0.006 species/km² (Table 7.1). As the areas become smaller, however, fewer opportunities for

comparison exist because specific mainland sites have received intensive study and, with the exception of El Verde, Puerto Rico (Reagan and Waide, 1996), no West Indian site has been studied in the manner of, for example, Santa Cecilia (Duellman, 1978) or La Selva (Guyer, 1990). Nevertheless, at specific sites in the West Indies, considerable evidence indicates species-rich herpetocommunities. For example, up to 24 species of anurans, including 20 species of *Eleutherodactylus*, occur together in the Castillon area (ca. 1.0 km²) on the north slope of the distal La Hotte range in Haiti, and 16 anuran species cooccur at Soroa, Pinar del Río, Cuba (Hedges, 1996c). At La Palma in the Cordillera Central of the Dominican Republic, seven species of *Anolis* coexist in a very limited area (Rand and Williams, 1969). Similarly, in the heavily disturbed courtyard of a hotel in the coastal town of Barahona (Barahona Province, Dominican Republic), Powell and colleagues (unpublished data) encountered 13 lizard species, including five species of *Anolis* (an additional six species, including two *Anolis*, could be added if the area within 5 km of Barahona was included).

In contrast, Duellman (1990), for example, reported 86 species of anurans (including 15 species of *Eleutherodactylus*), 31 lizards and amphisbaenians (but only 6 species of *Anolis*), and 53 snakes at Santa Cecilia (Upper Amazon of Ecuador); Donnelly and Guyer (1994) encountered 23 frog species at a single pond at La Selva (Costa Rica), and eight species of *Anolis* also occur there (Duellman, 1990). Surprisingly, only ten species of *Anolis* are recorded from the entire region of Amazonian Brazil (Avila-Pires, 1995).

Inventories of herpetofaunas in the West Indies indicate that, despite their richness, they often lack the diversity found on the Neotropical mainland (especially in rain forest habitat). However, these communities more than compensate for this lack of comparative species diversity with spectacular numbers of individual animals.

Distribution

Although many factors may be involved (e.g., physiography and proximity to other land masses), larger West Indian islands harbor more species of amphibians and reptiles than do smaller islands (e.g., Darlington, 1957; MacArthur and Wilson, 1967). However, few exposed land masses in the West Indies which sustain more than herb-stage vegetation (Lazell, 1972) are devoid of, at least, reptiles, and West Indian islands harbor from one to nearly 200 described species of frogs and reptiles. In examining the distributions of frogs, lizards,

and snakes, we have determined that nearly 40% have multiple-island distributions (MIDs; although perhaps occupying the same island bank), but the differences between frogs and reptiles is considerable and undoubtedly reflects the superior overwater colonizing ability of reptiles. Only 15.8% of the frog species have MIDs, whereas 47.2% of the lizards and 53.6% of the snakes have MIDs. Williams (1969) suggested certain characteristics of successful colonizers. Although anoles were the focus of his discussion, the criteria apply to other taxa as well: (1) inhabitants of open forest or savanna, ecotonal; (2) not a deep shade, rain forest, or montane taxon; and (3) an ecological generalist (not a specialist). More reptilian than anuran species meet these criteria.

In another analysis of factors that affect distributions, Schoener and Schoener (1983a) conducted experiments on 521 small islands in the Bahamas and concluded that the occurrence of lizard species (especially *Anolis*) is related to: (1) island area; (2) structural vegetation diversity; (3) abundance of vegetation and, in some instances, altitude (although the latter may merely reflect a correlation between plant diversity and increasing elevation); (4) distance to the nearest main island (more than distance to the nearest island); and (5) competitive interactions. Unpublished work based on data preliminarily presented in Burns *et al.* (1992) indicates that similar factors apply to the distribution of lizards on the Cayos Siete Hermanos, a small archipelago off the north coast of Hispaniola.

Population Densities

Although the West Indies (especially the Greater Antilles) harbor a rich anuran fauna (primarily *Eleutherodactylus*, which comprises nearly 80% of the known frog species), it is a reptile-dominated vertebrate fauna (>70% of known species), and lizards especially predominate. Not only do they attain high densities [Andrews (1979) suggested that food might be the only restraint on density] but also snakes and other predators prey more frequently on lizards than on invertebrates, frogs, birds, and mammals. Table 7.2 summarizes current knowledge of the densities of West Indian frogs, lizards, and snakes. In comparison to areas outside the West Indies, population densities determined for West Indian species (especially lizards) are singularly amazing.

Table 7.2. Population Densities of West Indian Frogs, Lizards, and Snakes^a.

Species	Island	Density (No./ha.)	Reference
Anura			
Leptodactylidae			
<i>Eleutherodactylus antillensis</i>	Puerto Rico	400	Stewart and Woolbright (1996)
<i>E. coqui</i>	Puerto Rico	100-23,000	Stewart and Pough, (1983); Stewart and Woolbright (1996)
<i>E. hedricki</i>	Puerto Rico	200	Stewart and Woolbright (1996)
<i>E. portoricensis</i>	Puerto Rico	800	Stewart and Woolbright (1996)
<i>E. richmondi</i>	Puerto Rico	100	Stewart and Woolbright (1996)
<i>E. wightmanae</i>	Puerto Rico	400	Stewart and Woolbright (1996)
<i>E. cundalli</i> , <i>E. gossei</i> , <i>E. johnstonei</i> , <i>E. planirostris</i>	Jamaica	Four species combined: 4,635	Stewart and Martin (1980)
Sauria			
Gekkonidae			
<i>Sphaerodactylus clenchi</i>	Hispaniola	4000	Cheng (1983)
<i>S. macrolepis</i>	Guana, B.V.I	1600-2600	Lazell (1991)
<i>S. vincenti</i>	Martinique	8200	Leclair and Provencher (1988)
Polychrotidae			
<i>Anolis aeneus</i>	Grenada	130-1080	Roughgarden <i>et al.</i> (1983)
<i>A. acutus</i>	St. Croix	2000-5600	Ruibal and Philibosian (1974)
<i>A. angusticeps</i>	S. Bimini	2500	Schoener and Schoener (1980)
<i>A. bimaculatus</i>	St. Eustatius	2220-3000	Roughgarden (1995)
<i>A. brevirostris</i>	Hispaniola	800	Moster <i>et al.</i> (1992)

Table 7.2 cont.

<i>A. cristatellus</i>	Guana, B.V.I.	4000-7200	Lazell (1991)
<i>A. cuvieri</i>	Puerto Rico	1.33	Dial <i>et al.</i> (1994)
<i>A. cybotes</i>	Hispaniola	367	Fobes <i>et al.</i> (1992)
	Hispaniola	136-144	Schell <i>et al.</i> (1993)
<i>A. distichus</i>	Abaco	9700	Schoener and Schoener (1980)
<i>A. evermanni</i>	Puerto Rico	2308	Dial <i>et al.</i> (1994)
<i>A. gingivinus</i>	St. Martin	760-12,980	Roughgarden <i>et al.</i> (1983)
<i>A. griseus</i>	St. Vincent	5500	Roughgarden <i>et al.</i> (1983)
<i>A. gundlachi</i>	Puerto Rico	1976	Reagan (1992)
<i>A. oculatus</i>	Dominica	2148	Bullock and Evans (1990)
<i>A. olssoni</i>	Hispaniola	608-650	Smith <i>et al.</i> (1992)
<i>A. pulchellus</i>	Puerto Rico	Up to 20,000	Gorman and Harwood (1977)
	Guana, B.V.I.	1000-1800	Lazell (1991)
<i>A. richardi</i>	Grenada	1630-1800	Roughgarden <i>et al.</i> (1983)
<i>A. sagrei</i>	Abaco	9700	Schoener and Schoener (1980)
<i>A. stratulus</i>	Puerto Rico	25,870	Reagan (1992)
	Puerto Rico	2339	Dial <i>et al.</i> (1994)
	Guana, B.V.I.	3000-5400	Lazell (1991)
<i>A. trinitatus</i>	St. Vincent	5000	Roughgarden <i>et al.</i> (1983)
<i>A. wattsi</i>	Barbuda	3510	Roughgarden <i>et al.</i> (1983)
	St. Maarten	5010	(1983)
	St. Christopher	4970	
	St. Eustatius	4780-9850	Roughgarden (1985)
Tropiduridae			
<i>Leiocephalus carinatus</i>	Bahamas	700	Schoener <i>et al.</i> (1982)
<i>L. schreibersii</i>	Hispaniola	143	Schreiber <i>et al.</i> (1993)
Iguanidae			
<i>Cyclura carinata</i>	Pine Cay, Caicos Island	90.3 (juv.) >31.1 (adults)	Iverson (1979)

Table 7.2 cont.

<i>C. pinguis</i>	Anegada, B.V.I.	2.03	Carey (1975)
Scincidae			
<i>Mabuya mabouya</i>	Dominica	751	Bullock and Evans (1990)
Teiidae			
<i>Ameivachrysolaema</i>	Hispaniola	144	Schell <i>et al.</i> (1993)
<i>A. fuscata</i>	Dominica	379	Bullock and Evans (1990)
Serpentes			
Typhlopidae			
<i>Typhlops richardi</i>	Guana, B.V.I.	Ca. 580	Lazell (1991)
Boidae			
<i>Corallus grenadensis</i>	Grenada	19-61	Henderson and Winstel (1992) and R. Henderson, Unpublished data
<i>Epicrates monensis</i>	Cayo Diablo	>100	Tolson (1988)

*All densities have been extrapolated to number/ha. In some cases, only the maximum density is presented for a given study, even when there was variation in densities from locality to locality.

Frogs

Relatively few data are available regarding frog densities. The highest recorded density for mainland *Eleutherodactylus* is 4586/ha for *E. stejnegerianus* in a Costa Rican cloud forest at 1200 m; *E. bransfordi* occurred at a density of 635/ha in wet forest at 100 m in Costa Rica (Scott, 1976). These data are closer to those for West Indian frog densities than are mainland densities for lizards, but still pale in comparison to >20,000 *E. coqui*/ha in Puerto Rico (Stewart and Pough, 1983; Stewart and Woolbright, 1996).

Lizards

Based on data for areas extralimital to the West Indies, the highest densities recorded are for a gekkonid (*Gonatodes antillensis*) on Bonaire (4200/ha; Bennet and Gorman, 1979), skinks (*Mabuya* spp.) in the Seychelles (up to 3600/ha; Cheke, 1984), and a temperate

skink (1500/ha; Turner, 1960). An anole (*Anolis bonairensis*) on Bonaire reached 1318/ha (Bennett and Gorman, 1979). Species of *Phelsuma*, diurnal geckos that inhabit anoline niches, attain densities of 200-300/ha on the Seychelles Islands (Cheke, 1984). On Guam, Rodda *et al.* (1995) determined lizard densities of up to 20,000/ha, but this included several species and most were introduced. With one exception, all of these records are from tropical islands. More significantly, on the Neotropical mainland, anole densities are low in comparison to those in the West Indies (Table 7.3). Andrews (1979) found anoles to be two or three times more abundant at a site on Dominica than at a site in Costa Rica and also that the most common species in the West Indies reached higher densities than the most common species on the Neotropical mainland. Andrews offered possible reasons for the disparity in mainland versus island population densities (discussed later). Lieberman (1986), in comparing forest leaf litter herpetofauna versus cacao litter herpetofauna at La Selva, Costa

Table 7.3. Population Densities of *Eleutherodactylus*, *Anolis*, and *Ameiva* on the Neotropical Mainland

Species	Location	Density (No./ha)	Reference
<i>Eleutherodactylus</i>			
<i>bransfordi</i>	Costa Rica	635	Scott (1976)
<i>fitzingeri</i>	Panama	2060	Heatwole and Sexton (1966)
<i>stejnegerianus</i>	Costa Rica	4586	Scott (1976)
<i>Anolis</i>			
<i>fuscoauratus</i>	Peru	18.1	Duellman (1987)
<i>humilis</i>	Costa Rica	91	Scott (1976)
<i>limifrons</i>	Panama	1886	Andrews (1979)
<i>nebulosus</i>	Mexico	900	Jenssen (1970)
<i>polylepis</i>	Costa Rica	224	Scott (1976)
<i>Ameiva ameiva</i>			
<i>quadrilineata</i>	Peru	4.6	Duellman (1987)
	Costa Rica	29.7-63.8	Hirth (1963)

Rica, found that the forest site contained greater species diversity but that the cacao site harbored greater densities of frogs and lizards.

Duellman (1987) presented biomass data for a lizard community at a rain forest locality in the Upper Amazon Basin of Perú. Twenty-one species accounted for 1040 g/ha. The only comparable data are those presented by Bullock and Evans (1990) for three widespread lizard species (*Anolis oculatus*, *Mabuya mabouya*, and *Ameiva fuscata*) on Dominica, where the maximum biomass occurred in coastal woodlands and was recorded as 44,700 g/ha. In lowland rain forest habitat on Dominica, the highest recorded biomass was 950 g/ha for *A. oculatus* alone; in comparison, in Amazonian Perú, *A. fuscoauratus* accounted for only 20.3 g/ha. Similarly, *Ameiva ameiva* accounted for 193.4 g/ha in Peruvian rain forest (in clearings) (Duellman, 1987), but *A. fuscata* accounted for up to 25,070 g/ha in coastal woodland habitat on Dominica (Bullock and Evans, 1990).

Snakes

Few data are available for population densities of snakes in the West Indies or on the Neotropical mainland. Scott (1976) calculated a density of 22 snakes/ha of all leaf-litter species combined in a Costa Rican wet forest, and Henderson (1974), based on mark and recapture study at mangrove edge in Belize, determined that *Oxybelis aeneus*, a slender, arboreal lizard eater, occurred at densities of 28-35/ha. At best we can provide a density index for the sake of comparison, but because the indices are based on samples in different kinds of habitat, comparisons must be viewed cautiously. Densities of vertebrate-eating snakes in the West Indies appear to be high when compared to continental data (Turner, 1977; Lillywhite and Henderson, 1993), although high snake abundance on islands outside of the West Indies has also been reported (e.g., Klauber, 1956:529). Density data are available for two arboreal boids in the West Indies: *Epicrates monensis*, a small (ca. 1.0 m SVL) saurophagous species, occurs at densities of >100/ha on a satellite island off Puerto Rico (Tolson, 1988), and *Corallus grenadensis*, a lizard- and mammal-eating species occurred at densities of 19-61/ha in agriculturally disturbed habitats on Grenada (Henderson and Winstel, 1992; R.W. Henderson, unpublished data).

Murphy *et al.* (1994) calculated rates of encounters with snakes in several tropical forest communities: two snake communities in Southeast Asia (Thailand and Borneo) yielded about 1 snake/day; two sites on the Neotropical mainland (Guyana, Ecuador) yielded 0.39 and 0.38 snakes/day; and average daily snake encounters at a site on the

continental island of Trinidad was 2.3. Although comparable data for a West Indian site are lacking, encountering 5-10 snakes/hr in some habitats (not rain forest) is not unusual (but representing only one or two species), snake "markets" (i.e., where money is offered to people living in a particular area in exchange for snakes, and dozens of people may join the hunt) have often yielded over 100 vertebrate-eating snakes within a 24hr period, and 955 specimens of the secretive arthropod-eating *Typhlops pusillus* were accumulated in approximately 36 hr (Schwartz and Henderson, 1991). Henderson and Henderson (1995) encountered up to 4.5 *C. grenadensis*/hr in xeric scrub on Grenada.

The population densities of amphibians and reptiles in the West Indies exhibit a predictable pattern. Arthropod-eating taxa for which we have some index of density (frogs, lizards, and typhlopoid snakes) often occur at high densities (sometimes phenomenally so); frog and lizard predators occur at lower densities, and mammal predators occur at the lowest densities. This pattern is not unique to the West Indies, and herpetofaunas on the Neotropical mainland exhibit similar relationships between diet and relative abundance (e.g., Duellman, 1978). The available evidence indicates that, in general, population densities of amphibians and reptiles in the West Indies, in a variety of habitats but including rain forest, are often many times higher than those on the Neotropical mainland. Assuming density compensation (MacArthur *et al.*, 1972; Case, 1975; Case *et al.*, 1979; Williamson, 1981), higher population densities are anticipated in the West Indies relative to the mainland, but the numbers are nevertheless extraordinary even relative to other islands.

Trophic relationships of the herpetofauna

Excluding turtles and crocodylians (a total of 12 species), the West Indian herpetofauna is composed trophically of 11 herbivores (iguanaid lizards; 1.8% of 622 species), 74 snakes that eat vertebrates (11.9%), and 537 species of frogs, lizards, amphisbaenians, and snakes that prey predominantly on arthropods (86.3%) (Table 7.4).

Trophic Generalists and Specialists

Although placing a species at one end or the other of a broad spectrum ranging from "specialist" to "generalist" is tempting, in reality most species fall somewhere in between. Such labeling of West Indian amphibians and reptiles is difficult for a suite of reasons: (1) Dietary analyses have usually been done without corresponding data on the relative abundance of prey species in the habitat; (2) we do not

Table 7.4. Diets of West Indian Frogs and Squamate Reptiles^a

Genus	No. of Species	Distribution	Diet
Frogs			
Bufonidae			
<i>Bufo</i>	12	G, L	A
Dendrobatidae			
<i>Colostethus</i>	1	L	A
Hylidae			
<i>Hyla</i>	3	G	A
<i>Osteopilus</i>	9	B, G	A, (F)
Leptodactylidae			
<i>Eleutherodactylus</i>	139	B, G, L	A
<i>Leptodactylus</i>	5	G, L	A
Lizards			
Anguidae			
<i>Celestus</i>	19	G	A
<i>Diploglossus</i>	3	G, L	A, (L)
Gymnophthalmidae			
<i>Bachia</i>	1	L	A
<i>Gymnophthalmus</i>	2	L	A
Polychrotidae			
<i>Anolis</i>	138	B, G, L	(P) ^b
Iguanidae			
<i>Ctenosaura</i>	1	W	P, A, (L)
<i>Cyclura</i>	8	B, G	P, A, (L)
<i>Iguana</i>	2	G, L	P
Tropiduridae			
<i>Leiocephalus</i>	23	B, G	A, (L), (P)
Gekkonidae			
<i>Aristelliger</i>	6	B, G	A
<i>Gonatodes</i>	1	G	A
<i>Hemidactylus</i>	4	B, G, L	A
<i>Phyllodactylus</i>	2	G, L	A
<i>Sphaerodactylus</i>	79	B, G, L	A
<i>Tarentola</i>	2	B, G	A
<i>Thecadactylus</i>	1	G, L	A
Scincidae			
<i>Mabuya</i>	2	G, L	A
Teiidae			
<i>Ameiva</i>	19	B, G, L, W	A, (L)

Table 7.4 cont.

<i>Cnemidophorus</i>	2	L, W	A
<i>Kentropyx</i>	1	L	A
<i>Tretioscincus</i>	1	W	A
Xantusiidae			
<i>Cricosaura</i>	1	G	A
Snakes			
Boidae			
<i>Boa</i>	1	L	M
<i>Corallus</i>	1	L	L, M, (B)
<i>Epicrates</i>	9	B, G	L, M, (B)
Colubridae			
<i>Alsophis</i>	11	B, G, L	F, L, M, (B)
<i>Antillophis</i>	2	G	F, L
<i>Arrhyton</i>	12	G	F, L
<i>Chironius</i>	1	L	F
<i>Clelia</i>	2	L	S(?), M
<i>Coniophanes</i>	1		
<i>Darlingtonia</i>	1	G	F
<i>Hypsirhynchus</i>	1	G	L
<i>Ialtris</i>	3	G	L, M, S
<i>Liophis</i>	4	L	F, L
<i>Mastigodryas</i>	1	L	F, L
<i>Nerodia</i>	1	G	Fi
<i>Tretanorhinus</i>	1	G	Fi
<i>Uromacer</i>	3	G	F, L
Leptotyphlopidae			
<i>Leptotyphlops</i>	8	B, G, L	A
Tropidophiidae			
<i>Tropidophis</i>	13	B, G	F, L, M, (B)
Typhlopidae			
<i>Typhlops</i>	23	B, G, L	A
Elapidae			
<i>Micrurus</i>	1	W	L, S
Viperidae			
<i>Bothrops</i>	2	L	M

^aDistribution: B = Bahamas, G = Greater Antilles (including Virgin Islands and Cayman Islands), L = Lesser Antilles, W = western Caribbean. Diet: A = arthropods, B = birds, F = frogs, Fi = fish, L = lizards, M = mammals, P = plants, S = snakes (letters designating prey that appear in parentheses indicate prey categories of lesser importance). Genera that are not native to the region have been omitted.

^bGiant species (>100 mm SVL) only (Bowersox et al., 1994b)

know specifically what most West Indian frogs and reptiles eat and although extrapolating what we know about Species A to Species B is tempting, it is also dangerous; (3) diet for a given species may not be the same from one site to the next, and a species that is a habitat "generalist" will exhibit trophic differences over its range; (4) community studies are virtually absent for the West Indian herpetofauna, and we do not know how, for example, syntopic leaf litter frogs and lizards partition trophic resources; and (5) the terms (specialist versus generalist) used can be misleading because there are different degrees of specialization. For example, the Hispaniolan endemic colubrid snake genus *Uromacer* has three recognized species: *U. catesbyi*, an active forager, *U. frenatus*, an ambush forager, and *U. oxyrhynchus*, another ambush forager. *Uromacer catesbyi* undergoes an ontogenetic shift in diet from primarily lizards (*Anolis*) to primarily hylid frogs (*Osteopilus dominicensis*); *U. frenatus* undergoes an ontogenetic shift from primarily scansorial lizards (*Anolis*) to a high percentage of ground-dwelling lizards (*Ameiva*); *U. oxyrhynchus* eats primarily *Anolis* throughout its life. Of the three species, *U. catesbyi* exhibits the most taxonomically dramatic diet shift (lizards to frogs), *U. frenatus* eats only lizards (a specialist?), but a variety of lizards (a lizard generalist?), and *U. oxyrhynchus* is an *Anolis* specialist. But even this oversimplifies the situation. For example, *U. frenatus* occurs in xeric areas where *Ameiva* occurs at high densities and numbers of *Anolis* are reduced, so its diet might reflect prey availability more than anything else (Henderson *et al.*, 1987c, 1988a). Lieberman (1986:56), in describing a Costa Rican leaf litter herpetofauna, concluded that "all species ate the arthropods in proportions significantly different from those found in the can traps...In a strict sense, none of the herpetofauna species are absolute dietary generalists. Each species has its own position along a continuum from absolute specialist to absolute generalist."

Frogs

Few diet analyses have been conducted for West Indian anurans, and the species most often studied to date have been leptodactylids. The available information, including purely anecdotal data, represent only 12 species (8.7% of the West Indian *Eleutherodactylus*). Jones' (1982) Puerto Rican study indicated that species of *Eleutherodactylus* are opportunistic, taking arthropod prey (including a high percentage of ants) in about the same proportions that they occur in the habitat. However, a disproportionate number of large prey items were taken. Similar types of studies on the Neotropical mainland indicate that

Table 7.5. Foraging and Diet Characteristics of Some West Indian Frogs and Lizards and Their Relationship to Island Size^a.

Genus/ Island area (km ²)	Foraging Mode	Adaptive Zone	Most Frequently Consumed Prey	Reference
Frogs				
Bufo				
<i>Bufo</i> 111,463	S	G	Hymenoptera (Formicidae)	Sampedro Marin and Fundora (1982)
Hylidae				
<i>Osteopilus</i> 111,463	S	S	Isopoda, [Orthoptera]	Meshaka (1996)
76,193			Coleoptera	Duer et al. (1992)
207			Diptera [Orthoptera]	Meshaka (1996)
185			Orthoptera	Meshaka (1996)
Leptodactylidae				
<i>Eleutherodactylus</i> 76,193	S	LG	Hymenoptera (Formicidae), [Coleoptera], [Orthoptera]	Howard <i>et al.</i> (unpublished manuscript)
76,193			Hymenoptera (Formicidae)	Howard <i>et al.</i> (unpublished manuscript)
11,526			Hymenoptera (Formicidae)	Schwartz and Henderson (1991)
11,526			Hymenoptera (Formicidae)	Schwartz and Henderson (1991)
11,526			Hymenoptera (Formicidae)	Schwartz and Henderson (1991)
11,526			Hymenoptera (Formicidae)	Schwartz and Henderson (1991)
11,526			Hymenoptera (Formicidae)	Schwartz and Henderson (1991)
8,865			Homoptera	Jones (1982)
49			Hymenoptera (Formicidae)	Jones (1982)
49			Areanae	Jones (1982)

Table 7.5 cont

<i>Leptodactylus</i>	S	G		
751			Orthoptera	Brooks (1982)
Lizards				
Gekkonidae				
<i>Aristelliger</i>	S	A		
<0.15			Coleoptera [Orthoptera]	Burns <i>et al.</i> (1992)
<i>Sphaerodactylus</i>	A	G		
76,193			Isoptera	Cunningham <i>et al.</i> (1993)
76,193			Coleoptera	Cunningham <i>et al.</i> (1993)
8,865			Acari, [Araneida], [Collembola], [Isopoda]	Thomas and Kessler (1996)
8,865			Collembola	Thomas and Kessler (1996)
Polychrotidae				
<i>Anolis</i> ("typical")	S	G, A		
<0.15			Hymenoptera (Formicidae)	Burns <i>et al.</i> (1992)
<0.15			Hymenoptera (Formicidae)	Burns <i>et al.</i> (1992)
ca. 10			Hymenoptera (Formicidae)	Schoener (1968)
ca. 10			Homoptera [Diptera]	Schoener (1968)
ca. 10			Diptera	Schoener (1968)
ca. 10			Diptera, [Hymenoptera (Formicidae)]	Schoener (1968)
20			Hymenoptera (Formicidae)	Roughgarden (1995)
20			Lepidoptera larva [Hymenoptera (Formicidae)]	Roughgarden (1995)
311			Hymenoptera (Formicidae)	Schoener and Gorman (1968)
751			Hymenoptera (Formicidae)	Andrews (1979)

Table 7.5 cont

1,102			Hymenoptera (Formicidae)	Schoener and Gorman (1968)
11,424			Hymenoptera (Formicidae)	Rand (1976b)
76,193			Hymenoptera (Formicidae)	Fobes <i>et al.</i> (1992)
76,193			Hymenoptera (Formicidae)	Lenart <i>et al.</i> (1994)
76,193			Hymenoptera (Formicidae)	Lenart <i>et al.</i> (1997)
<i>Anolis</i> ("giant")	A, S	A		
76,193			Lepidoptera (larvae)	Bowersox <i>et al.</i> (1994b)
311			Hymenoptera (Formicidae)	Schoener and Gorman (1968)
Tropiduridae				
<i>Leiocephalus</i>	S	L, G		
<0.15			Hymenoptera (Formicidae)	Burns <i>et al.</i> (1992)
<0.15			Hymenoptera (Formicidae)	Burns <i>et al.</i> (1992)
76			Hemiptera, [Isopoda]	Schoener <i>et al.</i> (1982)
1,544			Lepidoptera (larvae), [Plants], [Coleoptera]	Schoener <i>et al.</i> (1982)
76,193			Hymenoptera (Formicidae)	Schreiber <i>et al.</i> (1993)
76,193			Hymenoptera (Formicidae), [Coleoptera]	Lahey <i>et al.</i> (unpublished manuscript)
Anguidae				
<i>Celestus</i>	A	G		
76,193			Coleoptera, [Orthoptera]	White <i>et al.</i> (1992)
76,193			Dictyoptera, [Lepidoptera]	Cisek <i>et al.</i> (1990)
76,193			Diplopoda	Inchaústegui <i>et al.</i> (1985)
<i>Diploglossus</i>	A	G		

Table 7.5 cont

8,865			[Dermaptera]	Thomas and Kessler (1996)
Teiidae				
<i>Ameiva</i>	A	G		
<0.15			Dictyoptera	Burns <i>et al.</i> (1992)
91			Coleoptera	Censky (1996)
751			Oligochaeta, [Coleoptera]	Bullock and Jury (1990)
8,865			Gastropoda	Lewis (1989)
76,193			Dictyoptera, [Diptera]	Powell <i>et al.</i> (1989)
76,193			Isoptera, [Coleoptera larvae and adults]	Sproston <i>et al.</i> (unpublished manuscript)
76,193			Isoptera, [Coleoptera]	Sproston <i>et al.</i> (unpublished manuscript)v
76,193			Isoptera, [Orthoptera], [Arachnida]	Sproston <i>et al.</i> (unpublished manuscript)

*Under "Foraging Mode" A = active and S = sit-and-wait; under "Adaptive Zone" A = arboreal/scansorial, G = ground-dwelling, and LG = largely ground-dwelling. Usually only the orders that contribute the most prey items are listed. Prey orders in brackets indicate that they occurred nearly as frequently as the order listed first.

considerable interspecific variation exists in *Eleutherodactylus* diets (Lieberman, 1986). The data addressing opportunism are equivocal. According to Toft (1980), a disproportionate number of large prey items were taken by Panamanian species, which are characterized as being in a "non-ant specialist" guild. In contrast, Lieberman's (1986) Costa Rican study indicated that species of *Eleutherodactylus* took a wide range of prey sizes, consumed ants in proportion to their occurrence in the leaf litter, and should be considered generalists.

Andrews (1979), in discussing diet differences between mainland and West Indian anoles, suggested that ants in the diet of West Indian species of *Anolis* was an indication that island anoles were food limited relative to their mainland counterparts (but see below). This hypothesis could also be applied to *Eleutherodactylus*, but an alternative hypothesis is that West Indian frogs and reptiles are, in

general, eurytrophic, their trophic plasticity contributes to their high densities, and they are eating those prey items (e.g., ants) which they most frequently encounter. In a Puerto Rican study of *Eleutherodactylus coqui*, Woolbright and Stewart (1987) found that prey size was correlated with frog size (large frogs ate one or two large prey items, whereas smaller frogs ate several small items), and that foraging success was reduced during the dry season. In contrast, Lieberman (1986) found that arthropod abundance peaked at the end of the dry season at La Selva, Costa Rica (forest and cacao), suggesting that foraging success for leaf litter herpetofauna (including several species of *Eleutherodactylus*) would also peak.

In contrast to *Eleutherodactylus*, members of the endemic hylid genus *Osteopilus* (*dominicensis* and *septentrionalis*) prey often on coleopterans and orthopterans and these orders contributed substantially to the total volume of prey (Table 7.5); ants are virtually absent from their diets (Duer *et al.*, 1992; Meshaka, 1996). Sit-and-wait foragers in the bufonid genus *Bufo* [formerly *Peltophryne*; *guentheri* (Parsons, 1995) *lemur* (Schmidt, 1928), *longinasus* (Valdés de la Osa and Ruiz García, 1980), *pellocephalus* (Sampedro Marín and Torres Fundora, 1982)] also include a large number of ants in their diets (up to 84% by frequency of occurrence). Similarly, on the mainland, the diet of *Bufo haematiticus* at La Selva (Costa Rica) included a high proportion of ants (88%) in its diet, as did *Dendrobates pumilio* (63%) and *Gastrophryne pictiventris* (85%).

Lizards

Not surprisingly, most of what we know about the diet of West Indian lizards is based on *Anolis*. Andrews (1979) compared sites on the Central American mainland and on Dominica. She found that: Small items, particularly ants, are relatively more abundant in island than mainland habitats and more abundant in the diets of island than mainland anoles as well. The large numbers of prey in the stomachs of island anoles is a necessary consequence of small average prey sizes. Thus, a comparison of diets suggest that while mainland anoles have the "option" of feeding on large, high-yield prey items, island anoles do not.

She concluded that indirect evidence indicated "that anoles on islands are food limited relative to anoles on the mainland." Contrary to Andrews' conclusions, work by Guyer (1988a,b) on the Neotropical mainland has shown that food does indeed limit anole (*A. humilis*) population density: the numbers of adult males and females were observed to increase in the presence of supplemental food.

Subsequent work has illustrated further the importance of ants in the diets of West Indian anoles (e.g., Floyd and Jenssen, 1983; Burns et al., 1992; Table 7.5). Within the anoline fauna, exceptions to this rule are the so-called giant species (SVL > 100 mm), which prey most often on lepidopteran larvae, coleopterans, orthopterans, and fruit (Andrews, 1979; Bowersox et al., 1994b). We are uncertain why ants represent an inferior food source (Andrews, 1979). Although we know little or nothing specific about most Neotropical frog and lizard diets (mainland and insular faunas), except that most feed predominantly on arthropods, we do know that ants are the most ubiquitous and abundant (numbers and biomass) arthropods in the Neotropics (Wilson, 1991). If most West Indian species are opportunistic predators, predation on the most frequently encountered prey species is expected. Since frog and lizard species in the West Indies are capable of consuming up to 50,000-340,000 prey items/ha/day (Reagan, 1996), an abundant, presumably energetically low-cost food source seems a necessary prerequisite for maintenance of the highest frog and lizard population densities in the world.

When the diets of nonanoline, arthropod-eating lizards are examined (Table 7.5), a different picture emerges. Ants, although often included, do not comprise a significant proportion of the diets. Predation on ants may be a function of foraging strategy. Anoles are typically sit-and-wait foragers (a known exception being the active forager *A. valencienni*; Hicks and Trivers, 1983), as are the other conspicuous ant predators in the West Indies (members of the tropidurid genus *Leiocephalus*). Those species that do not include ants in their diets, or at least do not include them in large numbers, are active foragers, highly arboreal, or are largely nocturnal. Vitt and de Carvalho (1995) found that in the lavrado area of northern Brazil, lizards exhibit a similar, but not unexpected, dichotomy: sit-and-wait species (an *Anolis* and a tropidurid) feed on mobile invertebrates (including many ants), whereas active foragers (teiids) take prey that is predominantly inactive, clustered, and which includes relatively few ants. Since this is a reflection of lizard foraging mode and degree of prey mobility, that the same pattern prevails in the West Indies is not surprising.

Floyd and Jenssen (1983) suggested that "high availability and ease of capture are primary factors influencing the lizards' [*Anolis opalinus* on Jamaica] choice of food." Work by Schoener (1968) and Schoener et al. (1982) has indicated that island size (food per unit area is more abundant on larger islands and there is a possible absence of large insects on small islands) and the presence or absence of

congenerics can influence dietary composition and breadth. In closing, we contend that by eating the most abundant food (usually ants), lizard foraging "territories" require less space, thereby increasing the potential number of lizards that can occupy a particular area or volume of habitat. The opportunistic foraging of many West Indian frogs, lizards, and snakes is probably in large part responsible for the high population densities achieved by many species.

Snakes

Of the 74 known vertebrate-eating snake species in the West Indies, only two (*Chironius vincenti* and *Darlingtonia haetiana*) are frog specialists (Henderson and Schwartz, 1986; Henderson and Haas, 1993), three prey exclusively on lizards (but will accept other vertebrates in captivity), about 60 take frogs and lizards, none are bird specialists although large boids (e.g., *Epicrates striatus*, *Corallus* spp.) and colubrids (e.g., *Alsophis cantherigerus*) do occasionally include them in their diets, and eight or nine prey frequently (but not exclusively) on mammals. Figure 7.2 presents a partial food web focusing on Cuban populations of *Alsophis cantherigerus* and illustrates trophic relationships between members of a herpetofauna. But even with the few components included, the web in Figure 7.2 is oversimplified; *Osteopilus* will eat conspecifics and *Anolis* (Meshaka, 1996), and *Anolis* will eat other *Anolis*, etc.

Among colubrid snakes, Henderson and Crother (1989) found that of 707 prey items, 74.9% were lizards, 20.8% frogs, 1.7% snakes, 1.6% mammals, 0.4% fishes, and 0.3% birds; combined, frogs and lizards accounted for 95.7% of the prey of colubrids compared to only 1.9% for birds and mammals. Invertebrates were virtually absent from the diets of West Indian colubrids, whereas invertebrate predation is common in some (but not all; Vitt and Vangilder, 1983) communities on the Neotropical mainland. Predation on invertebrates by New World colubrids is phylogenetically restricted and species or groups that prey largely or exclusively on invertebrates (e.g., *Atractus*, *Dipsas*, *Geophis*) do not occur in the West Indies (Cadle and Greene, 1993).

At the generic level, *Anolis* comprised 56.8% of the prey items, *Eleutherodactylus* 13.5%, *Ameiva* 9.5%, *Osteopilus* 7.2%, and *Leiocephalus* 4.4%. Among lizards, 75.8% were anoles (and anoles comprise ca. 41% of the lizard species in the West Indies), and among the frogs, 64.6% were *Eleutherodactylus* (which comprise about 79% of the frog species). Although these data would be more meaningful if we had corresponding data on encounter rates for anoles and other

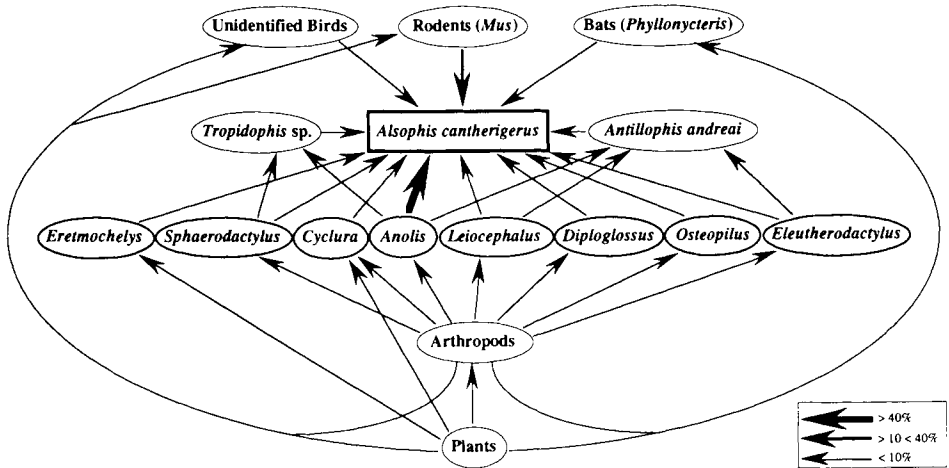


Figure 7.2. A graphic representation of trophic relationships with Cuban populations of the colubrid snake *Alsophis cantherigerus* as the focal species. Width of the arrows indicates percentage each prey group contributed to the diet of *A. cantherigerus*. No attempt was made to quantify the contributions of plants and arthropods to any other group (i.e., arrow thickness has no significance). The diet of *A. cantherigerus* was based on the examination of stomach contents of preserved specimens (Henderson and Sajdak, 1996).

lizards in the West Indies, considering that the colubrid fauna is composed of diurnal foragers and that anoles are diurnal, ubiquitous (occur from ground level to the crowns of trees in a wide range of habitats), and attain extremely high population densities, the high rate of predation on anoles is hardly a surprise. Those snake species that prey on lizards other than *Anolis* tend to be either ground-dwelling (e.g., *Hypsirhynchus ferox*) or ambush foragers (e.g., *H. ferox*, *Uromacer frenatus*), and all exhibit an ontogenetic shift in diet from anoles to nonanoline lizards.

Predation on *Anolis* and, to a considerably lesser extent, *Eleutherodactylus*, was widespread across geographic boundaries in the West Indies; all major islands and island groups supported colubrid snake faunas that preyed predominantly on anoles. Figures 7.3 and 7.4 illustrate the importance of anoles in the diets of West Indian colubrids despite different foraging modes and adaptive zones (Fig. 7.3) and regardless of island size (Fig. 7.4). The latter figure also illustrates how geography affects trophic niche breadths. Similarly, Rodríguez-Robles and Greene (1996) determined that in the Greater Antilles small (<800 mm SVL) and medium-sized (> 800 but < 1200

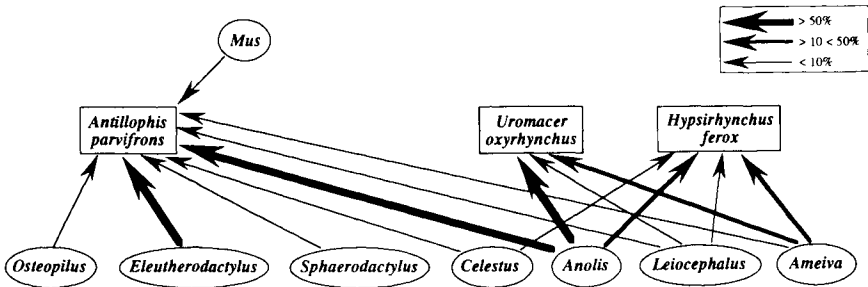


Figure 7.3. Regardless of foraging mode or adaptive zone, anoles are the primary prey of colubrid snakes in the West Indies. *Antillophis parvifrons* is a ground-dwelling active forager (data from Henderson et al., 1987a), *Uromacer oxyrhynchus* is an arboreal sit-and-wait strategist (data from Henderson et al., 1987b), and *Hypsirhynchus ferox* is a sit-and-wait ground-dweller (data from Henderson, 1984). All three species are diurnal, all are endemic to Hispaniola, and all occur sympatrically (and probably syntopically). Arrow width indicates percentage members of each genus contribute to the diet of the three snake species based on examination of stomach contents of preserved specimens.

mm SVL) snakes prey mainly or entirely on frogs and especially lizards, and that most large (> 1200 mm SVL) species eat primarily mammals. Among colubrids on the four largest islands in the Greater Antilles, species richness is positively correlated with prey species richness, and no island supports more than one large species that preys predominantly on mammals (i.e., *Epicrates* spp.).

Henderson and Crother (1989) noted,

On the neotropical mainland, anoles are not nearly as conspicuous, do not occur at such high densities, do not comprise such a major portion of the lizard fauna, and do not contribute as much to the diets of most snake species as they do in the Antilles.

Using data gleaned from Duellman (1978) and Dixon and Soini (1986), of 158 prey items of colubrid snakes, 48.1% were frogs, 21.5% were lizards, and 13.3% were birds and mammals combined; only 3.3% of the prey items were anoles and they comprised only 14.7% of the lizard prey (and 18-21% of the lizard species in the faunas). Similarly, Greene (1988) and Braker and Greene (1994) have noted the high incidence of frogs in the diets of snakes in Costa Rica (“about half of the species feed largely or entirely on frogs” at La Selva; Braker and Greene, 1994). Henderson (1993a,b) described the high incidence of lizards (almost exclusively *Anolis*) in the diets of

West Indian species of the arboreal boid genus *Corallus*, compared to the Neotropical mainland where lizards are nearly absent from tree

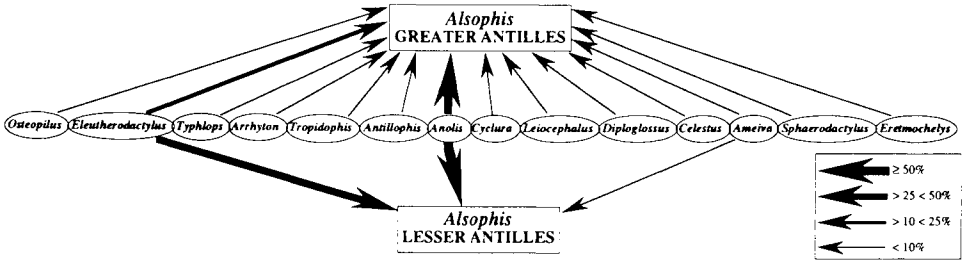


Figure 7.4. Although all West Indian species in the colubrid snake genus *Alsophis* prey heavily on *Anolis*, species occurring in the prey-rich Greater Antilles take a wide variety of frogs and reptiles. Lesser Antillean *Alsophis* are more limited in exploitable frogs and reptiles. Width of the arrow indicates percentage contribution members of each prey genus made to either Greater or Lesser Antillean *Alsophis* diets (based on examination of stomach contents of preserved specimens; Henderson and Sajdak, 1996).

boa diets. Phylogenetic analyses by Rodríguez-Robles and Greene (1996) suggested that size reduction occurred three times within *Epicrates*, once in mainland *E. cenchria* and twice within the Antillean clade. Their hypothesis “that small size evolved in Caribbean *Epicrates* in response to a shift in adult diet to smaller lizard prey” was supported by their analysis.

Trophically, anoles are the most widely exploited vertebrates in the West Indies. Fitch and Henderson (1987) noted that, for *Anolis bahorucoensis*, “every individual...is constantly in danger, from several directions simultaneously, and can survive only by instantaneous escape responses that demand priority over every other kind of activity.” The same may well apply to most other species of *Anolis* in the West Indies [with the possible exception of the giant (>100 mm SVL) species], and Henderson and Crother (1989) suggested that members of no other vertebrate-eating snake fauna of similar diversity prey to such a large extent on members of a single genus of prey organisms anywhere else in the world.

Overall, we know more about the diets of a higher percentage of colubrid snakes in the West Indies than either frogs or lizards. Although several species might be labeled specialists (*Darlingtonia*

haetiana, *Uromacer oxyrhynchus*), we feel that most are best described as opportunistic. Anoles account for more than 50% (by frequency of occurrence) of the prey items in the diets of most West Indian colubrids (Henderson *et al.*, 1988a; Henderson and Crother, 1989; Henderson and Sajdak, 1996) and they also make substantial contributions to the diets of boids and tropidophiids (Henderson *et al.*, 1987a; Henderson and Crother 1989; Henderson, 1993a). Colubrids that prey primarily or exclusively on invertebrates (the “Goo-eaters” of Cadle and Greene, 1993) are absent from the West Indies, as are all trophically “specialized” mainland snake genera. But despite the high incidence of anoles in the diets of West Indian vertebrate-eating snakes, we do not consider these snakes anole specialists. Species of *Anolis* are, more often than not, the dominant vertebrate species at a given site in the West Indies; this is not true on the mainland (e.g., Duellman, 1987). In 1991, at a disturbed site at Barahona, R. Powell (unpublished data) found about 135 *Anolis* to 19 *Ameiva* and 18 *Leiocephalus*. On Dominica, Bullock and Evans found the maximum density of *Anolis oculatus* at a coastal woodland site to be 25 times greater than maximum density for *Ameiva fuscata*. Conversely, the greatest difference wherein *Ameiva* was the dominant species was 2.1 times greater than for *Anolis*. At a xeric site on Hispaniola where the ground-dwelling, *Ameiva*- and *Leiocephalus*-eating colubrid *Hypsirhynchus ferox* occurred, Powell *et al.* (1996b) encountered 287 *Ameiva*+*Leiocephalus* compared to 107 *Anolis* (2.6 times more primarily ground-dwelling lizards than scansorial anoles). We believe, then, that West Indian vertebrate-eating snakes are exploiting those prey most frequently encountered, based on their adaptive zone (e.g., arboreal or ground-dwelling), foraging mode, and habitat.

Predators of Frogs and Reptiles

Based on published accounts and personal observations, we tabulated a list of over 100 predators of frogs and reptiles in the West Indies, and it is surely far from complete. Nearly 80% of these predators are other frogs and reptiles (Table 7.6). Virtually all the predation on frogs and reptiles by frogs and lizards is infrequent and opportunistic. Species of West Indian frogs and most reptiles that are primarily arthropod predators, but will prey occasionally on vertebrates, include: *Leptodactylus fallax* (Brooks, 1982; Lescure, 1979), *Osteopilus dominicensis* (Duer *et al.*, 1992), *O. septentrionalis* (Meshaka, 1996), *Hemidactylus haetianus* (Bowersox *et al.*, 1994a), *Anolis chlorocyanus* (Bowersox *et al.*, 1994a), *A. cybotes* (Fitch and Henderson, 1987; Lenart *et al.*, 1996), *A. vermiculatus* (González-

Table 7.6. Known Predators (invertebrates, anurans, etc.) of West Indian Frogs and Reptiles (listed by genus) Determined by Stomach Content Analysis or Field Observation.

Genus	Predators						
	Inv.	Anu.	Liz.	Sna.	Cro.	Bir.	Mam.
Anura							
<i>Bufo</i>	X			X			X
<i>Eleutherodactylus</i>	X	X	X	X		X	X
<i>Hyla</i>				X			
<i>Osteopilus</i>		X	X			X	
Sauria							
<i>Hemidactylus</i>				X			
<i>Sphaerodactylus</i>	X		X	X		X	
<i>Anolis</i>	X	X	X	X		X	X
<i>Leiocephalus</i>			X	X			
<i>Cyclura</i>			X	X		X	X
<i>Iguana</i>				X		X	X
<i>Mabuya</i>				X		X	
<i>Celestus</i>				X			
<i>Diploglossus</i>				X			
<i>Ameiva</i>				X		X	X
Testudines							
<i>Trachemys</i>					X		
Amphisbaenia							
<i>Amphisbaena</i>	X			X			
Serpentes							
<i>Typhlops</i>				X		X	
<i>Boa</i>		X					
<i>Corallus</i>							X
<i>Tropidophis</i>				X		X	
<i>Alsophis</i>				X		X	X
<i>Antillophis</i>				X		X	
<i>Arrhyton</i>				X			
<i>Ialtris</i>						X	
<i>Liophis</i>						X	
<i>Uromacer</i>						X	

Inv., Invertebrates; Anu., anurans; Liz., lizards; Sna., snakes; Cro., crocodilians; Bir., birds; Mam.,

Bermuda and Rodriguez-Schettino, 1982), *Leiocephalus barahonensis* (Lahey *et al.*, 1998). *L. schreibersii* (Jenssen *et al.*, 1989), *Cyclura carinata* (Iverson, 1979), *Ameiva chrysoleama* (Sproston *et al.*, 1998), and small *Crocodylus acutus* (Thorbjarnarson, 1988). Snakes and birds represent the most significant predators of frogs and reptiles, although predation by birds may vary from island to island (Wunderle, 1981; Adolph and Roughgarden, 1983, McLaughlin and Roughgarden, 1989). In any case, the list of bird species is certainly inadequate in providing an accurate picture of avian predation pressure. More complete analyses of trophic relationships in West Indian faunas (not restricted to the herpetofauna) appear in Goldwasser and Roughgarden (1993), Roughgarden (1995), Reagan (1996), Reagan *et al.* (1996), and Stewart and Woolbright (1996). All, however, reinforce the importance of anoles in West Indian food webs and analysis of a rain forest community on Puerto Rico indicated that frogs and lizards dominate nocturnal (*Eleutherodactylus*) and diurnal (*Anolis*) webs (Reagan, 1996; Reagan *et al.*, 1996).

Apparently, it is a widely held belief that West Indian anoles receive little predation pressure relative to mainland lizards (McLaughlin and Roughgarden, 1989; Roughgarden, 1995), and Andrews (1979), in discussing the high densities of anoles in the West Indies compared to the Neotropical mainland, commented that island anoles have fewer predators to contend with, noting that fewer snake species occurred at island sites. Certainly this is true, but this does not mean that fewer snakes are present. Almost all West Indian snakes, with the exception of scolecophidians, prey on anoles at some time in

Table 7.6 cont.

mammals.

Note: Predators are listed by species as follows: **Invertebrates:** ant (*Crematogaster brevispinosa*), spiders (*Olios antiguensis*, *Stasnia portoricensis*, *Oligoctenus ottleyi*, and *Avicularia laeta*), amblypygid (*Phrynos longipes*), scorpion (*Tityus obtusus*), centipede (*Scolopendra alternans*), crab (*Epilobocera situatifrons*); **Anurans:** *Eleutherodactylus coqui*, *E. wightmanae*, *Leptodactylus fallax*, *Osteopilus dominicensis*, *O. septentrionalis*; **Lizards:** *Anolis coelestinus*, *A. cristatellus*, *A. cybotes*, *A. equestris*, *A. grahami*, *A. gundlachi*, *A. lineatopus*, *A. richardi*, *A. stratulus*, *A. vermiculatus*, *Leiocephalus carinatus*, *Cyclura carinata*, *Ameiva exsul*, *A. plei*, *A. taeniura*, *Sphaerodactylus klauberi*; **Snakes:** virtually all boids, tropidophiids, and colubrids (about 70 species); **Crocodylians:** *Crocodylus acutus*, *C. rhombifera*; **Birds:** *Buteo jamaicensis*, *B. platypterus*, *B. ridgewayi*, *Falco sparverius*, *Tyto alba*, *Otus nudipes*, *Egretta thula*, *Florida caerulea*, *Bubulcus ibis*, *Coccyzus minor*, *Saurothera merlini*, *S. vieilloti*, *Crotophagous ani*, *Tyrannus dominicensis*, *Melanerpes portoricensis*, *Myiarchus tyrannulus*, *Mimus gilvus*, *Todus mexicanus*, *Turdus nudigenis*, *T. plumbeus*, *Margarops fuscatus*, *Ferninia cerverai*, *Quiscalus lugubris*; **Mammal:** *Didelphis marsupialis*, *Rattus rattus*, *Felis catus*, *Canis familiaris*, *Herpestes javanicus*.

their life history, and many prey predominantly or exclusively on anoles throughout their lives. In addition, some snakes, especially small colubrids and boids, may occur in very high densities in the West Indies and be the major predators of frogs and lizards. Similarly, although islands harbor fewer potential prey species for snakes, it does not mean that they harbor fewer potential prey items (Henderson and Bourgeois, 1993; Henderson and Sajdak, 1996).

Versatility

A herpetofauna derived largely from successful colonizers might be expected to exhibit the colonizing characteristics of its ancestors (Williams, 1969; see also Distribution). Because successful colonizers must be ecological generalists capable of surviving, if not thriving, in varied habitats, the question arises whether or not West Indian amphibians and reptiles include more species exploiting a broad spectrum of ecological variables than members of the mainland Neotropical herpetofauna.

Such data are lacking for most of the West Indies and island-mainland comparisons are difficult to make (e.g., assessment of vegetative zones or communities on islands and on the mainland may not have been comparably evaluated). Nevertheless, comparisons of distributions of Greater Antillean amphibians and reptiles with those of the mainland may be instructive. Unfortunately, the only comparable information exists solely for one major taxonomic group (snakes) in only one area (Honduras) (Wilson and Meyer, 1985). When we compared the distributions of Honduran snakes relative to physiographic regions and Holdridge's life zones with those of Hispaniola (Tables 7.7 and 7.8), however, no obvious pattern emerged.

The two snake faunas differ substantially. That of Honduras consists of seven families and 95 species (two were omitted from the following comparisons, see caption to Table 7.7), only two of which are scolecophidians. In contrast, the snakes of Hispaniola represent only five families (elapids and viperids are absent) and 28 species, but 13 are scolecophidian arthropod eaters. Nevertheless, only 12 Honduran snakes (13% of the snake fauna) occupy two-thirds (five or all six) of the physiographic regions identified by Wilson and Meyer (1985) and only four (4%) are found in all regions. On Hispaniola, with its varied topography and 18 identified physiographic regions (see caption to Table 7.7), six species (21%) occupy two-thirds or more (>12) of the regions considered and one species (4%) is found in

Table 7.7. Numbers and Percentages (in parentheses) of Honduran and Hispaniolan Snake Species Relative to the Number of Physiographic Regions in Which They Occur^a.

Location	Number of Physiographic Regions								
	1	2	3	4	5	6	7	8	9
Honduras	25 (27)	24 (26)	18 (19)	14 (15)	8 (9)	4 (4)	-	-	-
Hispaniola	7 (25)	5 (18)	3 (11)	0	2 (7)	1 (4)	1 (4)	1 (4)	1 (4)
Location	10	11	12	13	14	15	16	17	18
Honduras	-	-	-	-	-	-	-	-	-
Hispaniola	0	1 (4)	0	0	1 (4)	1 (4)	3 (11)	0	1 (4)

^aThe Honduran species *Micrurus ruatanus* and *Pelamis platurus* are excluded from consideration because the distribution of the former is restricted to a satellite island and the latter is marine. The six physiographic provinces of Honduras are those identified by Wilson and Meyer (1985). The 18 Hispaniolan regions are those of Henderson and Schwartz (1984) with subdivided areas combined and all satellite islands except Ile de la Gonâve excluded.

all 18. However, at the other extreme, 49 Honduran snakes (53%) are found in one-third or fewer (2 or less) regions and 25 (27%) are found in only one, whereas 18 Hispaniolan species (64%) are limited to one-third or fewer (6 or less) regions and seven species (25%) are restricted to one region. When distribution is examined relative to vegetative life zones (Table 7.8), differences are even more equivocal. Only five Honduran snakes (5%) occur in more than two-thirds (>6) of the eight life zones identified by Wilson and Meyer (1985) and none is found in all eight. Forty-two species (45%) are found in less than one-third (<2) of the life zones, with 24 (26%) restricted to one, but another 28 species (30%) are found in three life zones. On Hispaniola, only one species (4%) lives in more than two-thirds (7 or more) of nine life zones and 18 (64%) occupy one-third (3) or fewer life zones, with seven species (25%) restricted to only one zone.

Although these data suggest that a relatively few more Hispaniolan snakes occupy a greater variety of physiographic regions than do Honduran snakes, this does not hold true when considering distributions relative to vegetative life zones, and the numbers of specialists (those species occupying few physiographic provinces or

Table 7.8. Numbers and Percentages (in parentheses) of Honduran, Cuban, and Hispaniolan Snake Species Relative to the Number of Physiographic Regions in Which They Occur^a.

Location	Number of PhvsioGraphic Regions								
	1	2	3	4	5	6	7	8	9
Honduras	24 (26)	18 (19)	28 (30)	15 (16)	3 (3)	4 (4)	1 (1)	0	-
Cuba	5 (22)	5 (22)	3 (13)	1 (4)	0	1 (4)	1 (4)	3 (13)	4 (17)
Hispaniola	7 (25)	4 (14)	7 (25)	4 (14)	3 (11)	2 (7)	1 (4)	0	0

^aThe Honduran species *Micrurus ruatanus* and *Pelamis platurus* are excluded from consideration because the distribution of the former is restricted to a satellite island and the latter is marine. The eight vegetational formations of Honduras are those identified by Wilson and Meyer (1985). The "ecogeographic" distribution of Cuban snakes is from Rodríguez (1993). The nine Hispaniolan vegetational zones are taken from maps of Haiti and the Dominican Republic produced by the Organización de Estados Americanos.

vegetative life zones) are comparable. Nor does the vastly greater number of scolecophidians on Hispaniola substantially alter the results; the 13 species include some with restricted and some with more varied distributions in about equal numbers to those of macrostomatan snakes (although no leptotyphlopoid or typhlopoid snakes are included among the most versatile species).

Using data from Rodríguez-Schettino (1993; see also Rodríguez-Schettino, 1986 and Estrada, 1993b]) on the distribution of snakes in Cuba relative to "ecogeographic" zones, somewhat different results emerge (Table 7.8). We used only data from nine zones on "mainland" Cuba, disregarding snake distributions from Isla de la Juventud and offshore archipelagos. The percentages of snakes occupying one or two zones are comparable to those of Honduras and Hispaniola, but 17% of the snake fauna (4 species) occupied all nine zones in Cuba, whereas no snakes did this on Hispaniola or in Honduras. Thirteen Cuban species (56.5%) occur in one-third of the zones, and nine species (39%) occur in two-thirds. The most widespread Cuban species fell into one of two categories: (1) species belonging to genera represented by only one species (*Epicrates*, *Alsophis*, *Antillophis*, *Tretanorhinus*), or (2) species belonging to genera represented by more than one species wherein one species was widespread and congeners were geographically (and ecologically) restricted (*Typhlops*, *Tropidophis*, *Arrhyton*). The results of our

mainland-island comparisons for the sake of determining versatility of a fauna remain equivocal, but we suggest that further investigation of this topic would be fruitful.

In addition to the possibility that a large number of ecological generalists among West Indian species might be a reflection of their ancestral status as good colonizers, the versatility of these amphibians and reptiles is an important and intriguing consideration in lieu of historical and recent responses of the herpetofauna to human influences. To fully understand the ecology of many organisms, and this may be especially true of vertebrates on islands, the impact of human history must be taken into account (Pregill, 1986; Pregill *et al.*, 1994). As on the Yucatán Peninsula (Lee, 1996), the historical introduction of predators and competitors and habitat modifications in the West Indies (Henderson *et al.*, 1996) have ramifications for species abundance and distribution and may result in extirpation or extinction (Iverson, 1978; Henderson, 1992; Hedges, 1993; Kaiser and Henderson, 1994).

Henderson (1992) noted that the herpetofauna of the Lesser Antilles is especially versatile and appears to be tolerant of habitat degradation (notable exceptions include forest-adapted *Eleutherodactylus*, *Diploglossus montiserrati*, and *Chironius vincenti*), and to date no species has been lost to habitat destruction. Similarly, Hedges (1993) concluded that it was premature to declare any frog species in the West Indies extinct, although several species have not been collected in recent years (but see Joglar and Burrowes, 1996). Hedges (1993) also stressed, however, that habitat destruction (e.g., rampant deforestation) could have catastrophic effects on the frog fauna once all forests had been destroyed (e.g., forest cover in Haiti is now at <1% of historical levels). Based on the knowledge that (1) 99% of the original forest in Haiti has been eliminated and (2) apparently no Haitian frogs have become extinct, can we conclude that elimination of the last 1% will not have a catastrophic effect? Perhaps the anuran fauna of Haiti (and possibly all of Hispaniola) will endure as long as second growth forests, banana plantations, and similar degraded habitats persist because the fauna is euryoecious.

Perhaps better examples could be drawn from studies of West Indian lizards, and biologists working in the West Indies have been often struck by the exceedingly opportunistic foraging of lizard species. In the Bahamas, Schoener *et al.* (1982) observed *Leiocephalus* (1) feeding on bread, peanut butter, jelly, crackers, and cheese; (2) lapping water from ice cubes and quickly learning to successfully solicit those ice cubes; and (3) feeding on food scraps

dropped on the floor by diners at a hotel. One of us (R. Henderson), while eating canned beans and tuna on Great Bird Island (off Antigua), had *Anolis bimaculatus* and *Ameiva griswoldi* crawl into the cans to eat. Wijffels (1997) suggested that good food availability and absence of predators (and therefore a longer life span) contributed to the large size attained by *Anolis bimaculatus* in and around human dwellings on St. Eustatius. Artificial lighting has resulted in a number of traditionally diurnal anoles occupying nocturnal niches (Schwartz and Henderson, 1991; Powell and Henderson, 1992; Bowersox *et al.*, 1994a) more commonly associated with gekkonids (usually *Hemidactylus* spp.). The possible impact on gecko populations has not been assessed, but recent (May 1994) observations of *Anolis cybotes* and *Sphaerodactylus difficilis* in nocturnal syntopy in Cabarete, Dominican Republic, suggest that, because *A. cybotes* is an opportunistic predator of *S. difficilis* (Schwartz and Henderson, 1991; Lenart *et al.*, 1996), this temporal niche expansion may provide increased opportunities for saurophagy by *A. cybotes*.

Some species appear to have benefited from the presence and activity of humans. *Eleutherodactylus johnstonei* is widely distributed throughout the Lesser Antilles and many locales outside of the region (Kaiser and Hardy, 1994). The species obviously has taken advantage of its ecological versatility and opportunities for human-mediated transport to establish populations on previously frog-free islands and even in the presence of native congeners (Kaiser and Henderson, 1994), and "as habitat degradation progresses rapidly on most Caribbean islands, the advance of *E. johnstonei* will likely proceed in step, and at the expense of local endemics."

Another example is that of heliophilic anoles, which often are very abundant in human-modified habitats. *Anolis porcatus* is rarely seen in Cuban forests, except high in the canopy and in occasional tree gaps, and *A. sagrei* is never seen in densely forested areas (J. B. Losos, in litt.). Both species, however, are ubiquitous in open, disturbed habitats. Also, consider Hispaniolan anoles. A few species are nearly everywhere (e.g., *A. chlorocyanus/coelestinus*, *A. cybotes*, *A. distichus/brevirostris*, and the grass anoles) and these forms generally thrive in altered environments [although *A. chlorocyanus* has largely been displaced by introduced Cuban *A. porcatus* in urban Santo Domingo (Powell *et al.*, 1990b) and *A. cybotes* by the Puerto Rican bank generalist, *A. cristatellus*, in the less-than-pristine environs of La Romana (Fitch *et al.*, 1989)].

Henderson and Winstel (1995) and Henderson *et al.* (1996) documented exploitation of human-modified habitats by the arboreal

boa *Corallus grenadensis* in an area of mixed agriculture and uncultivated woodlands on Grenada. Not surprisingly, habitat devoid of trees or with only solitary trees (i.e., devoid of contiguous tree crown habitat) precluded the presence of *C. grenadensis*. Small tree boas (<500 mm SVL) were encountered most often in uncultivated woods (78.1%), and medium-sized tree boas (500-1000 mm SVL) occurred in uncultivated woods (64.3%) more often than in cultivated trees (35.7%), but used the latter more often than small snakes (21.9%). Large tree boas (>1000 mm SVL) were more often encountered in mango trees (48.2%) and other cultivated trees (22.2%) than in uncultivated areas (29.6%). The cultivated trees used by the tree boas (e.g., mango, breadfruit, nutmeg, cacao) did not occur on the island until, in some cases, 150-200 years before present. Similarly, the rodent species (*Oryzomys* sp.; L. Lippold, personal communication) presumably exploited by adult *C. grenadensis* on pre-Columbian Grenada is now extinct; the rodents (*Mus*, *Rattus*) that adult *C. grenadensis* now exploit as food did not occur on the island until about 1500.

However, unlike the highly visible examples presented above, a preponderance of West Indian amphibians and reptiles rarely occur in severely altered situations. These animals apparently are, at least in most cases, descendants of effective colonizers (Hedges, 1996; but see also Crother and Guyer, 1996), but many species have evolved in situ, presumably through natural selection, into ecological specialists. To find them one must look in specific habitats (e.g., *Anolis barbouri* in montane ravines with abundant leaf litter and well shaded by intact forest canopy; Flores *et al.*, 1994). Because individual habitats are frequently restricted in area, the distributions and population sizes of these species are generally small.

Summary

The following is a hypothetical scenario that offers speculation for why things are as they are in the West Indies. The West Indian herpetofauna is largely derived from the herpetofauna of the South American mainland, and most lineages appear to have reached the West Indies by overwater dispersal. Potentially, many of the colonizers were animals of coastal habitats, forest edge, and other open situations, making them more tolerant of high ambient temperatures and sun-drenched habitats than, for example, forest-dwelling species. This tolerance would, presumably, increase their

chances of surviving a prolonged overwater journey during which shade would be minimal.

The West Indian herpetofauna is depressed in the number of families and genera present but rich in species due, in large part, to the spectacular radiations of three genera (*Eleutherodactylus*, *Sphaerodactylus*, *Anolis*). West Indian frogs, lizards, and snakes tend to occur in extremely high densities compared to mainland species. Even in rain forest habitat, anoles, for example, occur at densities hundreds of times higher than those of mainland populations, and their biomass may approach the biomass of entire lizard communities on the mainland. In short, lizard communities on the Neotropical mainland consist of more *species of lizards*, but West Indian communities are composed of *more lizards*. In turn, the West Indian snake fauna preys largely on frogs and lizards, a food source that is ubiquitous and abundant. West Indian frogs and reptiles are, in general, habitat and trophic opportunists. Exceptions, of course, exist, especially in the rich faunas of the Greater Antilles where in situ speciation has produced habitat and trophic specialists. On the smaller islands of the Lesser Antilles, ecological generalization and versatility are even more widespread.

Future Emphasis

The characteristics of the West Indian herpetofauna that, in part, have accounted for the exciting ecological research that has already been done (inter and intransland diversity, the adaptive radiations of several genera, the extremely localized distributions of some species and the broad, multi-island distributions of others, and the rarity of some species and the ubiquity and phenomenally high densities of others) remain lures for future research.

Predicting the future is a risky venture; still, the belief that ecological investigations will become more important in the twenty-first century is easy to support. Island-bound human populations continue to grow and their need for food, living space, amusement, and resources capable of being exploited in search of wealth will inevitably increase the pace at which the habitats of West Indian animals are altered. As a result, ecological studies will be necessary as a springboard not only for posing and addressing intellectual questions, but for the very pragmatic consideration of issues pertaining to the conservation of the region's flora and fauna. With this in mind, biologists must be willing to examine interactions in severely altered habitats (Powell *et al.*, 1996b), contrary to our inclination to focus on

the remaining pristine areas. Also, scientists must collaborate actively with persons in those regional and national agencies whose responsibilities include conservation, agriculture, tourism, and urban development. Although many amphibian and reptilian species are ecologically versatile, others are not. Some anurans have very specialized habitat requirements (Hedges, 1993; Kaiser and Henderson, 1994; Joglar and Burrowes, 1996) and some endemic anurans apparently are experiencing difficulty withstanding the dual onslaught of habitat destruction and the introduction of *Bufo marinus* (e.g., Kaiser and Henderson, 1994). Turtles, both freshwater and marine species, are being heavily exploited for food. Many lizard species appear to coexist successfully with humans, and a few even thrive as human commensals, but exploitation of *Cyclura* for food and of many species for the pet trade has had a measurable effect. At least the larger species of snakes are vulnerable to continued human efforts at extermination and a number have been brought to the edge of extinction by the mongoose (Henderson, 1992; Tolson and Henderson, 1993).

In light of these realities and the resultant universal need to consider applications to conservation (Greene, 1994), we have chosen not to focus on the applied but rather on the more classical types of questions that have been overlooked to date or to which past or present work is leading. In addition to issues raised specifically by conservation biology and the need to continue ongoing investigations by many biologists, we believe there are three major areas in need of attention; all are conventional, all have been neglected, and each will ultimately contribute to our ability to conserve individual species, communities, and habitats.

Community Studies

The study of herpetological communities in the West Indies has been largely confined to congeners and almost exclusively centered on *Anolis* (e.g., Rand, 1967a; Rand and Williams, 1969; Moermond, 1979; Schoener and Schoener, 1980; Roughgarden *et al.*, 1983, Losos, 1992) and to a lesser degree *Eleutherodactylus* (Stewart, 1979; Jones, 1982). On the Neotropical mainland, analyses of taxonomically diverse frog (e.g., Toft, 1980; Lieberman, 1986), lizard (e.g., Rand and Humphrey, 1968; Duellman, 1987; Vitt and de Carvalho, 1995), and snake (e.g., Vitt and Van Gilder, 1983; Cadle and Greene, 1993) faunas, or of an entire herpetofauna (e.g., Duellman, 1978, 1990; Duellman and Mendelson, 1995), are more common. Similar studies are virtually lacking for the West Indies, although the recent work of

Bullock and Evans (1990) and Thomas and Kessler (1996) addresses taxonomically diverse communities. But these, like the recent syntheses of Reagan (1996), and Reagan *et al.* (1996), have focused on communities in moist montane forests, whereas similarly diverse communities in deserts, dry forests, and chaparral have been largely neglected (Powell *et al.*, 1996b). Studies by Censky (1996), Reagan (1996), and Tolson (1996) include references to competition, predation, and habitat destruction, respectively, involving sympatric species other than amphibians and reptiles. Because similar situations exist elsewhere, future community studies should not be limited to the herpetofauna. Rather, we should seek to build on the growing body of trophic data in order to elucidate more fully the entire gamut of relationships in which West Indian frogs and reptiles play such prominent roles. Regardless of the taxonomic breadth of community studies in the West Indies, geographically they constitute a fertile arena for inter- and intrainland comparisons.

Autecologies

With the exception of those noted earlier, autecological studies are sorely lacking for the West Indian herpetofauna. As Vitt and Zani (1996) noted in an article on a Neotropical mainland lizard:

A vast majority of insights leading to major discoveries have been based on patterns revealed by analyses of large data sets taken directly from the natural history literature...but we still lack data for most species. Logistics of field research in tropical countries has contributed partially to the general lack of data for many [species in many] areas but lack of appreciation for the value of individual species studies...has undoubtedly contributed indirectly by making it difficult to acquire research support.

Even the basic natural history of frogs, with the notable exception of *Eleutherodactylus coqui*, has been largely ignored. Some species of *Bufo* and *Osteopilus* are very common, and species of *Eleutherodactylus* are frequently ubiquitous. Considering the current global interest in amphibian populations, island populations of frogs should be more attractive than ever as subjects of intensive ecological pursuits.

Despite intensive efforts, many aspects of sea turtle ecology remain a mystery, and the few remaining viable populations of freshwater turtles and crocodylians deserve attention. Along with anurans in general, detailed investigations of nonanoline lizard ecology are virtually absent from the literature. Interest in *Cyclura* has remained fairly constant, but they are large and spectacular in appearance. The diminutive gekkonids in the genus *Sphaerodactylus*

offer opportunities at the other size extreme. They often occur in high densities (Table 7.2), little is known about social interactions, and nothing is known about movement ecology or habitat utilization. Considering the diversity and distribution of *Sphaerodactylus*, our lack of knowledge is surprising. Likewise, the large gekkonids of the genus *Aristelliger*, a magnificent *Ameiva* fauna, and a host of intriguing anguids in an amazing array of sizes beg for attention.

Considering the disproportionate amount of attention anoles have received in the past 30 years, it is surprising that an autecological approach to studying anoline ecology has been ignored. One spectacular segment of the anoline fauna has been neglected completely: the giant species, and these lizards (e.g., *A. barahonae*, *A. smallwoodi*) are large enough to monitor with radio telemetry (Losos *et al.*, 1990). With the exception of the boines, the snake fauna has, been largely ignored ecologically. The dwarf boas (*Tropidophis*) and many colubrids (e.g., *Alsophis*, *Antillophis*, *Uromacer*) are common enough for mark and recapture methods or large enough for the use of radio telemetry.

Amphibians and Reptiles in Modified Habitats

Few, if any, ecologically oriented studies of amphibians and reptiles in the West Indies have been conducted in habitats that have not been affected, to one degree or another, by humans. The long history of humans in the West Indies provides a fertile ground for examining their role in geographic distribution and ecology. Lee (1996), for example, has done extensive analyses of the role of the Mayan culture in influencing the present-day herpetofauna (distribution and ecology) of the Yucatán Peninsula of México. In the West Indies, Kaiser (1992) has masterfully documented how trade-mediated human introductions may have been the single most important factor in creating the current distribution of *Eleutherodactylus* in the Lesser Antilles. Henderson *et al.* (1996) described and documented post-Columbian changes in the ecology of the arboreal boid *Corallus grenadensis* in Grenada; this species has had to adapt to new habitats, new prey species, and new predators during the past 500 years. Similarly, Henderson *et al.* (1987a) provided a scenario which described a shift in prey species by the boid *Epicrates striatus* on Hispaniola from now-extinct rodents and insectivores to European-introduced *Mus* and *Rattus*; the shift probably became complete in the early twentieth century.

Opportunities abound for investigations of human-disturbed habitats (e.g., Meshaka, 1996), because few, if any, species of frogs

and reptiles in the West Indies have not been forced to survive in human-modified habitats. For example, at one site in Barahona, Dominican Republic, at least 13 lizard species coexist in a severely altered area of 1400 m^2 (Fobes *et al.*, 1992; Schell *et al.*, 1993b; Schreiber *et al.*, 1993). Similarly, Olson (1995) observed amphibians and reptiles to be diverse and common in Haitian courtyards. Studies of anoles using fence posts and coconut palms, geckos and frogs exploiting the prey-attracting properties of electric lights and the cover of human dwellings, or *Alsophis* using sheets of discarded plywood for cover are all common examples of amphibians and reptiles exploiting human modifications of habitat. Some are subtle, others obvious, but all are examples of portions of the herpetofauna adapting to human disturbances. The West Indian herpetofauna, among other insular and mainland faunas, is in a state of flux. The introduction of predators (*Herpestes javanicus*: Baskin and Williams, 1966; Henderson, 1992; cats and dogs: Iverson, 1978; *Rattus rattus*: Tolson, 1988), prey species (*R. rattus*, *Mus musculus*: Henderson *et al.*, 1996), and competitors (e.g., *Eleutherodactylus johnstonei*: Kaiser and Henderson, 1994; various *Anolis*: Fitch *et al.*, 1989; Losos *et al.*, 1993) contribute to modifications in the present-day distributions of many species (Case and Bolger, 1991; Case *et al.*, 1992; Losos *et al.*, 1993). Determining how the herpetofauna is responding to these modifications, be they subtle or flagrant, should be a top priority.

Evolutionary Relationships

Brian I. Crother

*Department of Biology
Southeastern Louisiana University
Hammond, Louisiana 70402*

Nothing in evolution makes sense except when seen in the light of phylogeny.

—Jay M. Savage, 1997 Society of Systematic Biologists Presidential Address

Introduction

The West Indies hosts at least 585 species of reptiles and amphibians in some 70 genera representing Anura (157 species, 11 genera), Testudinata (8 species and 4 genera; excluding sea turtles), Sauria (317 species and 29 genera), Serpentes (99 species and 24 genera), and Crocodylia (4 species and 2 genera) (Schwartz and Henderson, 1991). Given these large numbers of taxa for a quite definable biogeographic unit (see Chapter 10), the task of estimating the phylogenetic relationships within these groups is and has been no simple task. However, the continuation of the task has become more important than ever with the current and looming extinction crises and the consequent necessity to understand global biodiversity. To identify crisis areas for conservation, or just simply the “best” (= most diverse?) areas for preservation, methods have been developed that employ phylogenies as the baseline data (e.g., Crozier, 1992, 1997; Faith 1992, 1994; Forey *et al.*, 1994; Vane-Wright *et al.*, 1991). Thus, at what one might characterize as one of the cutting edges of applied biology is phylogeny. Although the application of phylogeny to conservation may seem surprising, it is not surprising how central phylogeny has become in comparative studies of arguably all biological disciplines. Its importance can perhaps be measured by the number of texts that have been recently published on the comparative

method in biology: e.g., Brooks and McLennan (1991), Eggleton and Vane-Wright (1994), Harvey and Pagel (1991), and Stearns (1992). In addition to conservation and comparative biology in the broad sense (whether it be organismal or molecular), phylogeny has also become the cornerstone for the study of historical biogeography, regardless of the method (Crother and Guyer, 1996). Finally, phylogenies, especially when constructed from different data sets, can be used to assess the accuracy of phylogenetic methods themselves.

Based on the necessity of phylogeny in biology, both applied and basic, it has become important to address phylogenetic questions of all groups. This, with the aid of modern methods, has led to the production of phylogenies at rates never seen before, which means phylogenies are being reconstructed that are in some cases less than optimal and thus have led to and will lead to controversy and debate over their veracity (accuracy). This chapter will attempt to review all the phylogenetic (not taxonomic) work that has been conducted with Caribbean herpetofaunal groups and in the process discuss some of the debates that have already arisen in our attempts to produce best estimates of phylogeny. [Hedges (1996) made general comments on the relationships of most West Indian taxa, but because those comments were not accompanied by data I have eschewed including his notions in the following review.] In this review, I will also assess the phylogenies for general robustness, which admittedly will be biased in favor of conscientious cladistic analyses and hypotheses based on multiple data sets. I will also point out taxonomic areas in need of work and possible directions of future work.

Anura

The great majority of the West Indian anurans are endemics, and it is these groups that will be focused on in the succeeding discussion. Thus for example, for taxa such as *Rana grylio*, *Hyla crucifer*, and *Gastrophryne carolinensis* there will not be a discussion of phylogeny.

Bufonidae

There are two genera of bufonids found in the West Indies, one which is endemic. The genus *Bufo* is represented by the widespread and introduced *B. marinus* and as such will not be given further attention. The endemic genus, as concluded by Pregill (1981a) is *Peltophryne* and it is composed of 10 species (following Schwartz and Henderson, 1991).

The relationships within the genus are not well understood and no cladograms have been presented. Ruibal (1959) and Schwartz (1972) seemed to suggest that *P. peltacephala*, *gundlachi*, and *cataulaciceps* formed one group and that *empusa*, *fluviatica*, *guentheri*, *lemur*, and *taladai* formed a second group with *longinasa* apparently not related to either group. Tihen (1962) considered the Caribbean taxa possibly related to a group consisting of North and South American species, but this was broken up by Martin (1972). Cei *et al.* (1972) hypothesized that *peltacephala* shared a relationship with the *Bufo granulosa* group of South America. Finally, Pregill (1981a) discussed the cranial morphology and its evolutionary implications for the West Indian taxa. Unfortunately, but correctly so, Pregill (1981a) did not construct a phylogeny from his data because of outgroup difficulties. However, he did make some general phylogenetic statements about possible relationships. Pregill (1981a) suggested that *fluviatica* and *longinasa* were the most primitive taxa; *lemur* and *guentheri* were probably sister taxa; *taldai*, *gundlachi*, *cataulaciceps*, and *empusa* may form a monophyletic group; and *peltacephala* was unique and not yet placeable.

The intergeneric phylogeny, as the intrageneric phylogeny, has not been estimated in any detail. In a phylogenetic analysis focusing on the position of *Frostius*, Cannatella (1986) noted that *Peltophryne* (as well as *Crepidophryne* and *Rhamphophryne*) was easily separated from all other neotropical bufonids but did not include it in the analyses and resultant cladograms. However, Graybeal and Cannatella (1995) later noted that the monophyly of *Peltophryne* was questionable.

In summary, the genus *Peltophryne* remains unaddressed phylogenetically at both the interspecific and intergeneric levels; i.e., baseline phylogenetic hypotheses have yet to be constructed for this group. Also, other than Pregill's (1981a) work, the previous comments on interspecific relationships have been "only conjectural treatment" (Pregill, 1981a:273).

Hylidae

The level of phylogenetic knowledge for the hylid radiations in the West Indies is similar to that of the bufonids; no cladistic hypotheses have been put forth even though the taxa have received attention, albeit none as a group since Trueb and Tyler (1974). Dunn (1926a) considered the Jamaican hylids, *Calyptahyla crucialis*, *Hyla marianae*, *H. wilderi*, and *Osteopilus brunneus* to form a monophyletic unit derived from His-

paniola. Dunn (1926a) also considered the members of the Hispaniolan group, *Hyla heilprini*, *H. pulchrrilineata*, *H. vasta*, and *Osteopilus dominicensis* to be each others' closest relatives. And finally, Dunn (1926a) considered the entire Antillean radiation (thus including the Cuban *septentrionalis*) to be derived from a single invasion event, therefore implying their monophyly. Noble (1927) and Shreve and Williams (1963) discussed other possibilities but essentially concluded to follow Dunn (1926a). Duellman (1970), in his monograph on Middle American hylid frogs, disputed both of Dunn's (1926a) island groups. He cautiously accepted Trueb's (1970) first discussion on the casque-headed frogs, in which she suggested a *septentrionalis* group composed of *septentrionalis*, *dominicensis*, *vasta*, *brunneus*, and *lichenata*. Acceptance of this group breaks up both of Dunn's (1926a) groups. Duellman (1970) went further with the Hispaniolan hylids and indicated that *heilprini* and *pulchrrilineata* were unique lineages and that neither were related to the *septentrionalis* group of Trueb (1970) or the other *Hyla* of Jamaica. Schwartz and Fowler (1973) reviewed the ideas of Dunn (1926a), Shreve and Williams (1963) and Trueb (1970) and decided that they had little argument against Trueb's *septentrionalis* group. Schwartz and Fowler (1973) also noted, in similar fashion to Duellman (1970), that the relationships among the small hylids of Jamaica and Hispaniola were unclear and entertained the idea put forth by Dunn (1926a) that they were neotenic forms of the larger hylids in the *septentrionalis* group.

The most recent work on the entire Antillean radiation of hylids is that of Trueb and Tyler (1974). They noted that there were three recent introductions (*Oloolygon rubra* from South America to St. Lucia, *Hyla squirella* from North America to Grand Bahama, and *H. cinerea* from North America to Puerto Rico) and that their affinities are not with any of the nine endemic taxa. Trueb and Tyler (1974) rejected the monophyletic and neotenic hypotheses of Dunn (1926a), and based on morphological differences suggested that the West Indian hylids were composed of six distinct lineages, each related to mainland components and not to each other. These lineages are *marianae-wilderi*, *heilprini*, *pulchrrilineata*, *septentrionalis-dominicensis-brunneus*, *vasta*, and *lichenata*. The hypothesized mainland relationships of *heilprini* possibly were ("highly reminiscent of", Trueb and Tyler, 1974:32) with the *albomarginata* group, the *septentrionalis* group with *Osteocephalus*, *vasta* with the *boans* group, and *lichenata* perhaps with *Osteocephalus* or *Trachycephalus*. The other taxa could not be placed.

The possible disparate relationships among the West Indian hylids is suggestive of a complex history for the group and would thus make estimation of their phylogenetic relationships difficult. However, based on the information of Trueb and Tyler (1974), any future cladistic study on the West Indian hylids must not assume monophyly of the group and must include numerous mainland taxa in order to clarify the extent of the nonmonophyletic nature of these hylids.

Leptodactylidae

The genus *Leptodactylus* is represented in the West Indies, according to Schwartz and Henderson (1991), by five species, three of which are endemic (*albilabris*, *dominicensis*, *fallax*) and two that are South American (*insularum*, *wagneri*). However, the recognition of these taxa was disputed by Heyer (1978, 1979). Heyer (1978) presented evidence which indicated that *albilabris* and *dominicensis* were conspecifics and consequently he synonymized *dominicensis*. Heyer (1970) figured a phylogeny of the *melanotus* group that included *wagneri*, but later (1994) he considered West Indian *wagneri* to be possibly confounded with other populations and still considers its status to be unresolved. Curiously, Schwartz and Henderson did not mention Heyer's work, nor did they discuss reasons for the acceptance of *dominicensis*. As a general statement about *Leptodactylus* relationships, they apparently do not form a monophyletic group, with each taxon possibly being associated with a different mainland lineage. *Leptodactylus fallax* is a member of the *pentadactylus* species group and is hypothesized to belong to a clade of unspecified relationships which include *flavopictus*, *knudseni*, *labyrinthicus*, and *pentadactylus*. *Leptodactylus albilabris* (*sensu* Heyer) is a member of the *fuscus* species group and is hypothesized to belong to a clade of unspecified relationships which include *bufonius*, *fuscus*, *gracilis*, *labialis*, *latinasus*, *mystaceus*, and *mystacinus*. Bogart (1974) presented karyotype data for *dominicensis* (= *albilabris*) that did not dispute its membership in the *fuscus* group. Explicit phylogenetic hypotheses are yet to be constructed for either of these presumptive clades.

Unlike *Leptodactylus*, the genus *Eleutherodactylus* (over 130 species) has had several explicit phylogenetic hypotheses put forth based on morphological (Joglar, 1989; Kaiser *et al.*, 1994b,c; Lynch, 1996), allozyme (Hedges, 1989a,b, 1991; Kaiser *et al.*, 1994d, 1996; Smith *et al.*, 1981), immunological data (Hass and Hedges, 1991), and multiple data sets (Kaiser, 1996). Hedges (1989a) and Joglar (1989) attempted to ex-

amine all the species, but only Hedges presented hypotheses for most of the taxa (Fig. 8.1). Joglar (1989) presented summarized phylogenies. Perhaps the best way to assess the state of West Indian *Eleutherodactylus* phylogeny would be to search for congruence among the various data sets. Starting at the most inclusive level, the phylogenies of Hedges (1989a) and Joglar (1989) are difficult to reconcile because of the summarized nature of Joglar's phylogeny, which was based on a cladistic analysis of morphological characters. However, Joglar suggested that the *auriculatus* group, *unistrigatus* group, and the *recordii* group [these groups are somewhat similar in content to Hedges's (1989a) groups] form parts of a polytomy which also included *Syrrhopus* and *Tomodactylus*. Sister to this polytomy was the *inoptatus* group.

At best, Joglar's hypothesis is consistent with one of Hedges distance-based (Cavalli-Sforza distance) trees. Hedges presented two trees based on allozymes with all of the taxa he studied (84 species), one constructed with UPGMA (unweighted pair-group method of averages) and the other with distance Wagner. As mentioned previously, the UPGMA tree was consistent with Joglar's hypothesis in that the *inoptatus* group was outside the other groups. However, the two hypotheses of Hedges (1989a) were not congruent with each other and in fact were quite different in the relationships of the groups. The UPGMA tree suggested (((I,II)III)IV), whereas the distance Wagner tree showed ((I,IV)III)(II,III). Of these four major groups of Hedges, none were monophyletic, but in the distance Wagner tree his group I (composed of Jamaican taxa) did fall out in a single clade.

Hedges (1989a,b) presented several different hypotheses on the relationships of the Jamaican group. In a paper exclusively on Jamaican *Eleutherodactylus*, Hedges (1989b) constructed two distance-based phylogenies (UPGMA and distance Wagner with Cavalli-Sforza distance) and a character-based phylogeny (FREQPARS). None of these hypotheses are congruent with each other nor with the trees in the larger work (1989a).

Hass and Hedges (1991) constructed a phylogeny of five *Eleutherodactylus* taxa (of different subgenera) from immunological distance data clustered with distance Wagner. Comparison of these relationships (*gossei*, *nubicola*)*planirostris*(*montanus*, *inoptatus*) to the allozyme relationships, (((*gossei*, *nubicola*)*inoptatus*)*montanus*)*planirostris*) and (((*gossei*, *planirostris*)(*nubicola*,*montanus*))*inoptatus*), reveals the absence of congruence between the immunological data and the allozyme data.

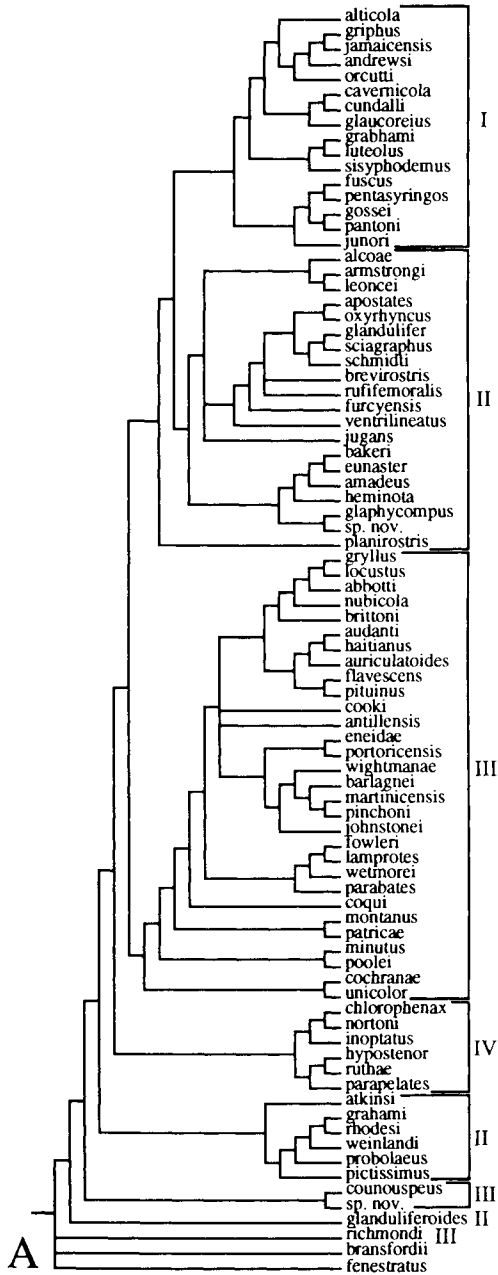
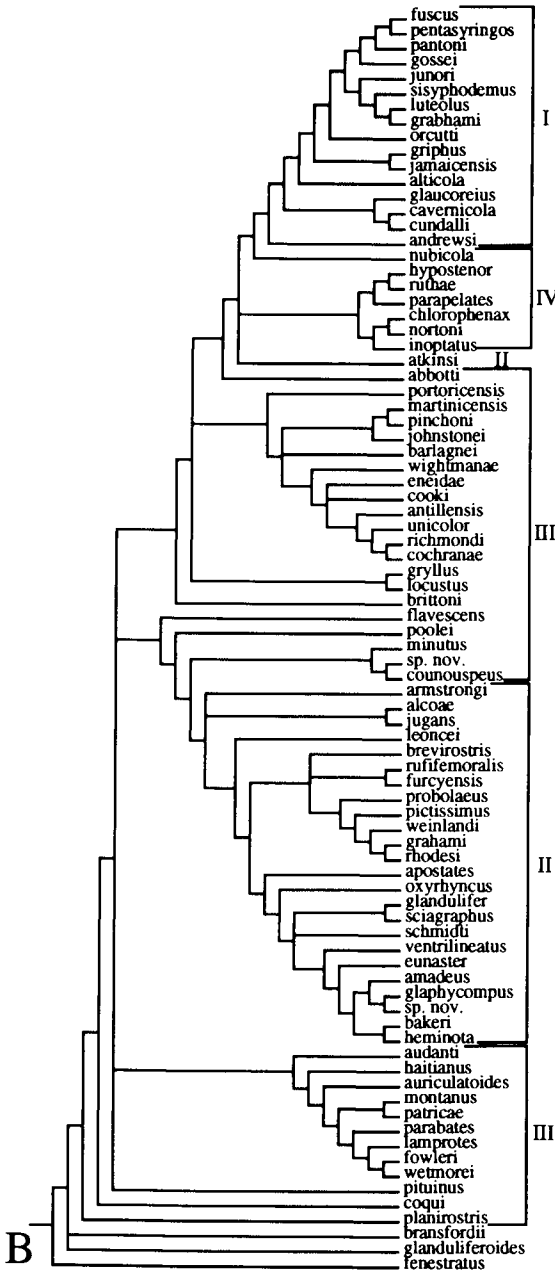


Figure 8.1. Phylogeny estimates of West Indian *Eleutherodactylus* based on allozyme data. (A) UPGMA tree. (B) Distance Wagner tree. Branch lengths do not correspond to distance estimates (re-drawn from Hedges, 1989).



Smith *et al.* (1981) published an allozyme-based distance phylogeny of part of the Puerto Rican *Eleutherodactylus* species assemblage. In comparison with the results of Hedges (1989a) there is great disparity. Hedges, in his complete trees, depicts the Puerto Rican bank species as nonmonophyletic, which is an assumption made by Smith *et al.* However, in one of Hedges' (1989a:356) character-based trees, the Puerto Rican taxa are included in the same clade (unresolved relationships) but joined with the Lesser Antillean taxa.

Joglar (1983) also published an allozyme study of the Puerto Rican *Eleutherodactylus*, but his study had a more complete sampling than Smith *et al.* (1981). However, Joglar's study shared other limitations with the Smith *et al.* study: distance-based phenograms and assumed monophyly. Joglar presented two hypotheses from two different distances, both clustered with UPGMA. The two phenograms shared some similarities in sister taxa and basal lineages but otherwise the two trees differed in structure. In some aspects the Joglar and Smith *et al.* trees were consistent, but the consistency differed between the two Joglar trees. As far as comparison with the Hedges (1989a) hypothesis is concerned, the remarks are essentially the same as for the comparison between the Smith *et al.* (1981) and the Hedges tree: congruence is lacking. The species pairs that are resolved in Hedges [(*portoricensis*, *enieidae*) and (*gryllus*, *locustus*)] were not found by Joglar. Again, as in the Smith *et al.* study, perhaps the assumption of monophyly confounded the results, or perhaps as noted by Page and Lydeard (1994), the data derived from sequential electrophoresis (the method employed by Hedges, 1989a) is intractable for phylogenetic analysis because the relationships between the numerous character states are uncertain.

To complicate matters, Lynch (1996) reexamined the phylogeny of the subgenus *Pelorius* which was erected by Hedges (1989a) and assigned six Hispaniolan species. Hedges found two three-taxon sister clades, one resolved and the other not. Lynch (1996), employing out-group comparisons, could not find resolution in either clade and concluded that support for the two clades was at best weak or simply absent.

Kaiser (1996) summarized multiple (five) data sets into a consensus hypothesis for the Lesser Antillean (LA) *Eleutherodactylus*. He concluded that the LA assemblage was not monophyletic, with the northern clade probably related to Greater Antillean forms and the southern clade probably related to South American taxa. Again, the search for congruence between Kaiser's conclusions and Hedges' work failed. Hedges

(1989a) presented four different hypotheses for his LA representatives. The two distance-based hypotheses (UPGMA and distance Wagner) were incongruent (Fig. 8.1) with each other and with Kaiser (1996). Hedges (1989a) also presented two character-based hypotheses, one which placed the LA taxa in an unresolved clade with the Puerto Rican taxa and the other which separated the LA taxa but left them unresolved.

As noted by Hedges (1989a), the groups he recovered are largely similar to the groups recognized by Dunn (1926a) and Schwartz (e.g., 1965c, 1969c, 1976b.). However, because of the persistent incongruities among the modern hypotheses (as noted previously) questions arise as to why resolution has remained elusive. The obvious answers are inconsistent approaches to phylogeny reconstruction coupled with data perhaps inadequate for resolving the problem. Also, compounding the problem is simply the size of the group. This is an enormous group of over 130 taxa and thus represents a Ruthian task. Possibly the best way out of the current morass of West Indian *Eleutherodactylus* phylogeny would be through the analysis of additional data sets and a subsequent total-evidence approach. Then perhaps a stronger phylogenetic signal will emerge to provide a robust hypothesis of West Indian *Eleutherodactylus* evolution.

Sauria

Amphisbaenidae

This morphologically peculiar and phylogenetically enigmatic squamate group (e.g., Gans, 1967, 1978; Rage, 1982; Gauthier *et al.*, 1988; Presch, 1988; Schwenk, 1988) is represented in the West Indies by two genera, *Amphisbaenea* and *Cadea*, and 14 species. Equally as intriguing as the morphology and evolution is their biogeographic distribution in the West Indies (the global distribution is also an eye-catcher for the biogeographer): the Greater Antilles sans Jamaica. Although the group is seemingly tantalizing for systematists and certainly has not been ignored (e.g., Vanzolini, 1951; Zug and Schwartz, 1958; Gans and Alexander, 1962; Thomas, 1965a, 1966d), no explicit phylogenies have been produced for the West Indian radiation.

Vanzolini (1951) presented a systematic rearrangement but did not discuss relationships. Zug and Schwartz (1958) examined only *Cadea*, and Thomas (1965a, 1966d) mostly described new diversity. Only Gans and Alexander (1962), in their monograph on West Indian *Amphisbaenea*

and *Cadea*, even remotely considered relationships. At the outset they expressed caution at the evaluation of phylogeny and decided that any explicit statements would be made at a later date. Gans and Alexander did note that the taxa showed high similarity in external characters and that the inclusion of internal characters did little to further sort out phylogeny. A single suggestion of relationships was made based on body annuli. Three groups were identified: *innocens* - *cubana cubana*, *cubana barbouri* - *manni* - *caeca*, and *bakeri* - *fenestrata*. However, they stressed that no overall pattern emerged other than that *manni* appeared distinct from the others.

The apparent inability of morphology to elucidate relationships even within the West Indies assemblage might suggest an ancient and homoplastic group. The ancient aspect is almost certainly the case, given that its current global distribution may have been affected by the breakup of Pangea (Gans, 1990). Perhaps sequences from conservative DNA regions are required to address the problem, and apparently at this time molecular data are being collected (C. Gans, personal communication).

Anguidae

This family is represented in the West Indies by five genera and 23 species. Four of the genera and 22 of the species belong to the subfamily Diploglossinae and all these species are endemic. The fifth genus (and 23rd species) is *Ophisaurus ventralis*, which was introduced to Grand Cayman Island, where it is possibly now established (Schwartz and Henderson, 1991). The other genera in the West Indies are *Celestus*, *Diploglossus*, *Sauresia*, and *Wetmorena*, with the latter two genera endemic to Hispaniola. The definition, and thus the contents of these genera remain in flux, and perhaps the main reason is because of the absence of any explicit and robust phylogenetic hypotheses.

Ideas of relationship and content have been suggested and discussed and have largely been centered around scuttelation, sheathed or exposed claws, and osteoderm patterns (Boulenger, 1885; Dunn, 1939; Grant, 1940b; Savage and Lips, 1993; Schwartz, 1970; Strahm and Schwartz, 1977; Underwood, 1959a). The key character for delimiting the taxa was sheathed or exposed claws until Strahm and Schwartz (1977) recognized that the claws were not retractile. The character Strahm and Schwartz (1977) relied on was osteoderm pattern, and based on this character they reorganized *Celestus* and *Diploglossus*. Savage and Lips (1993) reemphasized the importance of the sheathed or exposed claw character (and

deemphasized the osteoderm character following Wilson *et al.*, 1986) and once again reorganized the genera along traditional lines. Savage and Lips (1993) attempted a phylogenetic analysis using eight external morphological traits but achieved only an unresolved polychotomy. Thus, phylogenetic support for their classification remained elusive. However, Savage and Lips (1993) presented a biogeographic argument to support their classification over that of Strahm and Schwartz (1977). Regardless, the phylogeny of the Caribbean anguid fauna remains unresolved.

Gekkonidae

The gekkonid assemblage is rather interesting in that although there are seven genera (*Aristelliger*, *Gonatodes*, *Hemidactylus*, *Phyllodactylus*, *Sphaerodactylus*, *Tarentola*, *Thecadactylus*) found in the Caribbean region, none are endemic. *Sphaerodactylus* is by far the largest radiation, but *Aristelliger* is the best candidate for a true West Indian genus, with five of the six species found exclusively in the West Indies and the other taxon located on western Caribbean islands and Belize. Overall, this genus has received little attention and so the interspecific phylogeny remains purely speculative. Schwartz and Crombie (1975) proposed that *Aristelliger* was composed of two distinct groups, one containing *cochranae* and *barbouri* [this followed Hecht (1951), who actually placed these taxa in the subgenus *Aristelligella*] and the other composed of the other four taxa (*georgeensis*, *hechti*, *lar*, *praesignis*). Within this latter group, Schwartz and Crombie (1975) went only so far as to say that *hechti* is probably closer to *lar* and *praesignis* than to *georgeensis*. Interestingly, they cautioned that additional species may be hidden under these four names and further speculation on phylogeny seemed premature. It is surprising that recent attempts have not been made to estimate the phylogeny of this genus; it seems perfect for an allozyme data set.

The phylogenetic position of the genus *Aristelliger* was figured by Russell (1979) based on aspects of foot structure. Russell placed *Aristelliger* as the sister to a clade composed of *Gekko* and *Chondrodactylus*, two Old World taxa.

As currently understood, the genus *Gonatodes* is represented in the Caribbean islands of Cuba, Jamaica, Hispaniola, and Grand Cayman by a single species, *albogularis* (but note that subspecific differentiation is recognized; Vanzolini and Williams, 1962). To my knowledge, no explicit interspecific phylogenetic hypotheses have been published for the genus. The position of the genus has been speculated upon many times

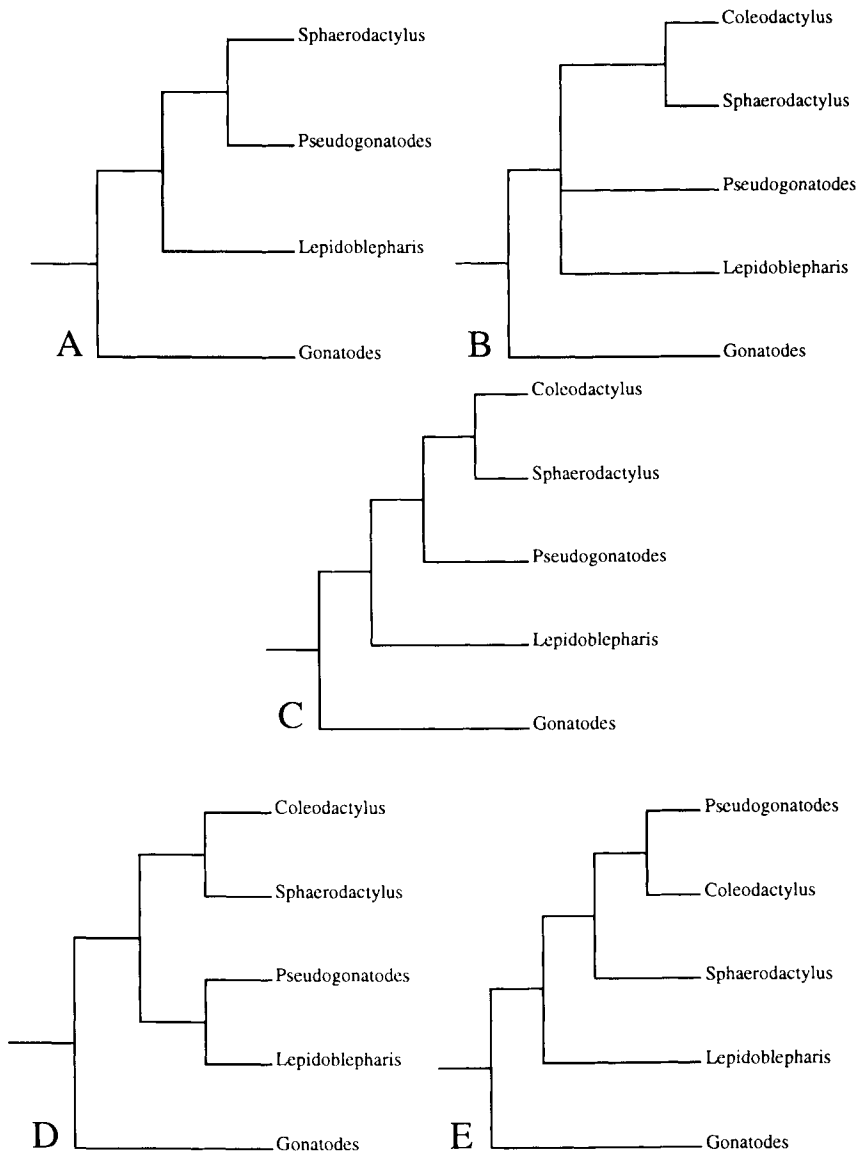


Figure 8.2. Various phylogeny estimates of gecko genera *Gonatodes* and *Sphaerodactylus*. (A) Noble, 1921, (B) Park, 1926, (C) Vanzolini, 1968, (D) Russell, 1972, (E) Kluge, 1995 (redrawn from Kluge, 1995).

(Noble, 1921; Parker, 1926b; Vanzolini, 1968; Russell, 1972); in addition, a rigorous cladistic analysis has also been conducted (Kluge, 1995). Remarkably, these five hypotheses are congruent on the placement of *Gonatodes*: basal to the other sphaerodactyl genera (Fig. 8.2).

The apparently amazing dispersal capabilities of gekkonid lizards are exemplified in the genus *Hemidactylus*. Five species are recognized in the Caribbean region: two are clearly human introductions (*garnotii*, *turcicus*) and so will not be considered further, two others are most probably natural trans-Atlantic migrants (*brookii*, *mabouia*), and the fifth is a Lesser Antillean-northeastern South American endemic. Kluge (1969) argued strongly within a phylogenetic context that the widespread *brookii* and *mabouia* were conspecifics with their African populations. He also convincingly argued that they must have arrived to the New World via trans-Atlantic rafting and not by human introduction. Kluge (1969), (when computer-assisted phylogeny estimation was still primordial) constructed a Wagner diagram and found *mabouia* and *brookii* as sister taxa but also depicted *brookii* as paraphyletic with regard to the New World endemic *palachthus*. Based on Kluge's hypothesis, I suspect biochemical data may be useful here in teasing out patterns of differentiation. At the generic level, Russell (1979) suggested that *Hemidactylus* was the sister of part of *Cyrtodactylus*.

Another broad-ranging taxon with representatives in the Caribbean is *Phyllodactylus*. Two species are recognized, *wirshingi* on Hispaniola and Puerto Rico and *pulcher* on Barbados. Schwartz (1979f) argued that *wirshingi* was a natural resident and had undergone subsequent diversification, at least at the subspecific level. The relationships of these taxa have not been explicitly examined, although Dixon (1962) speculated that *wirshingi* was closely related to *martini* of the Dutch Leeward Islands. Russell (1979) included *Phyllodactylus* and placed it as the sister to the clade containing *Aristelliger*.

The latest list (Powell *et al.*, 1996a) of Caribbean *Sphaerodactylus* totaled 78 species (another dozen or so are found in Central and South America), which represents the third largest herpetofaunal radiation in the region [behind the obvious *Eleutherodactylus* and *Anolis* (as conceptualized by Williams, 1976, and not Guyer and Savage, 1986; but see the following discussion of polychrotid lizards)]. Given the size of the radiation it is no surprise that many ideas have been put forth on the relationships of various parts of the radiation and that really only one

worker (Hass, 1991, 1996) has attempted to resolve the phylogeny of the entire West Indian radiation.

Three different hypotheses have been presented about the relationships of the genus: (*Pseudogonatodes*, *Sphaerodactylus*) (Noble, 1921); (*Coleodactylus*, *Sphaerodactylus*) (Parker, 1926b; Vanzolini, 1968; Russell, 1972); and ((*Coleodactylus*, *Pseudogonatodes*) *Sphaerodactylus*) (Kluge, 1995). Kluge's hypothesis represents the only detailed cladistic analysis and so at this time should be considered the best estimate of *Sphaerodactylus* relationships at the generic level.

The intrageneric relationships of *Sphaerodactylus* are a bit more complicated. As with other groups, various speculative comments have been made about the relationships of parts of the genus. For example, King (1962:48) in the following quote suggested the Lesser Antillean forms were not monophyletic: "The species in the Lesser Antilles are closely related to and possibly conspecific with species widespread in the Greater Antilles (*S. macrolepis* with *S. difficilis*, *S. grandisquamis*, and *S. notatus*) and on the mainland (*S. molei* with *S. lineolatus*)." Schwartz and Thomas (1983) addressed King's notions in a study of the *difficilis* complex. Based on six morphological characters, they figured a dendrogram (as noted in Chapter 1, the only tree to be found in a Schwartz taxonomy paper) of the complex and found little to support King's relationships but did support King's ideas of nonmonophyletic island radiations. Also, Schwartz and Garrido (1985) indicated that the Cuban assemblage of *Sphaerodactylus* was derived from multiple areas, and thus was also not monophyletic. Such narratives need not be further examined because in the case of *Sphaerodactylus*, modern attempts at estimating the phylogeny have been put forth (Hass, 1991, 1996).

Hass (1991) employed allozyme data derived from sequential electrophoresis (Singh *et al.*, 1976) to "determine the major groups of West Indian *Sphaerodactylus* and not the relationships within these groups." Hass included 48 species in this study and presented a character-based bootstrap proportion consensus tree and a distance Wagner tree (she also presented a cladogram showing the character distributions but because this tree was only 1 of 1000, the relationships are not discussed). A strict consensus tree of the parsimony analysis was not shown (presumably because of lack of resolution: Hass mentioned that only four species pairs were resolved). The character-based bootstrap tree was mostly unresolved and did not have a high proportion for the support of West Indian monophyly. The distance tree was fully resolved (as it must be) but out

of the 45 nodes, only two had bootstrap proportions less than 90%, 13 had proportions less than 50%, and four had proportions of zero. Therefore, the robustness of this tree, although fully resolved, must be questioned.

Page and Lydeard (1994) questioned the phylogeny and in a parsimony reanalysis they essentially obtained the same largely unresolved tree that Hass (1991) presented. Page and Lydeard suggested the problem lay in the data derived from sequential electrophoresis. Hass reported 152 alleles from 15 loci and Page and Lydeard noted that the relationship between the character states could not be known and that as the number of states increase, the informativeness of those characters decreases

Hass (1996) added a mitochondrial DNA (mtDNA) data set (16S gene, 330 bases) to the fray with a reduced number of taxa (28). Phylogenetic hypotheses were derived from neighbor-joining (distance) and maximum parsimony (character) analyses. Twenty-one most parsimonious trees were found and although there were no measures to interpret the robustness of the parsimony tree or the fit of the data, the majority-rule consensus tree had only four nodes not found in 100% of the 21 trees. The neighbor-joining and parsimony trees were largely consistent, but as might be expected they were not fully congruent (Fig. 8.3). Most of the incongruence was in the internal nodes, although some of the smaller terminal groups also were rearranged. Even so, Hass (1996) seems to have at least corroborated some of the traditional groupings within *Sphaerodactylus*, such as the *argus* series, *ramsdeni* group, *nigropunctatus* group, and part of the *difficilis* complex. She also found (with three exemplar species), contra to previous assertions, a monophyletic LA clade.

Resolution in terms of relationships of these groups must await further analyses and overcome a possibly detrimental sampling problem. An immediate possibility in terms of analysis would be to combine the allozyme data with the mtDNA for the 20 overlapping taxa. (On the other hand, this may be inappropriate given the independent history of the mtDNA. Because of this, the mtDNA is probably more appropriately employed as a consilient data set.) The covariation in the mtDNA data may help organize the multitude of alleles in the allozyme data set. The sampling problem refers back to work done by Harris and Kluge (1984) on Central American *Sphaerodactylus*. Their cladistic study could not conclude that the mainland species were monophyletic. If this is correct, then the only possible relatives would be in the West Indies. Thus, the

implicit assumption employed in Hass' studies that the West Indian *Sphaerodactylus* are monophyletic may be incorrect. This may be the reason why the relationships among the groups appear unstable.

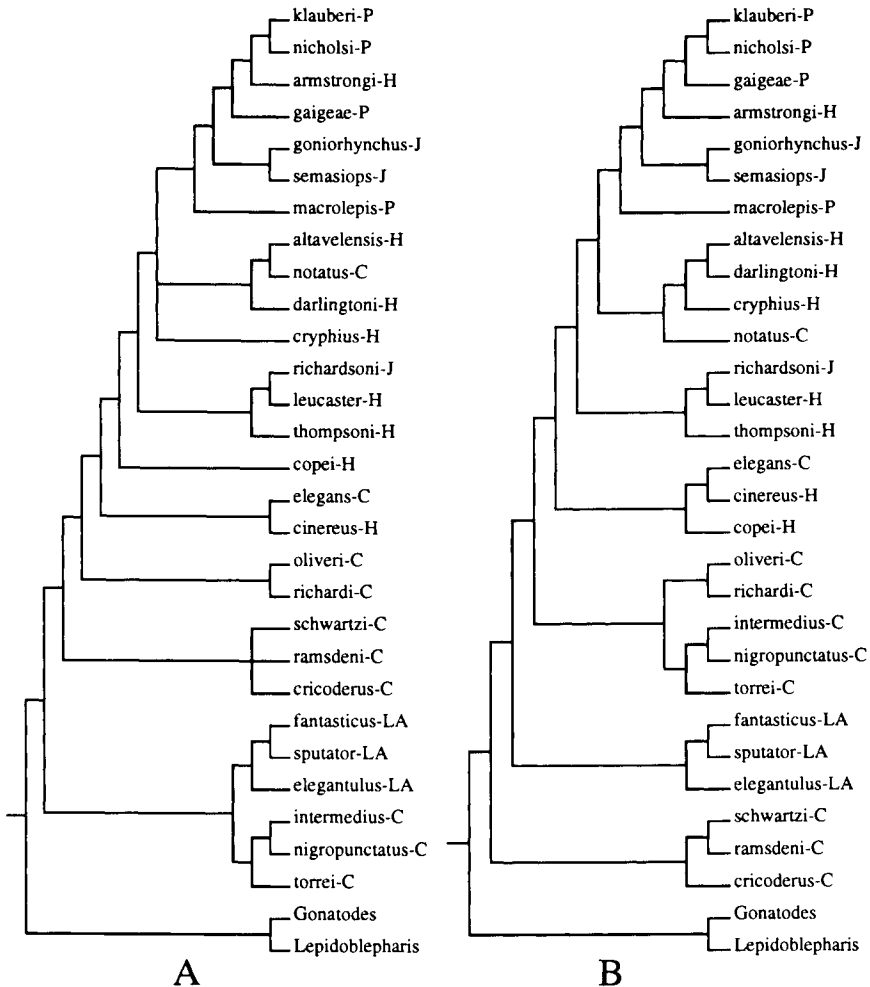


Figure 8.3. Phylogeny estimate of *Sphaerodactylus* based on mitochondrial 16S DNA sequence. (A) Majority-rule consensus of 21 most parsimonious trees. (B) Neighbor-joining tree (redrawn from Hass, 1996).

Essentially no phylogenetic work has been conducted with *Thecadactylus* (*rapicauda*; the genus is monotypic). By implication of Kluge's (1967) classification of *Thecadactylus* into the Gekkoninae, the genus is related to other West Indian genera such as *Aristelliger* and *Tarentola*. Kluge (1967) inferred, based on the absence of postanal slits, *Thecadactylus* probably shared a close history with *Aristelliger* and *Lygodactylus*. Hoogmoed (1973:64) on the other hand considered Kluge's (1967) hypothesis "a little far fetched" and opined that the loss of postanal sacs, which Kluge considered a synapomorphy, was independent.

Tarentola was revised by Joger (1984) and he placed the single New World species into its own subgenus, *Neotarentola*. The phylogenetic relationships of this taxon were not discussed

Iguanidae

The cladistic revolution in classification, that being the storage of information of monophyly and sister relationships instead of grades, brought about great changes in large groups that were subjected to cladistic analysis and subsequently re-classified. The Iguanidae of 10 years ago bears little resemblance to the current family in terms of its contents due to the efforts of Frost and Etheridge (1989). Following Frost and Etheridge, the Caribbean has only two representative genera, *Iguana* and *Cyclura*. The former contains two species (*iguana* and *delicatissima*) and the latter contains eight extant species (*ricordi*, *carinata*, *nubila*, *cyclura*, *cornuta*, *collei*, *rileyi*, *punguis*). Two taxa known only from skeletal material (*mattea* Miller, 1918; *portoricensis* Barbour, 1919) were sunk into synonymy with *punguis* by Pregill (1981b).

Barbour and Noble (1916) and Schwartz and Carey (1977) considered *Cyclura* to be the sister to *Ctenosaura*. Avery and Tanner (1971) proposed that *Cyclura* was the sister to a clade composed of *Ctenosaura* and *Sauromalus*. Kevin de Queiroz (1987) demonstrated that *Iguana* and *Cyclura* were each others closest relatives and a subsequent analysis (Norell and de Queiroz, 1991) with fossil taxa found *Cyclura* to be the sister of an *Iguana-Pumilia* (extinct) clade. The Norell and de Queiroz (1991) analysis fully resolved the Iguanidae phylogeny, which had previously been considered as part of an unresolved trichotomy with *Ctenosaura* and an *Amblyrhynchus-Conolophus-Sauromalus* clade (de Queiroz, 1987; Etheridge and de Queiroz, 1988; Frost and Etheridge, 1989). The most recent estimate of iguanid phylogeny (Sites *et al.*, 1996) combined the morphological data with sequences from two regions of

mtDNA. Sites *et al.* performed a rigorous phylogenetic analysis which yielded a strikingly different hypothesis from that of Norell and deQueiroz (1991). Sites *et al.* found *Cyclura* to be the sister to a large clade that included *Ctenosaura*, *Amblyrhynchus*, *Conolophus*, *Iguana* and *Sauromalus* (Fig. 8.4). In this same study, *Iguana* and *Sauromalus* were supported as sister taxa. Given the nature of the Sites *et al.* (1996) study, their hypothesis must be considered the current best estimate of the placement of *Cyclura* and *Iguana*, although one must keep in mind that the novel hypothesis may be a result of independent evolution of the mtDNA. Interestingly, *Cyclura* appears to have diverged relatively early in the history of the iguanas, perhaps hinting that the divergence was associated with the earliest proto-Antilles vicariance events.

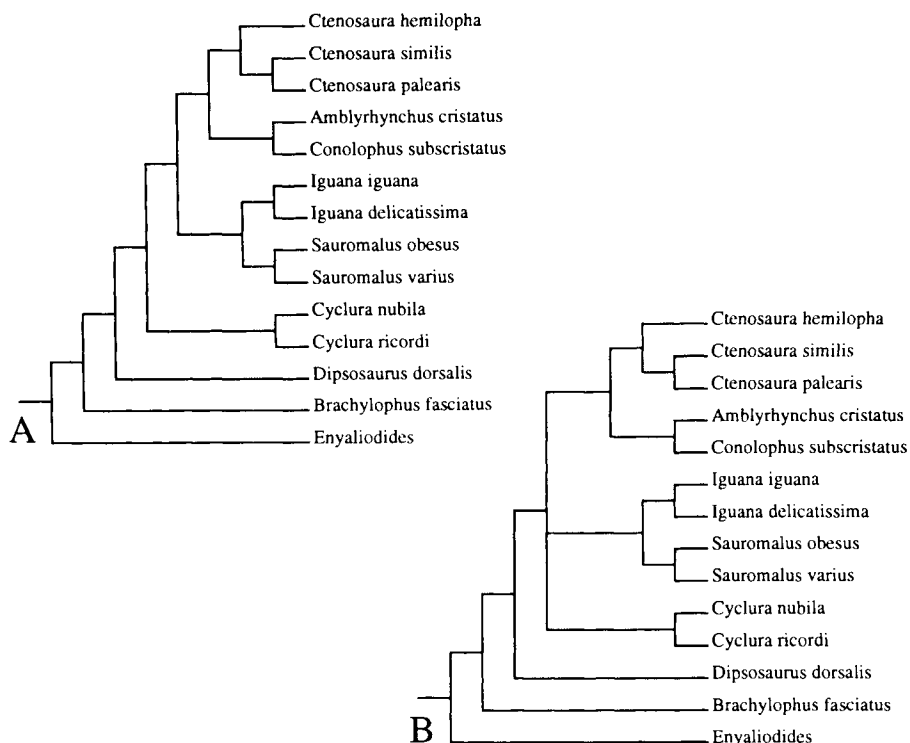


Figure 8.4. Phylogeny estimate of iguanid taxa based on mitochondrial DNA. (A) Maximum-parsimony tree with characters of three or more homoplasies deleted, (B) Consensus tree of the two most parsimonious trees derived from consistent characters only (redrawn from Sites *et al.*, 1995).

Given that *Iguana* contains only two species, which was verified by Lazell (1973), it is safe to say that the phylogeny is worked out. Perhaps the nagging question about *I. iguana* in the Caribbean is whether or not they are distinct from the mainland or are recent imports through Carib Indian movements. Lazell (1973) concluded that the Caribbean populations were distinct and as such populated the islands via natural means and not human transport. A recent report by Censky *et al.* (1998) of *I. iguana* dispersal in the LA corroborated Lazell's hypothesis.

The intrageneric phylogeny of *Cyclura* has been discussed and estimated, with the first key study addressing the question being that of Schwartz and Carey (1977). Based on the examination of 39 external morphological characters, they proposed a hypothesis of evolutionary history. They suggested that two groups were present, one composed of *ricordi* and *carinata* and the other composed of the remaining taxa (the fossil taxa were not included in the study). In the latter group, they believed that *pinguis* was outside of the other five taxa, and the relationships of these five could not be resolved with respect to each other. Schwartz and Carey (1977: 90) made a peculiar observation and conclusion about a subspecies of *rileyi* and the species *cyclura*,

It well may be that *C. r. cristata* should be regarded as either a distinct species, or that it should be combined nomeclatorially with *C. cyclura*. Neither course recommends itself; the first beclouds the affinities, the second violates our concepts of *cyclura* both in juvenile pattern and in head scutellation.

This problem is instructive in that it suggests homoplasy may be a factor in resolving this group, and that if *rileyi* is in fact a paraphyletic species, fundamental systematic problems remain in our understanding of *Cyclura*.

Hollingsworth (1998) inferred the phylogeny of *Cyclura* while attempting to understand the history of *Sauromalus*. Hollingsworth examined morphology and included polymorphic characters and coded them for analysis two ways. One approach followed Campbell and Frost (1993, polymorphisms were assigned discrete states) and the other coding scheme followed Wiens (1995) in which the frequencies of the polymorphisms were coded into bins. Regardless of the potential problems associated with frequency data of any kind (e.g. Crother, 1990; Murphy, 1993), Hollingsworth (1998) corroborated Schwartz and Carey's (1977) hypotheses with the exception that Hollingsworth found *pinguis* as the

sister to all the *Cyclura* (((*rileyi*, *collei*)*nubila*)*cornuta*, *cy-chlura*)(*ricordii*, *carinata*)).

At the time of this writing, there is an ongoing study using mtDNA sequence to address the phylogeny of *Cyclura* (C. Malone, personal communication).

Polychrotidae

The representative taxa of this iguanian family include the anole genera *Chamaelinorops* (monotypic), *Chamaeleolis* (three species), and *Anolis* [140 species; whether *Anolis* is composed of multiple genera sensu Guyer and Savage (1986) remains in debate; see below for further discussion]. It is clear that these three genera plus *Phenacosaurus* form a monophyletic group (Etheridge, 1960; Williams, 1977; Peterson, 1983a; Guyer and Savage, 1986; Frost and Etheridge, 1989), and this "anole" group seems clearly related to *Polychrus*. In contrast, the hypothesized relationships among these genera have varied. The phylograms of Etheridge (1960, redrawn here as Fig. 8.5) indicate a paraphyletic *Anolis*, with the other genera internested. My interpretation of Etheridge differs from the tree of Paull *et al.* (1975; also a redrawing of Etheridge) even though they also depict a paraphyletic *Anolis* (Fig. 8.5). Williams (1977) presented another hypothesis and he drew a monophyletic *Anolis* with *Phenacosaurus* as the sister (((alpha, beta) *Phenacosaurus*) *Chamaeleolis*) *Chamaelinorops*) (Fig. 8.5). Wyles and Gorman (1980) presented immunological distance data that they interpreted as suggesting *Chamaelinorops* was more closely related to other Hispaniolan *Anolis* than were other *Anolis* (thus implying a paraphyletic *Anolis*). Case and Williams (1987) employed morphology and allozymes to test the Wyles and Gorman (1980) hypothesis and successfully rejected the paraphyly notion as described by Wyles and Gorman (1980). Guyer and Savage (1986), in their provocative reanalysis of anole phylogeny, also found a monophyletic *Anolis* (sensu Etheridge, 1960) relative to the other anole genera. Hass *et al.* (1993) brought mtDNA and immunological data (ID) to the question of *Chamaeleolis*, *Chamaelinorops*, and *Anolis* relationships (they did not include *Phenacosaurus*). Based on their ID and mtDNA trees they concluded that *Anolis* was paraphyletic with regard to the other two genera. An interesting observation on the Hass *et al.* (1993) study was the choice of outgroups. The distantly related tropidurid genus *Leiocephalus* was used as the outgroup for the ID analysis. In the mtDNA analysis, *Leiocephalus*, *Polychrus*, and *Eumeces* were used and tellingly,

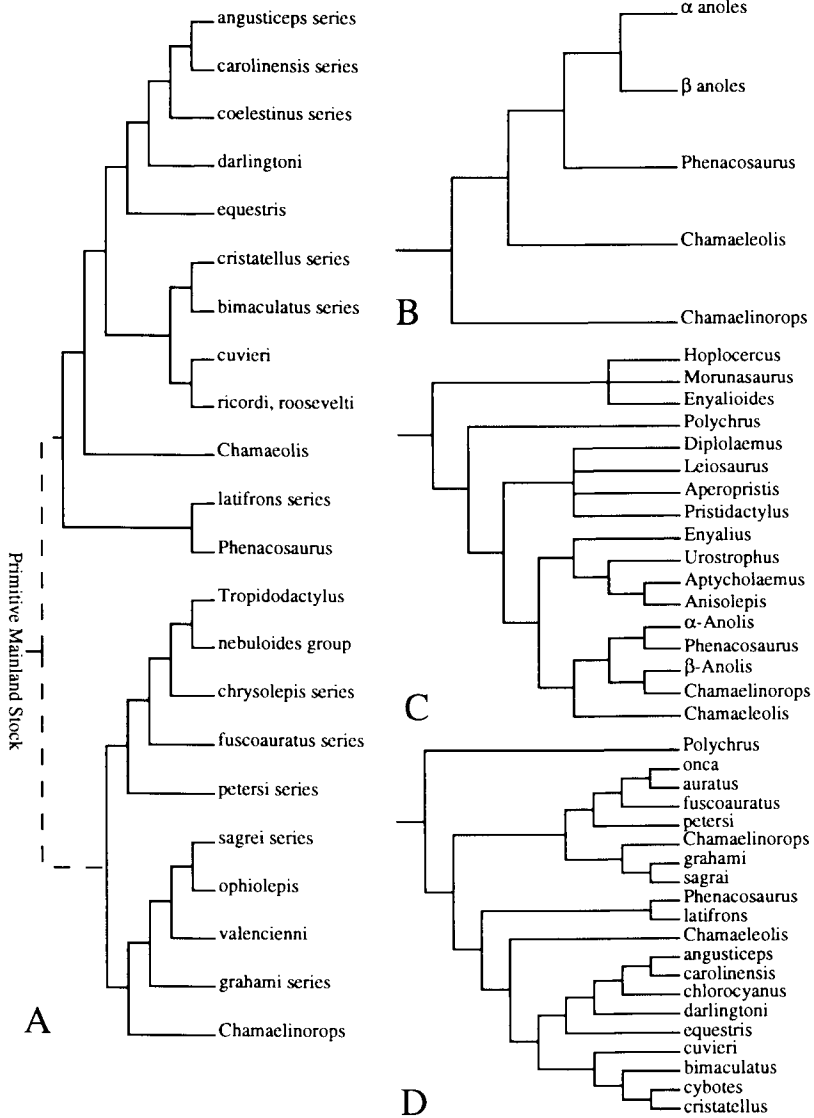


Figure 8.5. Phylogeny estimates of higher level anoline relationships. These are essentially interpretations of Etheridge, 1960. (A) Interpretation herein. (B) Hypothesis of Williams, 1977. (C) Interpretation of Etheridge by Paull *et al.*, 1975. (D) Interpretation of Etheridge by Guyer and Savage, 1986.

Leiocephalus, and not *Polychrus*, fell out as the sister to the anoles. Morphologically, evidence is lacking for a *Leiocephalus* - anole relationship (e.g., Etheridge, 1960, 1966; Frost and Etheridge, 1989; Pregill, 1992), whereas evidence for a closer *Polychrus* - anole relationship is not lacking (e.g., Etheridge, 1960; Peterson, 1983a,b; Guyer and Savage, 1986; Frost and Etheridge, 1989). As far as the relatively simple question of intergeneric relationships is concerned, there currently is no resolution.

Resolving the relationships within *Anolis* (*sensu* Etheridge) is a far more daunting task than estimating the phylogeny of anole genera. It is daunting for the simple reason that there are so many taxa. Savage and Guyer (1989) listed 286 species total, and of these 140 are found in the Caribbean islands and the Bahamas. What is so difficult about 140 taxa is that the approximate number of rooted trees that one must whittle down to a single phylogeny is about 3×10^{279} , which is far greater than the number of particles in the known universe. Given that, is it any wonder that although this group has received much attention over the past couple decades, little has been agreed on by the various workers?

Before I discuss the various interspecific studies, I want to give attention to the study by Guyer and Savage (1986). Guyer and Savage synthesized the osteological data of Etheridge (1960), the multitude of immunological distance studies, and the available karyotypic data to construct a consensus tree of anole relationships. Based on this synthesis they subsequently re-classified *Anolis* and broke it into several genera: *Anolis*, *Ctenonotus*, *Dactyloa*, *Norops*, and *Semiurus*. The erection of these genera was based on the discovery of five monophyletic groups in their final tree, which in itself is not such a controversial method. Although Schwartz and Henderson (1988) immediately embraced the Guyer and Savage classification, a subsequent volume (Schwartz and Henderson, 1991) did not, and two pointed rebuttals (Cannatella and de Queiroz, 1988; Williams, 1989a) to Guyer and Savage followed. Guyer and Savage (1992) contested the arguments against their original study, added more data, and conducted further analyses. Interestingly, the 1992 work by Guyer and Savage is almost never cited and the reasons given for the consistent dismissal of their phylogeny and classification are based on the Cannatella and de Queiroz (1988) and Williams (1989a) rebuttals. Have the Guyer and Savage (1986, 1992) studies and subsequent classification been given a fair scientific treatment? I recommend that all current and aspiring anolologists reread these four papers and pay special attention to the Guyer and Savage 1992 paper in which they argued point

by point their work, included suggestions of Cannatella and de Queiroz and Williams, add more data to the analysis, and yet resolve essentially the same set of relationships (Fig. 8.6). I find these to be compelling reasons why the Guyer and Savage studies should not be overlooked and should be subjected to the same test of rejection as any other hypothesis. Whether or not their hypothesis is rejected, it is certain that Guyer and Savage stirred up much activity toward resolving the immense problem of anole phylogeny and classification.

Below the genus level, hypotheses of anole relationships are many and are largely incongruent. Although there are earlier systematic studies on anoles, the dissertation of Etheridge (1960) is effectively the baseline for all subsequent anole phylogeny hypotheses. Etheridge proposed two main divisions of anoles, the Alpha Section and the Beta Section. Both sections have representatives in the West Indies, dominated in species numbers by the Alpha Section. Etheridge figured phylogenies for both sections but mostly at the level of series (Fig. 8.5). In general Etheridge considered the sections monophyletic with respect to each other and that they were derived from a mainland ancestor(s). The Beta Section was confined to Jamaican and Cuban forms and the Alpha section was found elsewhere throughout the Caribbean region.

Lazell (1966) examined the relationships of the Jamaican *Anolis reconditus* and figured a theoretical schema of four taxa which he considered a monophyletic group. Lazell disagreed with Etheridge (1960) by not considering *garmani* part of this group. Lazell (1972) later turned his attention to the Lesser Antillean anoles. Here, he presented two diagrammatic schemes which depicted the hypothesized temporal sequence of colonization events coupled with relationships. These figures are difficult to interpret, and Lazell readily admits his diagrams are not objective renderings. He stated (1972:98),

To reconstruct the evolution of the groups on an objective foundation is impossible. I shall reconstruct it, then, on a subjective basis. The result cannot be shown to be either right or wrong; one need not believe it or disbelieve it; there is no need to like it or dislike it, there is no necessity to even look at it. In what follows, I shall make abstract representations of real objects and arrange them in patterns that appeal to me. The couching of phrases in speculative terms now ends, but I practice no deception. Even as a mobile constructed out of odd bits of hospital apparatus is not medicine, so this is not science. In all respects, it is the purest of art.

One certainly cannot object to or criticize such honesty. Thankfully, however, the epistemological, theoretical, and empirical foundations of modern systematics are solid, and as such it can be argued that phylogenetics is science, not art, and thus the hypotheses are correctly viewed as ideas open to refutation.

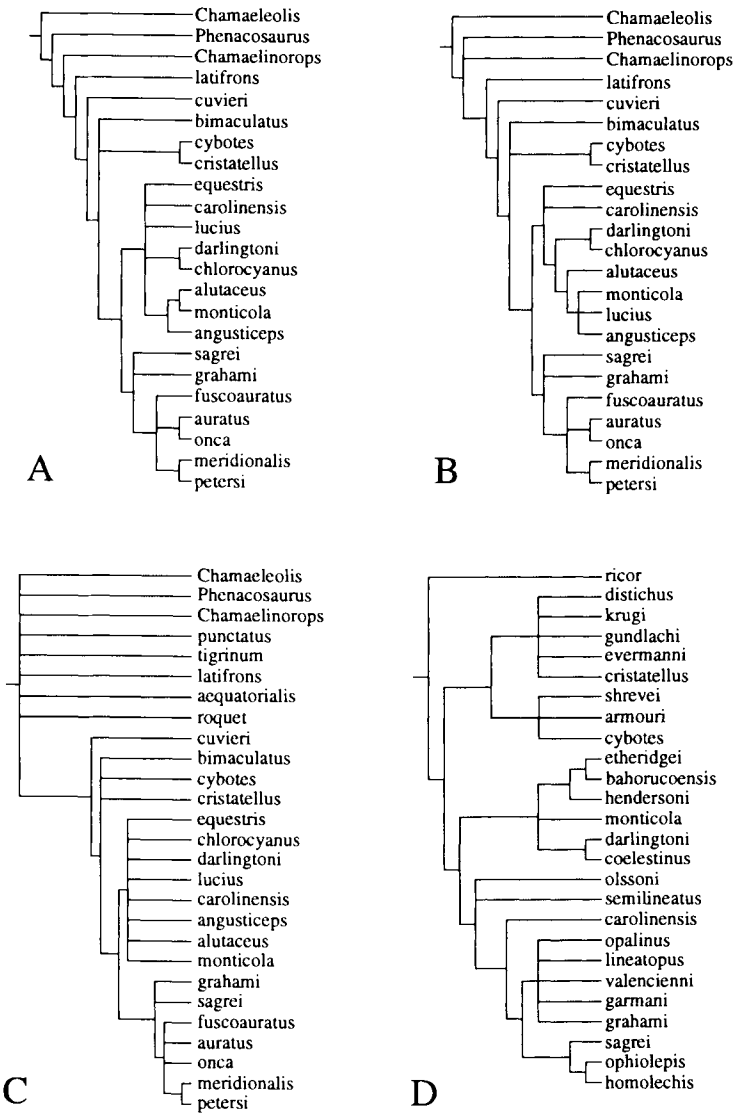


Figure 8.6. Phylogeny estimates of anoline lizards derived from the parsimony analysis of osteological data. (A) Estimate based on modified data set from Guyer and Savage (1986); successive approximation weighted analysis. (B) Estimate based on modified data set from Cannatella and de Quieroz (1989); successive approximation weighted analysis. (C) Estimate based on modified data set from Williams (1989). Unweighted analysis. (D) Estimate from combined osteology, allozyme, and karyotype data recovered with successive approximation weighted analysis (redrawn from Guyer and Savage, 1992).

Gorman and associates were the first to consistently put forth phylogenetic hypotheses of anoles based on objective criteria, and they remained essentially the lone group to do so for almost 20 years. Albeit, it can easily be argued that the first phylogenies were every bit as subjective as Lazell's.

Gorman and Atkins (1969) figured a proposed phylogeny of *bimaculatus*, *acutus*, and Puerto Rican species groups based on karyotypes and lactate dehydrogenase mobility. Although the *roquet* group was also discussed, a phylogeny was not forthcoming until 1974. Yang *et al.* (1974) produced a genetic distance-based dendrogram which was interpreted as an estimate of phylogeny. The phylogeny was at odds with the Gorman and Atkins (1969) tree and thus forced a reevaluation of their colonization scenario. For heuristic purposes, I recoded the Yang *et al.* allozyme data set with the locus as the character and the alleles as the states. Where polymorphisms occurred, frequencies >0.90 were considered fixed and frequencies <0.90 were coded as uncertain. Based on Gorman and Atkins (1969), *Anolis luciae* was used to root the tree alone and together with *blanquillanus* and *bonairensis*. A branch and bound search on PAUP 3.1.1 (Swofford, 1993) recovered 56 most parsimonious trees at 26 steps with high descriptive statistics (CI = 0.938, RI = 0.923, RC = 0.888). However, the strict consensus tree yielded a complete polytomy of the ingroup taxa except for *blanquillanus* and *bonairensis*, which formed a separate clade.

In 1976 Gorman and Kim took the same approach as Yang *et al.* (1974) to estimate the relationships of the other Lesser Antillean anoles: the *bimaculatus* group. A single fully resolved dendrogram was obtained. Following the protocol I described previously, I also reanalyzed these data. For this analysis I employed the *wattsii* complex and *acutus*, as discussed by Gorman and Kim (1976), as outgroup taxa. The branch and bound search was stopped at 4200 most parsimonious trees, each at 21 steps (CI = 0.905, RI = 0.889, RC = 0.804). The strict consensus tree corroborated some of the clades found by Gorman and Kim. The biggest surprise was that *acutus* (considered outside of the other groups) was internested within the *bimaculatus* group as the sister to *pogus*, results which are apparently at odds with chromosomal and morphological evidence. When *acutus* was used as the sole outgroup, the monophyly of the *wattsii* series could not be substantiated. These results are at odds with widely held views of anole relationships (e.g., Williams, 1976; Roughgarden *et al.*, 1987; Roughgarden, 1995). However, Roughgarden *et al.*

(1987) and Roughgarden (1995) also disagree with Gorman and Kim (1976) and the reanalysis herein in the placement of *leachi* and *bimaculatus* (Fig. 8.7). The relationships of the eastern Caribbean anoles remain unresolved, probably for the reasons discussed later.

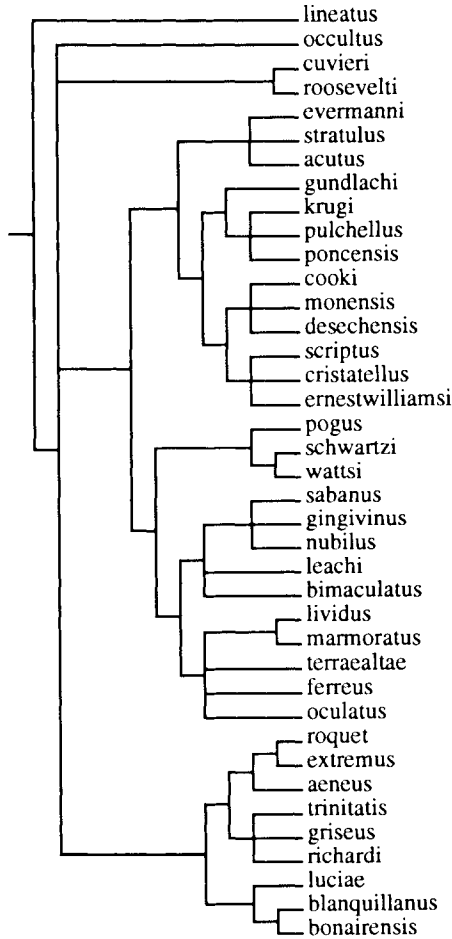


Figure 8.7. Phylogeny estimate of eastern Caribbean anoles based on squamation, karyotypes, allozymes, and immunological data. This is essentially a hand drawn consensus tree of the aforementioned data. The relationships in the *bimaculatus* group are the same as in Roughgarden (1987) and Losos (1992) (redrawn from Roughgarden, 1995).

The phylogenetic relationships of Puerto Rican anoles have been figured several times (Williams, 1972; Gorman *et al.*, 1980a, 1983; Burnell and Hedges, 1990; Roughgarden, 1995). Williams (1972) synthesized osteological, allozyme, and karyotypic data to arrive at his phylogeny. As an interesting aside compare the following quote from Williams (1972:74) about his feelings on his proposed phylogeny with the previous quote of Lazell (1972): "The several kinds of evidence are in very good agreement and the dendrogram can be taken to be a highly probable representation of real relationships."

Gorman *et al.* (1980a), in an incomplete sampling of the Puerto Rican taxa found *monensis* and *cooki* to be sisters and *cristatellus* paraphyletic with regard to *scriptus*. Later, Gorman *et al.* (1983) employed allozymes to examine the relationships of Puerto Rican bank anoles and anoles from elsewhere. They conducted two studies, one with all 10 Puerto Rican species and five others, and a second with eight Puerto Rican species and three others. Both a phenogram (UPGMA) and a cladogram (Wagner) were constructed for each study. The results were incongruent among and between studies and methods. Based on these results and karyotype data, Gorman *et al.* (1983) also presented a summary cladogram. None of the Gorman *et al.* (1983) trees were congruent with Williams' (1972) hypotheses, the main reason probably being the non-monophyletic nature of the Puerto Rican assemblage. The various analyses in Burnell and Hedges (1990) and the schema of Roughgarden (1995) further suggest that the Puerto Rican anoles are not monophyletic. Given that, attempts at understanding the phylogenetic relationships of Puerto Rican anoles without broad taxon sampling are/were doomed to inaccuracy.

Like Puerto Rico, Jamaica has an enticingly small number of anole species and as such has been subjected to phylogenetic analysis under the assumption that the assemblage is monophyletic (with the exception of *sagrei*). Underwood and Williams (1959) evaluated the Jamaican anoles and offered a systematic arrangement for them. Hedges and Burnell (1990) drew a phylogeny of Jamaican anoles as monophyletic, which they considered an interpretation of Underwood and Williams (1959). Interestingly, Underwood and Williams (1959) did not propose a monophyletic radiation of Jamaican anoles, and in fact were convinced otherwise (Underwood and Williams, 1959:9): "These are grouped only as we see them in the island. Without a doubt the Jamaican groups are only parts of more widely distributed groups." Lazell (1966) could be inter-

puted in the same way: if *garmani* and *valencienni* were not related to the other Jamaican taxa, did their relationships belong elsewhere?

In the two distance analyses of their allozyme data of 49 West Indian species, Burnell and Hedges (1990) found the Jamaican anoles monophyletic, including *sagrei*, but in the parsimony analysis they formed two groups, one related to Hispaniolan forms and the other related to members of the *carolinensis* series. Examination of the bootstrap values shown on these trees may hint at the robustness of the data and thus of the trees. The best of the three trees had only 3/44 (6.8%) nodes with a bootstrap proportion >80%. The monophyly of Jamaican anoles should be questioned.

Hedges and Burnell (1990) argued that they demonstrated the monophyly of Jamaican anoles by finding them monophyletic relative to five other species (and with *sagrei* as the only beta anole). I argue that monophyly relative to five other taxa when another 200+ species exist is not a clear demonstration. Although I think Hedges and Burnell may be correct because of historical biogeographic considerations, their demonstration is not convincing.

The anole assemblages of Cuba and Hispaniola are large in number (54 and 42 species, respectively) and decidedly nonmonophyletic. As such, no detailed phylogenetic hypotheses of all the taxa have been proposed. However, Poe (1998) included 37 species in a study of twig anole (Williams, 1983) relationships. His combined data set approach rejected twig anole monophyly and corroborated the hypotheses that the Cuban and Hispaniolan radiations were non-monophyletic. Regardless, estimation of the phylogenies of the radiations will only be forthcoming in large, inclusive studies which attempt to understand anoles as a whole.

Overall, the systematics of the West Indian anoles (as well as the rest of the anoles) remain in flux, although this was largely not the case after the publication of Williams' (1976,a,b) two influential papers on the classification of anoles. He used a number of formal and informal categories to imply relationship (monophyly?), and these implications have been widely followed since [although Williams (1989a) has argued otherwise]. Guyer and Savage (1986) figured the implied relationships of Etheridge (1960) and/or Williams (1976a,b) (Fig. 8.5).

Although the Etheridge/Williams scheme has been widely used, it has also been challenged. The assumptions of monophyly and relationship have been questioned numerous times from multiple data sources; for example, Gorman *et al.* (1980), Wyles and Gorman (1980), Shochat

and Dessauer (1981), Guyer and Savage (1986, 1992), Burnell and Hedges (1990), and Hass *et al.* (1993). Gorman *et al.* (1980b) used albumin immunological data and found incongruence between their study and their interpretation of Williams (1976a,b) for eastern Caribbean anoles. Wyles and Gorman (1980) used immunology again and could not support the classic relationships based on osteology. Shochat and Dessauer (1981) followed along the same path and noted discrepancies. The studies by Guyer and Savage (1986, 1992) were discussed previously and suffice to say they challenged the classic classification based on multiple lines of evidence. Burnell and Hedges (1990) employed data derived from sequential electrophoresis, found their results incongruent with the Etheridge/Williams scheme, and reclassified the West Indian anoles following the category conventions of Etheridge and Williams. Hass *et al.* (1993) examined albumin immunology and 16S mtDNA sequence and also found relationships at odds with Etheridge and Williams.

Although the previous studies all reject the Etheridge/Williams ideas of anole relationships, they are not all in agreement among themselves (see the accompanying figures). Several probable reasons exist for the incongruence: sample, methodology, and data. Exemplar sampling or incomplete samples, while perhaps sometimes necessary, can result in misleading phylogenies and certainly incongruence when different studies include different samples. Inconsistent methodological approaches can lead to perceived incongruences, when in fact the data indeed covary on the same history (Kluge, 1991). Different data sets, although they theoretically covary on the same history, can also result in different hypotheses (e.g., gene trees versus species trees). Of course, the combination of these three problems probably acts synergistically to exacerbate the incongruency. With anole phylogeny, I suspect this is what has occurred.

Where does that leave anole systematics? At the time of this writing a number of labs are bringing DNA sequences, more complete taxon sampling, and I predict consistent phylogenetic approaches to test the controversial ideas of not only Guyer and Savage but also Etheridge, Williams, and all the rest who have sought to understand anole phylogeny.

Scincidae

The Scincidae is taxonomically depauperate in the Caribbean, with only a single genus and two species present. The genus *Mabuya* is widespread, with a range that includes the Greater and Lesser Antilles (exclusive of Cuba), northern South America into Panama, Africa, Madagascar, and southern Asia. In the region of interest here, one of the two species is endemic to Hispaniola (*lineolata*) and the other (*bistriata*) encompasses the remainder of the West Indian distribution of the genus (Schwartz and Henderson, 1991).

The relationships of New World *Mabuya* remain enigmatic. Dunn (1935) suggested that their affinities were with African forms but that relationships between *mabouya* and *lineolata* were obscured by the fact that *lineolata* was unique and highly differentiated from its geographic counterpart. Since Dunn's comments, no studies have attempted to better understand the phylogeny of *Mabuya*. Perhaps the closest to addressing *Mabuya* phylogeny was Greer's (1986) work on the monophyly of the Lygosominae. Clearly, detailed phylogenetic studies of *Mabuya* are needed and would help clarify the relationships of the Caribbean forms.

Teiidae

This New World lizard family is represented in the West Indies by four genera: *Ameiva*, *Bachia*, *Cnemidophorus*, and *Gymnophthalmus*. As currently understood, there are 20 species of *Ameiva*, 1 species of *Bachia*, 2 species of *Cnemidophorus*, and 2 species of *Gymnophthalmus*. Amazingly (or perhaps not amazing at all), for such a widespread and important radiation there have been no modern, rigorous phylogenetic studies of these taxa. Interestingly though, there have been two schemes of phylogeny drawn, one on *Bachia* and relatives (Dixon, 1973) and the other on *Ameiva* (Barbour and Noble, 1915). Otherwise, any notions of relationship for these are only narrative (one might consider the phylograms as drawn narratives) and some require interpretation.

Barbour and Noble (1915) presented their concept of the relationships of *Ameiva* in diagrammatic fashion (Fig. 8.8). In general, they recognized three distinct groups of *Ameiva*: (1) an *A. ameiva* group that contained the South America, Lesser Antillean, Puerto Rican and Hispaniolan species (*vittipunctata* = *chrysoleama*), (2) an *A. undulata* group with relatives in Cuba, Jamaica, and Hispaniola, and (3) a Bahamian - Puerto Rican bank group.

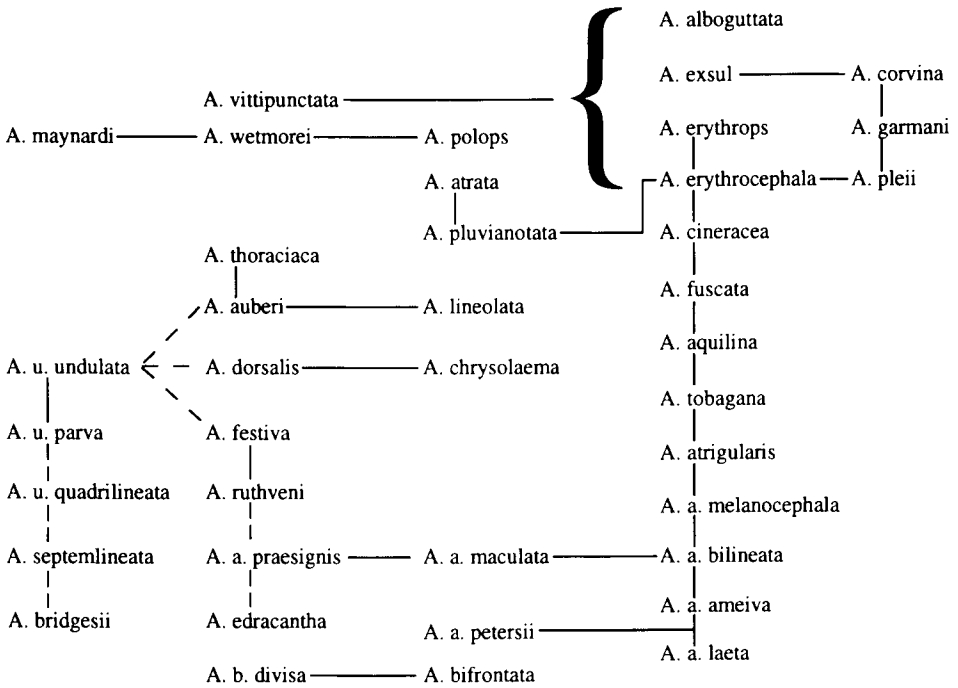


Figure 8.8. Phylogeny estimate of *Ameiva* based on morphology. Solid lines represent close relationship, dashed lines indicate the taxa are related but with unknown taxa intervening (Interpreted and redrawn from Barbour and Noble, 1915).

Cochran (1941) did not agree with Barbour and Noble and in her revisions she repeatedly broke up Barbour and Noble’s second and third groups. For example, Cochran suggested that *wetmorei* and *lineolata* were related and that *polops* and *taeniura* (considered *lineolata* by Barbour and Noble, 1915) were closest relatives, and she synonymized a South American member (*vittipunctata*) with a Central American member (*chrysolema*).

Baskins and Williams (1966) evaluated the LA *Ameiva* and drew some tentative conclusions about relationships. They considered the LA *Ameiva* to be composed of three lineages, a northern group (*erythrocephala*, *griswoldi*, *pleii*, *pluvionotata*, *corvina*, and *fuscata*), a southern group containing *cineracea* and *major*, and a third group composed of

only *ameiva*. Within the northern group, Baskins and Williams only speculated that *erythrocephala* and *griswoldi* were sisters relative to the rest that had uncertain affinities. By default, the two extinct members of the southern group were considered to have sister-taxa status and *ameiva* was related to mainland taxa. They also speculated that if the Lesser Antillean taxa had any relationship to Greater Antillean taxa, it might be with *chrysolema* and/or *exsul*.

Heatwole and Torres (1967) examined the Puerto Rican Bank taxa and concluded that *wetmorei*, *exsul*, and *polops* were not related but derived from independent stocks. They did not speculate as to what these stocks may have been.

Schwartz (1970) proposed a close relationship between *auberi* and *dorsalis*, which is at odds with the schema of Barbour and Noble (1915). He further suggested that *chrysolema* might possibly be part of that group. He also accepted Cochran's (1941) idea that *taeniura* and *polops* were closely related but then added that *maynardi* and *wetmorei* also were relatives of *taeniura-polops*. No mention was made of *lineolata*, which Cochran (1941) thought was related to *wetmorei*. Schwartz (1970) also argued that the entire West Indian *Ameiva* radiation was derived from a South American ancestor.

Dixon (1973) reviewed the relationships of *Bachia* and related genera and presented in a phylogram his concept of phylogeny. The lone West Indian form, *B. heteropus* (found on the Grenada Bank and northeast South America), was depicted as being the sister to *pallidiceps*, which is a northern South American taxon that ranges into Panama.

The species of *Cnemidophorus* are confined to tiny islands in the western Caribbean and the LA. *Cnemidophorus lemniscatus* is widespread on the Central and South American mainland and is also found on two small islands off Nicaragua. The other species, *vanzoi*, is restricted to an islet of St. Lucia. Initially, *C. vanzoi* was described as an *Ameiva* (Baskins and Williams, 1966), but Presch (1971) demonstrated that it belonged to *Cnemidophorus* based on distinctive tongue morphology. As an *Ameiva*, the distribution made some sense, as a *Cnemidophorus*, it appears highly unusual. Presch (1971) considered *vanzoi* to be a member of the *lemniscatus* group.

Thomas (1965d) reviewed the *Gymnophthalmus* of the southeastern Caribbean and concluded that *G. pleei* (found on the central LA islands of Guadeloupe, Dominica, Martinique, and St. Lucia) was related to *G. lineatus* (Curaçao and mainland). The other species, *G. underwoodi*

(Guadeloupe, St. Vincent, Barbados) could not be placed but Thomas (1965d) suggested its affinities lay with mainland forms.

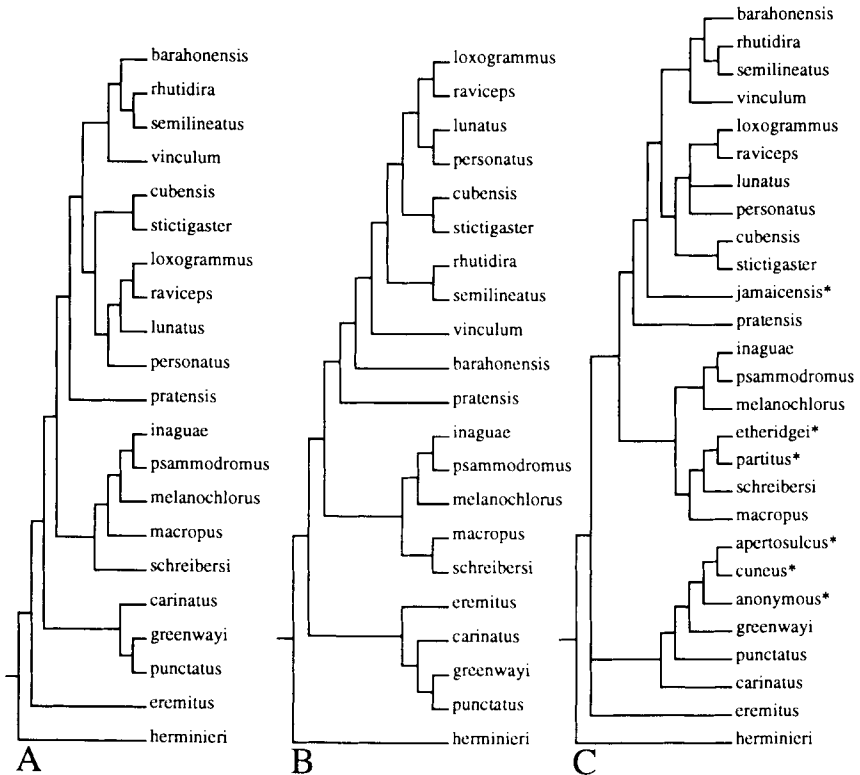


Figure 8.9. Phylogeny estimates of *Leiocephalus* based on parsimony analyses of morphological data. (A) Tree derived from an analysis with the software HENNIG86. (B) Tree derived from an analysis of the same data as A but with the software PAUP. (C) Strict and Adams consensus tree of parsimony analysis which included fossil taxa (denoted by asterisks) (redrawn from Pregill, 1992).

Tropiduridae

A single endemic genus (*Leiocephalus*, as diagnosed by Etheridge, 1966) representative of the Tropiduridae is found in the Caribbean islands. Frost and Etheridge (1989) proposed that *Leiocephalus* was the sister taxon to a South American clade composed of the *Stenocercus* group, *Tropidurus* group, and *Uranoscodon*. Pregill (1992) supported the Frost and Etheridge (1989) hypothesis with additional morphological characters.

The genus *Leiocephalus* is fairly diverse in numbers of species, and they are arguably the most distinctive terrestrial lizards on the islands. There are either 21 or 22 extant species (depending on whether you refer to Pregill, 1992, or Schwartz and Henderson, 1991) and all possess a remarkably upwards curled tail [thus the “curly-tailed lizard” moniker; Pregill (1992) colorfully described the tail as resembling a “coiled watch spring”]. In addition to the extant species, there are two recently extinct and another six which are probably late Holocene in age but no older than late Pleistocene (Pregill, 1992).

Schwartz was by the far the most prolific worker on *Leiocephalus*, publishing no less than 16 papers on variation and taxonomy in the genus (see Pregill, 1992, for reference list). However, the only intrageneric phylogeny proposed was by Pregill (1992). The phylogeny was morphology based, included the fossil taxa, and was analyzed within a cladistic framework. The evidence was well documented and the phylogenies fairly well supported. As such, the conclusions of Pregill (1992) should be considered good estimates of *Leiocephalus* phylogeny. The trees without fossil taxa differ somewhat from the trees with fossil taxa, so both are depicted herein (Fig. 8.9). The phylogenies suggest *Leiocephalus* had a complex evolutionary history, i.e., the island radiations were not found to be monophyletic and may reflect the complex geological history of the Caribbean region or indicate a taxon of great dispersal abilities.

Xantusiidae

A single member of this enigmatic family is found in the Caribbean: the Cuban endemic *Cricosaura typica*. Superficially, with the family containing only three (or possibly four) genera, the phylogenetic problem would appear to be simple. This, however, has not been the case, with the frustration aptly described by Bezy (1972:1): “Not only have xantusiid lizards been troublesome to students of ‘higher classification,’ but

those unfortunate taxonomists who have been lured into extensive studies of the systematics of the family have suffered greater torments.” Over 25 years later Bezy’s comment remains true.

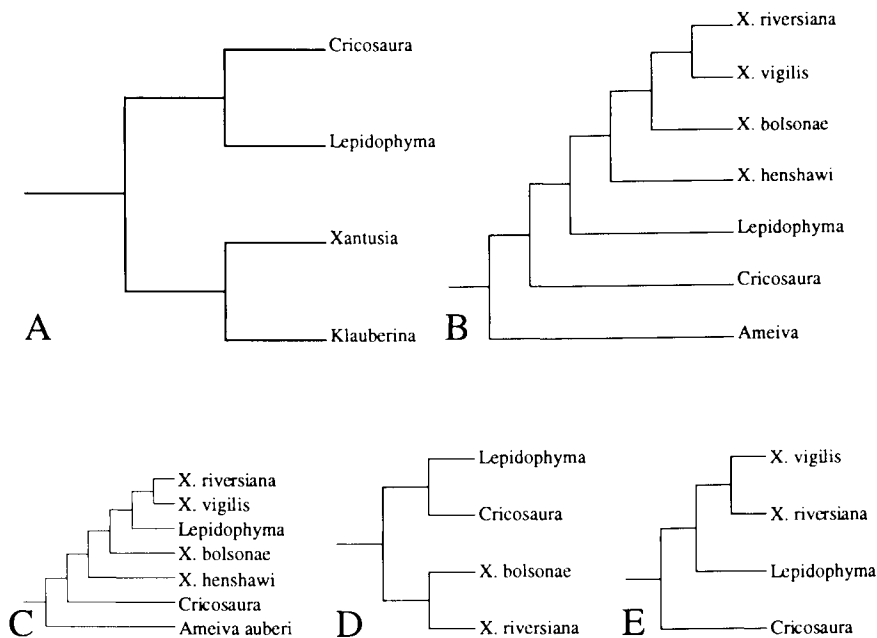


Figure 8.10. Phylogeny estimates of *Cricosaura*. (A) Estimate derived from the phylogenetic analysis of morphological characters (redrawn from Crother *et al.*, 1986). (B) Estimate derived from analysis of mtDNA sequence (redrawn from Hedges *et al.*, 1991). Neighbor-joining tree from combined sequences. Branch lengths do not correspond to distance estimates. (C-E) Phylogeny estimates of *Cricosaura* relative to other xantusiids based on reanalyses of mitochondrial DNA used in Hedges (1991) and with added morphological data (redrawn from Crother and Presch, 1993). All trees are based on parsimony analyses. (C) cytochrome b data alone, unweighted, ambiguities removed, (D) combined molecular and morphological data unweighted with base ambiguities, (E) combined data weighted with base ambiguities.

Savage (1955, 1963, 1964) can be considered the first worker to discuss the evolutionary relationships of *Cricosaura* to the other xantusiid genera. Savage (1955, 1964) examined external and osteological morphological characters and suggested that *Klauberina* (= *Xantusia riversiana*) was the ancestor of the other genera but that *Cricosaura* was its closest ally, although distantly related. So distant in fact, that Savage (1963) placed *Cricosaura* in its own subfamily. Interestingly, in interpretations of Savage's relationship hypotheses, neither Schatzinger (1980) nor Crother *et al.* (1986) depicted a *Cricosaura*–*Klauberina* clade (Fig. 8.10). Schatzinger (1980), in a study of *Palaeoxantusia*, diagrammed *Cricosaura* as the sister to the rest of the genera and noted that his conclusions were similar to those arrived at by Savage. Crother *et al.* (1986) in a cladistic reanalysis of Savage's morphological data, depicted two different Savage hypotheses, one phenetic and the other evolutionary. The phenetic tree was the same as Schatzinger's diagram, and the evolutionary tree placed *Lepidophyma* as the sister of *Cricosaura*. I am hard pressed to explain the absence of the *Cricosaura*–*Klauberina* hypothesis in Crother *et al.* (1986). Regardless, Crother *et al.* found *Cricosaura* to be the sister taxon of *Lepidophyma*.

Bezy (1972) presented karyotypic data on all the xantusiids except for *Cricosaura*, but Hass and Hedges (1992) later published the karyotype of *Cricosaura*. Hass and Hedges concluded that karyotype data were consistent with a hypothesis they (Hedges *et al.*, 1991) proposed based on mitochondrial DNA. This hypothesis stated that *Cricosaura* was the sister to the other genera. However, based on Bezy's (1972) conclusions concerning the evolution of diploid number and number of acrocentric chromosomes, the assertion of Hass and Hedges (1992) is incorrect. Bezy (1972) suggested that diploid number and acrocentric counts have evolved from high numbers to low numbers. As such, Bezy considered *Xantusia* primitive ($2N = 40$, 11 pairs acrocentrics), *Lepidophyma* derived ($2N = 38$, 9-10 pairs), and, to follow Bezy's logic, *Cricosaura* the most derived ($2N = 24$, 6 pairs). This scenario suggests a *Cricosaura*–*Lepidophyma* relationship, which is at odds with Hass and Hedges (1992) and Hedges *et al.* (1991) but in concordance with Crother *et al.* (1986).

The Hedges *et al.* (1991) study has been the only one to employ DNA sequence to the question of xantusiid relationships, and perhaps as a result of this the authors (Hedges *et al.*, 1991:768) make the claim that their study "provides the first robust estimate of intergeneric relation-

ships in this family” As mentioned previously, they concluded that *Cricosaura* was the sister to the other xantusiid genera (Fig. 8.10). Their conclusions however were challenged by Crother and Presch (1993), who reanalyzed the data and demonstrated that in fact a robust estimate was not achieved by Hedges *et al.* (1991). Instead, the results of the re-analyses yielded all manner of relationships (Fig. 8.10), strongly indicating that the final word on xantusiid relationships is yet to come (Crother and Presch, 1994; Hedges and Bezy, 1993, 1994).

Yet another data set has been examined in xantusiid lizards: scale microstructure. Peterson and Bezy (1985) found *Cricosaura* as the sister to *Xantusia*, but in a later study (Bezy and Peterson, 1988), they retreated from their hypothesis and concluded that scale microstructure failed to resolve the relationships among the xantusiid genera.

What can be concluded about the state of phylogenetic knowledge for *Cricosaura*? Although much effort has been expended on reconstructing the phylogeny of xantusiids, it seems clear that further work remains.

Serpentes

The snake fauna in the West Indies is represented by seven families, 23 genera and 108 species [Powell *et al.*, (1996a) listed 107 species of snakes; I include *Micrurus ruatanus*, an endemic in the western Caribbean]. The Colubridae make up the largest fraction of the assemblage with 17 genera (six endemic) and 48 species (43 endemic). Although the remainder of the families (Boidae, Elapidae, Leptotyphlopidae, Tropidopneidae, Typhlopidae, and Viperidae) contain no endemic genera, all but five of the species are endemic to the West Indies/Caribbean islands. Of the 11 nonendemic species, only two (*Elaphe guttata* and *Diadophis punctatus*) are considered to be human introductions (as such, these two taxa will not be discussed further).

Boidae

Three genera of boids (*Boa*, *Corallus*, and *Epicrates*) are found in the Caribbean islands and none are endemic. *Boa* and *Corallus* are represented by their widespread mainland species, *constrictor* and *hortulanus*, respectively. *Epicrates*, on the other hand, is represented by 10 endemic species, found throughout the Greater Antilles and the Bahamas.

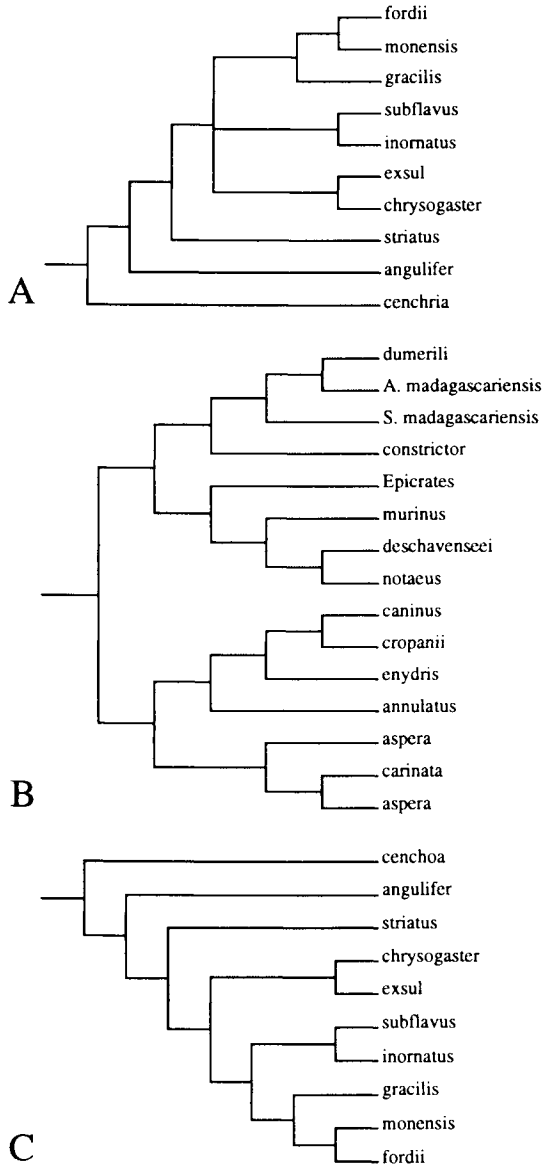


Figure 8.11. Phylogeny estimates of boine snakes. (A) Estimate of the interspecific relationships of the genus *Epicrates* derived from a parsimony analysis of morphological and biochemical data (Kluge, 1989) (A, B, redrawn from Kluge, 1991). (B) Most parsimonious estimate of boine phylogeny based on morphological data (Kluge, 1991). (C) Phylogeny estimate of *Epicrates* based on a parsimony analysis of morphology and lipid data (redrawn from Tolson, 1987).

Differentiation of *Boa constrictor* apparently has not occurred at the specific level in the LA but some variation from the mainland form is present, which led Lazell (1964) to assign subspecific epithets to the island forms. Lazell (1964) considered the island *Boa* to be the end of a step cline. Until recently, *Boa* was monotypic, containing only *constrictor*. Kluge (1991) synonymized *Acrantophis* and *Sanzinia* with *Boa* based on his phylogenetic analysis of boine snakes. According to Kluge (1991) the sister group to *B. constrictor* contains the Madagascan species *manditra*, *dumerili*, and *madagascariensis* (Fig. 8.11). Heise *et al.* (1995) included *B. constrictor* in their mtDNA based phylogeny of snakes and most peculiarly found *Boa* as the sister to colubroids instead of to *Python*, a result not supported by either Kluge (1991) or Cundall *et al.* (1993). The reasons for this peculiar arrangement may be in the mtDNA data set. The mtDNA may not be conserved enough for the question of higher snake phylogeny, the estimate may be befuddled by ancestral polymorphism problems, and it is possible that the phylogeny reflects a gene tree instead of a species tree. (Obviously it is a gene tree. The implication is that the gene evolution did not track the species evolution.) These matters are for discussion elsewhere.

The phylogenetics of the species *Corallus hortullanus* (= *enydris*, McDiarmid *et al.*, 1996) has been addressed at the intrageneric (Kluge, 1991) and intraspecific (Henderson and Hedges, 1995) levels. Henderson and Hedges examined single individuals from diverse geographic localities, including St. Vincent, Grenada, and Trinidad. They used a 307-base pair stretch of cytochrome *b* in the mtDNA to address the monophyly of *hortullanus* and to discover the possible origin of the West Indian populations. Monophyly was supported but the intraspecific results were intriguing in that Panama clustered with Trinidad, whereas the West Indian individuals were sisters, in a clade with Guyana, Brazil, and Peru. At the interspecific level, Kluge found *hortullanus* nested within the other two species of *Corallus* and *Xenoboa* (((*caninus*, *cropanii*) *hortullanus*) *annulatus*) (Fig. 8.11) and given these results he synonymized *Xenoboa* with *Corallus*.

Sheplan and Schwartz (1974) were the first workers to examine the systematics of all the species of Antillean *Epicrates* in a single work. At the time of their work, the systematics of *Epicrates* was, as they stated, "in a chaotic state." Based on the external characteristics of these snakes, they attempted to add order to the chaos. In retrospect, given the recent studies of Tolson (1987) and Kluge (1989), Sheplan and Schwartz (1974)

provided strong predictions about *Epicrates* phylogeny. They suggested two groups, one comprising the smaller-size species and the other comprising the larger species. The small species are apparently monophyletic (with the possible exception of *exsul*), but the larger species are not (Kluge, 1989; Tolson, 1987). Sheplan and Schwartz suggested that *angulifer* was basal, that *inornatus* and *subflavus* were sisters, and that *fordii* and *monensis* were also sister taxa. All these points were reaffirmed by Kluge (1989) and Tolson (1987). The only major difference between Sheplan and Schwartz (1974) and Kluge (1989) and Tolson (1987) was the placement of *exsul*. Kluge and Tolson both relied on morphological, osteological, and skin and scent lipids for their phylogenies (Kluge added 44 more morphological characters to Tolson's original data set). In Kluge's study, when the morphological data were analyzed separately *exsul* was placed in the smaller-sized clade, *a la* Sheplan and Schwartz. Tolson's morphological data alone never supported that arrangement. Tolson and Kluge's studies did differ and their results differed only in the level of resolution (Fig. 8.11). Tolson used *Corallus* as the outgroup, whereas Kluge used a hypothetical ancestor composed of both *Corallus* and *Eunectes*, and of course the two studies differed in that Kluge added 44 morphological characters. Regardless, their total evidence phylogenies were 100% consistent with each other, departing only in the relative positions of *chrysogaster-exsul* and *inornatus* -*subflavus* clades. Kluge's hypothesis was simply unresolved at that point.

At the intergeneric level several hypotheses have been figured for the relationships of *Epicrates*. Kluge (1991) provided interpretations of narratives and redrawings of these relationships, including Schwaner and Dessauer (1981), Underwood (1976), and McDowell (1979). These phylogenies and Kluge's (1991) were all generally similar in depicting the possibility of an *Epicrates*-*Corallus* relationship.

Colubridae

As mentioned previously, the colubrid snakes are represented in the Caribbean/West Indies by 48 species, 43 of which are endemic. Also mentioned previously, two of these (*Diadophis punctata* and *Elaphe guttata*) are considered human introductions and will not be considered further.

Only two species of colubrine snakes are found in the Antilles and both are endemics. *Chironius vincenti* and *Mastigodryas bruesi* are both found on St. Vincent but *C. vincenti* ranges further south throughout the

Grenadines and Grenada. These genera of colubrids have been included in but one phylogenetic study, an extraordinarily ambitious allozyme project conducted to address the phylogeny of snakes (Dowling *et al.*, 1996). Without using *vincenti* or *bruesi*, but using other representatives of these genera, Dowling *et al.* (1996) placed *Chironius* and *Mastigo-dryas* in a large clade composed of a somewhat eclectic collection of colubrids (Fig. 8.12). Do these genera really share a close relationship with *Dasyplestis*?

Dowling *et al.* (1996) used slowly evolving loci as data as discovered through sequential electrophoresis. Typically, allozyme data reach their phylogenetic utility limits at the genus level (intrasubfamilial level?; Buth, 1984). Even with the sequential protocol, clearly the data were strained to construct a snake phylogeny with such ancient origins. This was evidenced by the numerous notations by the authors concerning misplaced taxa. It can be safely said that further work needs to be done on the phylogeny of these taxa.

A single natricine species, *Nerodia clarkii*, is distributed across the northern coast of Cuba in the Antilles, but the extent of the distribution is throughout the coast of the southeastern United States from Texas to Florida. Lawson (1987) conducted the only explicit phylogenetic analysis of *Nerodia* taxa. He used allozymes and performed both discrete (with parsimony) and distance-based analyses. The parsimony tree suggested a *clarkii-sipedon* clade, two of the distance based estimates indicate a *clarkii-fasciata* clade, and the third distance tree shows *clarkii* as the sister to a large clade that contains four other species of *Nerodia* (not including *sipedon* or *fasciata*) and *Thamnophis couchii*. The lack of congruence is a problem, but Kluge (1991) correctly pointed out that incongruence derived from comparison of phylogenies based on differing methodologies and philosophies only serves to obscure the evidence present in the data. Because of my allegiance to the cladistic parsimony paradigm, I suggest that *clarkii-sipedon* might be the best estimate of relationship only tentatively, however, given recent improvements in our understanding of coding allozyme data. Unfortunately, Lawson's (1987) data could not be recoded and reanalyzed because the outgroup data were not included in the publication.

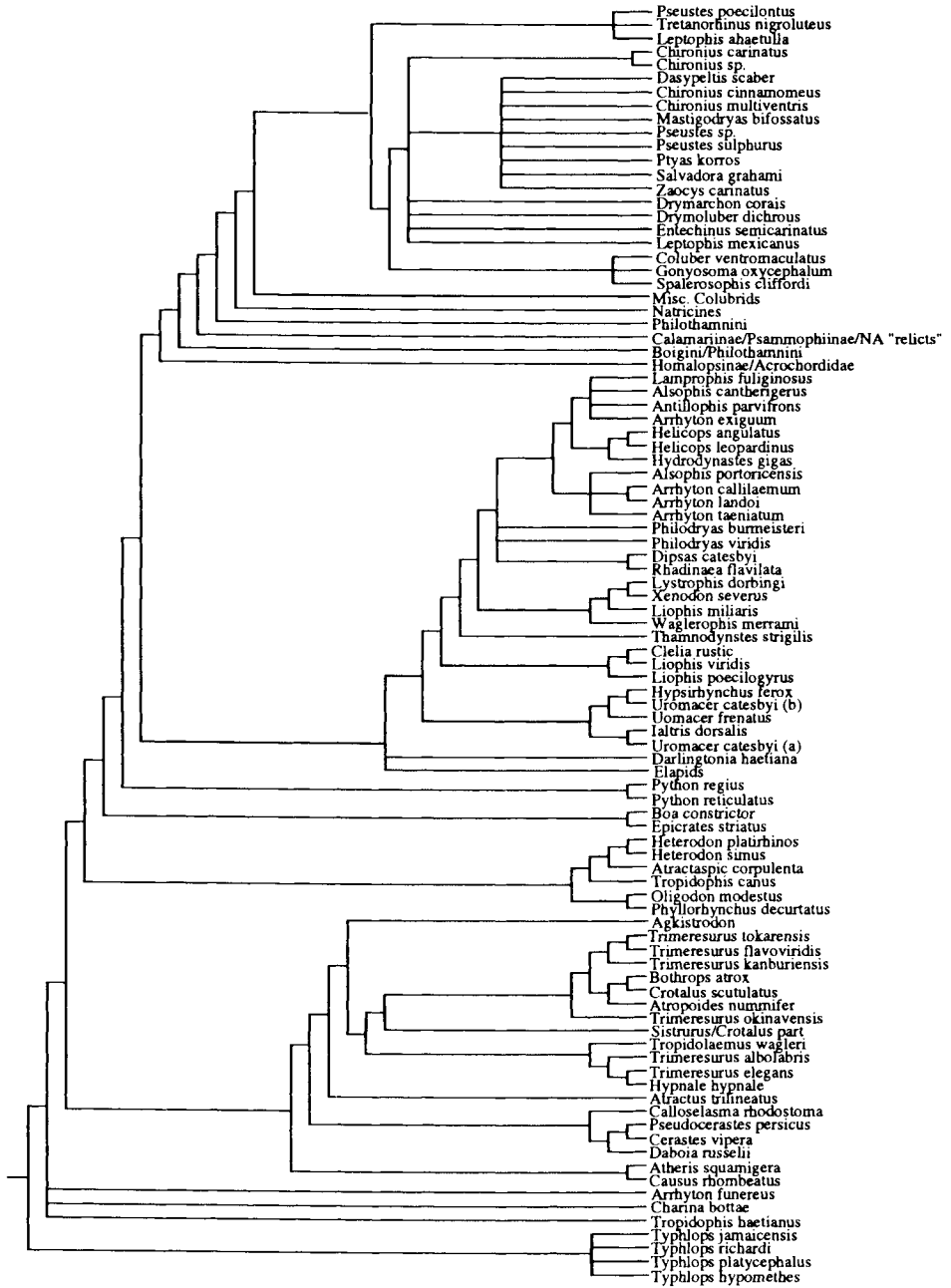


Figure 8.12. Phylogeny estimate of snakes based on a UPGMA analysis of allozyme data (condensed and redrawn from Dowling *et al.*, 1996).

The remaining 43 species belong to the still enigmatic xenodontine-dipsadine group. Several of these taxa have been variously considered part of a single radiation (*Alsophis*, *Antillophis*, *Arrhyton*, *Darlingtonia*, *Hypsirhynchus*, and *Uromacer*) or several unrelated monophyletic groups. Because of this history, these taxa will be considered together. The other genera, *Clelia*, *Coniophanes*, *Liophis*, *Pseudoboa*, and *Tretanorhinus*, have not been considered associated with the former taxa or each other and thus will be reviewed separately.

Two species of *Clelia*, *errabunda* and *clelia*, are recognized on the islands and are both from the southern LA (Underwood, 1993). In a narrative, Underwood (1993) suggested that the previous two *Clelia* species were in the same group as *scytalina* and *equatoriana*. However, Dessauer *et al.* (1987), in a phenogram based on immunological distance data, found *scytalina* and *clelia* to be unrelated, with *Clelia* in fact paraphyletic with respect to *Pseudoboa* and *Oxyrhopus* (which in turn is paraphyletic with respect to *Clelia* and *Pseudoboa*). *Pseudoboa newiedii* was suggested to be the sister to *P. nigra* (Dessauer *et al.*, 1987), and in Cadle (1984a) *Clelia* and *Pseudoboa* are shown as sister taxa. Jenner and Dowling (1985) also show *Clelia* and *Pseudoboa* as sister taxa. Dowling *et al.* (1996), in the same allozyme paper mentioned previously, included a single species of *Clelia* (*rustica*) and it fell out as the sister to *Liophis viridis* (Fig. 8.12).

The western Caribbean island of Isla San Andrés hosts the endemic *Coniophanes andresensis*. No phylogenies have been estimated which include this species but the genus has been explicitly included in phylogenetic analyses (Cadle, 1984a,b, 1988; Crother, in review; Jenner and Dowling, 1985), and Myers and Campbell (1981) discussed the possible relationships of *Coniophanes*. Crother (in review) tried two methods of coding allozyme data and found the position of *Coniophanes* to be unstable. With the locus as the character and polymorphisms coded as uncertain, the genus fell out in a clade containing *Geophis*, *Imantodes*, *Conophis*, *Crisantophis*, and *Enulius* (Fig. 8.13). With the data coded using the presence and absence of alleles (alleles as characters), *Coniophanes* was placed as a basal lineage. Jenner and Dowling (1985) presented a hypothesis that depicted *Coniophanes* as the sister to *Lep-todeira*, but the tree included only eight xenodontine genera and was largely an attempt to fit morphological data with immunological data. Myers and Campbell (1981) proposed in a "theory of relationships" a sister relationship between *Rhadinaea* and *Coniophanes*, and Cadle

(1984b), based on immunological distances, partly supported Myers and Campbell (1981) and suggested that *Coniophanes* was related either to some part of *Rhadinaea* or to *Urotheca* (= *Pliocercus*). In a phylogeny of exemplar taxa, Cadle (1984a, 1988) found *Coniophanes* as either sister to a *Sibon-Geophis* clade or to a *Leptodeira-Eridiphas* clade. Finally, in a study on *Tretanorhinus*, Pinou and Dowling (1994) placed *Coniophanes* with *Sibon*.

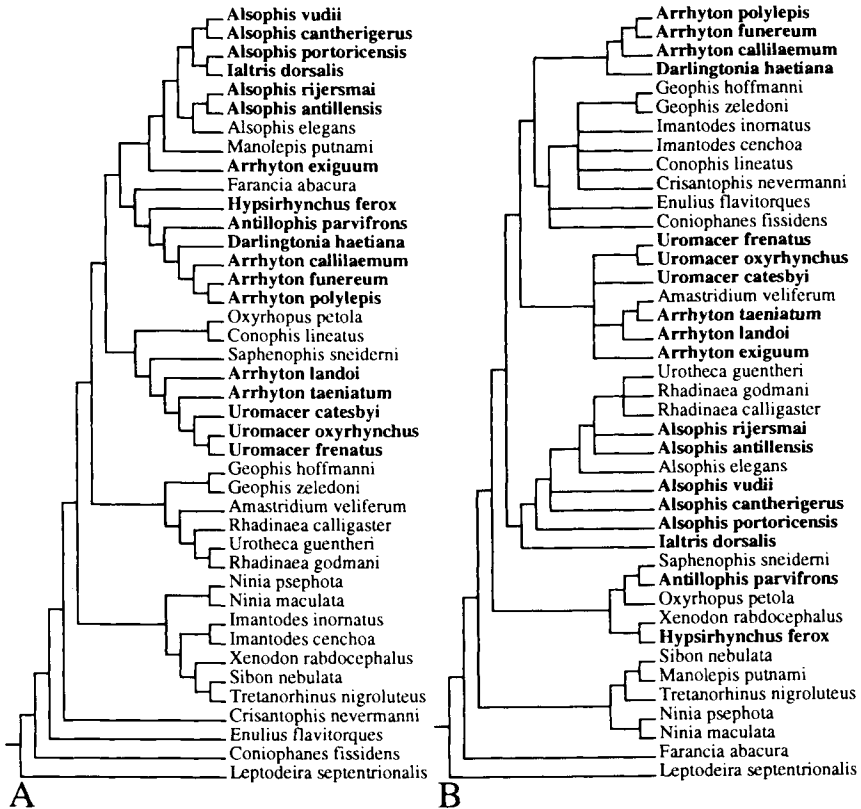


Figure 8.13. Phylogeny estimate of xenodontine snakes based on allozyme data. Bold type denote West Indian species. (A) Strict consensus tree of parsimony analysis with the alleles considered characters. (B) Strict consensus tree of parsimony analysis with uncertain polymorphisms removed (redrawn from Crother, in review).

Tretanorhinus is the only other genus with species on both the islands and the mainland. *Tretanorhinus variabilis* is found on Cuba and the Cayman Islands. Three species are found on the mainland, from southern Mexico into northern South America (Colombia and northwestern Ecuador). There have been no interspecific phylogenetic studies conducted on this genus, and, in fact, the mainland and island forms have not been compared in the same paper to date. Dunn (1939) worked on the mainland forms and Wood (1939) revised the island forms. In neither study did the authors speculate on the overall interspecific relationships of all the species (Dunn did mention that *taeniatus* and *moquardi* were related). Pinou and Dowling (1994) demonstrated that *Tretanorhinus* (using only the Cuban *variabilis*) was not a natricine but clearly related to dipsadines. Their phylogeny shows *Tretanorhinus* relative to *Lep-todeira*, *Sibon*, and *Coniophanes*. Dowling *et al.* (1996), in their attempt to sort out snake phylogeny with allozymes, recovered a clade that contained *Pseustes*, *Leptophis*, and *Tretanorhinus*. The placement of *Tretanorhinus* in this most peculiar clade was by the authors admission, a misplacement. Crother (in review), like Dowling *et al.* (1996), used *nigroluteus* to represent the genus and found it to share a relationship with *Sibon* and with dipsadines in general, a hypothesis similar to that of Pinou and Dowling (1994).

The genus *Liophis* is represented in the Caribbean islands by seven species: *perfuscus* on Barbados, *juliae* on Dominica, Guadeloupe and Marie-Galante, *ornatus* on St. Lucia, *cursor* on Martinique, *melanotus* on Grenada (and Trinidad, Tobago, and northern South America), and *triscalis* on Curaçao [*triscalis* is included in this discussion because of Dixon's (1981) comments]. Maglio (1970), based on morphological data, inferred a phylogeny (Fig. 8.14) of the Caribbean *Liophis* (considered *Dromicus* by Maglio), with the exclusion of *triscalis* because of its distribution in the Dutch Leeward Islands (outside the scope of Maglio's work). The species *melanotus* was placed as the sister to the rest of the endemic Antillean forms. Dixon (1981) took exception with Maglio's concept of relationships among the Caribbean *Liophis*. First, Dixon did not consider *melanotus* closely related to the eastern Caribbean species, but did believe that *triscalis* should be part of the Antillean group. He also concluded that those five Caribbean species (not including *melanotus*) were not closely related to any mainland forms of *Liophis*. That is the current status of our knowledge on the phylogeny of the Caribbean *Liophis*.

The remaining genera of snakes (*Alsophis*, *Antillophis*, *Arrhyton*, *Darlingtonia*, *Hypsirhynchus*, *Ialtris*, and *Uromacer*) found in the West Indies have been variously considered either as multiple lineages (three or four) or as a monophyletic group. As such, they will initially be discussed together with regard to the question of monophyly. Dunn (1932) did not present an explicit phylogeny but argued that this assemblage of snakes was not monophyletic. Maglio (1970) presented the first explicit phylogenetic hypotheses for these taxa and he agreed with Dunn (1932) on one point: the assemblage was not monophyletic. Maglio (1970) depicted three groups: (1) the *cantherigerus* group (*Alsophis*, *Hypsirhynchus*, *Uromacer*), (2) the *andreae* group (*Antillophis*), and (3) the *funereum* group (*Arrhyton*, *Darlingtonia*) (Fig. 8.14). Cadle (1984a) collected immunological data for a subset of taxa from Maglio's *cantherigerus* group and, based on the small amount of differentiation exhibited, suggested that if his sample was typical of the West Indian radiation, then that radiation must be "rather compact" (Cadle, 1984a:16; I interpret this as meaning monophyletic, especially given his 1985 biogeographic scenario). In phylogenies of exemplars, Cadle placed *Alsophis* with *Hydrodynastes* and with *Clelia-Helicops*. Crother (in review), like Maglio (1970), recovered multiple groups of the West Indian taxa (Fig. 8.13). The mainland members of the groups varied depending on the allozyme coding method. As far as the West Indian members were concerned, *Arrhyton exiguum*, *Antillophis parvifrons*, and *Hypsirhynchus* were the only unstable taxa. Otherwise, one group contained *Alsophis* and *Ialtris* (a genus Maglio could not place), a second group contained the Jamaican *Arrhyton* (*funereum*, *callilaemum*, *polylepis*) and *Darlingtonia*, and a third group included *Uromacer* and the Cuban *Arrhyton* (*taeniatum*, *landoi*). Dowling *et al.* (1996) also found the Antillean xenodontines to be nonmonophyletic, with taxa scattered throughout a large xenodontine clade and *Arrhyton funereus* [sic] as part of the unresolved basal lineage between the scolecophidians and the rest of the snakes. It may be safe to assume that based on the phylogenetic evidence compiled to date, the monophyly of these taxa is seriously in doubt, although the relationships among these groups remain in question.

Concordance of interspecific hypotheses of phylogeny are also lacking. In the Antilles the genus *Alsophis* is currently composed of 11 species, several of which are extremely rare or possibly extinct. As with most of the xenodontine genera in the Antilles, Maglio (1970) was the first to estimate phylogenetic relationships for *Alsophis*. Maglio sug-

gested a paraphyletic genus, inserting *Hypsirhynchus* and *Uromacer* as a clade related to *ater* (Fig. 8.14). *Alsophis vudii* and *cantherigerus* formed a clade that was the sister to the previously mentioned *ater* clade. *A. portoricensis* was nested within a clade of LA species and the Hispaniolan taxa were split: *melanichnus* with the LA clade and *anomalus* with the other clade. The proposed sister group to Maglio's paraphyletic *Alsophis* contained *Philodryas* and *Conophis*.

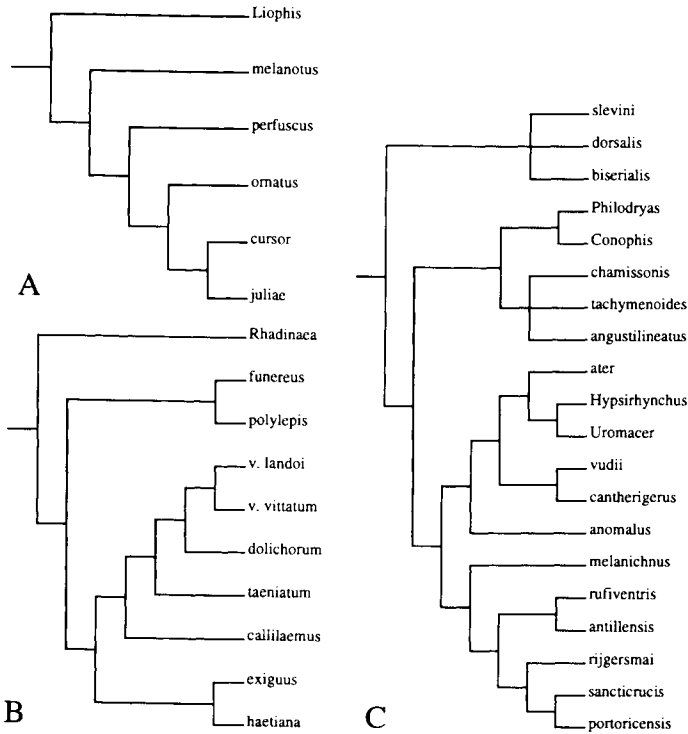


Figure 8.14. Phylogeny estimate of West Indian xenodontines based on morphology. (A) Estimate of *Liophis*. (B) Estimate of *Arrhyton* and *Darlingtonia*. (C) Estimate of *Alsophis*, *Uromacer*, and *Hypsirhynchus*. Specific epithets all refer to *Alsophis* (redrawn from Maglio, 1970).

Cadle (1984a) presented immunological data from *cantherigerus*, *vudii*, *rufiventris*, and *Hypsirhynchus* and concluded that his findings were totally consistent with Maglio's (1970) phylogeny. The species

cantherigerus and *vudii* were essentially indistinguishable (sister taxa?), with *Hypsirhynchus* nearer to them than *rufiventris*, a LA *Alsophis*. The immunological data did not support a *Philodryas* or *Conophis* relationship with the Antillean *Alsophis*. However, Jenner and Dowling (1985) depicted a *Philodryas* - *Alsophis* relationship in their tree. Cadle (1988), in a depiction of some broad colubroid relationships, placed *Alsophis* in a clade with *Clelia* and *Helicops*.

The allozyme data of Crother (in review) also suggested a paraphyletic *Alsophis*, but with respect to different taxa. The preferred hypothesis (note that both hypotheses place *Ialtris* and *A. elegans* with the Antillean *Alsophis*) in Crother (in review) depicts two clades, one composed of the Greater Antillean and Bahamian species (*vudii*, *cantherigerus*, *portoricensis*) and *Ialtris dorsalis* and the other clade composed of the Lesser Antillean species *rijersmai* and *antillensis* plus the South American *Alsophis elegans* (Fig. 8.13). These results support the *vudii*-*cantherigerus* sister relationship (also supported in Crother and Hillis, 1995) and the close relationship among the LA taxa. However, in no cases were *Uromacer* or *Hypsirhynchus* found nested within *Alsophis*. The relationship of *Ialtris* with *Alsophis* was mentioned as a possibility by Maglio (1970) but only because he otherwise could not ascertain the place of *Ialtris*. Schwartz and Rossman (1976) strongly questioned Maglio's comment but held off making a decision until further data could be collected. Based on the allozyme data, Crother (in review) suggested synonymizing *Ialtris* with *Alsophis* to make the latter genus monophyletic. However, the Dowling *et al.* (1996) study indicated a very different picture of *Alsophis* and *Ialtris*. First, the two *Alsophis* representatives (*cantherigerus* and *portoricensis*) were not found to be related and *Ialtris* was even more distant, internested in fact in a clade composed of *Uromacer* and *Hypsirhynchus*, which will be discussed later.

The genus *Antillophis* has two species, *andreae* on Cuba and *parvifrons* on Hispaniola. Because the interspecific relationships represent a trivial problem, the focus will be on the possible sister taxa to the genus. Maglio (1970) suggested that *Antillophis* was derived from a mainland stock, specifically something close to *Saphenophis boursieri*, and that both these groups might be derived from mainland *Alsophis*. Myers (1973) took exception to this hypothesis and did not find evidence to suggest that *boursieri* and *Antillophis* were closely related (certainly not to the point of being placed in the same genus, as mentioned by Maglio). Crother (in review) included *Antillophis parvifrons* and *Saphenophis*

sneideri in his study. Crother's final conclusion was that *Antillophis* could not be reliably placed because of its instability on the trees. However, interestingly, in the locus-as-character-based phylogeny, *Antillophis* fell out as the sister to *Saphenophis*!

The Dowling *et al.* (1996) work also included *A. parvifrons* and in their tree *Antillophis* fell out in a clade containing *Alsophis*, *Arrhyton*, and the southern African lycodontine *Lamprophis* (Fig. 8.12).

The West Indian endemic genus *Arrhyton* currently has 12 species, but Crother (in review) has suggested a change that reduces the number to nine. Because of the apparent similarity and closeness of relationship with *Darlingtonia*, these taxa will be considered together. Maglio (1970) depicted a paraphyletic *Arrhyton* (Fig. 8.14), which included the genus *Darlingtonia*. His phylogenetic concept regarded *funereum* and *polylepis* as a basal sister group; *exiguum* and *Darlingtonia* as sisters, and the other Jamaican form, *callilaemum*, as the sister to a clade composed of the Cuban species. Schwartz and Garrido (1981b) thought the phylogeny was reasonable, but countered it with an alternative. They agreed with the Cuban monophyly (and broke the taxa into three groups), but thought *funereum*, *polylepis*, *callilaemum*, and *exiguum* also belonged in a single group (implied monophyly?). However, Schwartz and Garrido (1981b) and later Hedges and Garrido (1992b) concluded that more data were needed to get a better understanding of *Arrhyton* phylogeny.

Dowling *et al.* (1996) included five species of *Arrhyton* and *Darlingtonia* and their phylogeny supported none of the previous ideas except possibly that *exiguum* perhaps does not belong to *Arrhyton* (Schwartz and Garrido, 1981b). Otherwise, the Cuban forms were not monophyletic (the Jamaican *callilaemum* fell out as the sister to *landoi*), *Darlingtonia* was not related to *Arrhyton*, and *funereum* was not related to other *Arrhyton*. Crother (in review) found *Arrhyton* not to be a cohesive group but rather more along the lines of Schwartz and Garrido (1981b) than Dowling *et al.* (1996). The best supported clade indicates a sister relationship between *Darlingtonia* and the Jamaican *Arrhyton*, with *Darlingtonia* the sister to a clade that shows *funereum* and *polylepis* as sister taxa relative to *callilaemum* (Fig. 8.13). Buden (1966) first suggested the *funereum*-*polylepis* relationship. Crother and Hillis (1995) supported the monophyly of Jamaican *Arrhyton*, but the relationships of *callilaemum* and *funereum* were reversed. The robustness of the *Darlingtonia*-Jamaican *Arrhyton* clade led Crother (in review) to subsume the Jamaican taxa into the genus *Darlingtonia*. The two Cuban taxa in-

cluded in Crother (in review) fell out in the same clade in both trees but never as sister taxa and always with *Uromacer*. The Puerto Rican *exiguum* was unstable (Fig. 8.13) and thus its relationships could not be placed.

Maglio (1970) considered the monotypic *Hypsirhynchus* as the closest relative to *Uromacer* and this clade was nested within the genus *Alsophis* (see discussion on *Alsophis*). As with most of the West Indian snakes, little has been done regarding the phylogenetic position of *Hypsirhynchus ferox*. In Cadie's (1984a) work he somewhat confirmed Maglio's notion that *Hypsirhynchus* belonged within *Alsophis* when he reported that *ferox* was more similar to some species of *Alsophis* than other *Alsophis*. Dowling *et al.* (1996) also supported the *Hypsirhynchus* – *Uromacer* relationship, but Crother (in review) did not. In the Crother study, *Hypsirhynchus* proved to be unstable, from being variously placed in a South American clade paired with *Xenodon* to being in a clade with *Darlingtonia*, *Antillophis*, and *Farancia*.

The final colubrid snake genus to be considered is *Uromacer*. This genus is endemic to Hispaniola, and the three species possess a distinctive treesnake morph. Horn (1969) was the first to speculate on the interspecific relationships of *Uromacer*. My interpretation of his arguments is that *catesbyi* is the sister to a clade of the more derived *frenatus*–*oxyrhynchus*. Maglio (1970) amazingly did not discuss the relationships within *Uromacer* but did emphatically point out the monophyly of the genus. Schwartz (1970) seemed to agree with Horn's (1969) idea of relationship, stating that there was a certain division between the short-snouted forms (*catesbyi*) and the long-snouted forms (*frenatus*, *oxyrhynchus*). Dowling *et al.* (1996), however, suggested a nonmonophyletic *Uromacer*, with one population of *catesbyi* and *frenatus* related to *Hypsirhynchus* and another population of *catesbyi* related to *Ialtris*. Crother (in review) found more traditional relationships within *Uromacer*, one in which ((*frenatus*, *oxyrhynchus*) *catesbyi*) was found, which is congruent with the early ideas, and the other tree simply has *catesbyi* as unresolved but in the same clade as *frenatus*–*oxyrhynchus*.

So where are we with regard to West Indian colubrid snake phylogeny? It is clear that we know little about the relationships of most of these taxa, but at least most taxa have been examined within some sort of a phylogenetic framework. However, most of the approaches are not rigorous discrete character-based parsimony, and no other sophisticated approaches have been applied because largely the data have not gone be-

yond allozymes (with the exception of Crother and Hillis, 1995). Much work is yet to be done before the West Indian colubrid assemblage, especially the xenodontines, is considered resolved.

Elapidae

Two species of coralsnakes are found on islands in the western Caribbean: *Micrurus nigrocinctus* and *M. ruatanus*. *Micrurus nigrocinctus* is found on the Greater Corn Islands and Corn Island, east of Nicaragua (this range of this species also includes all of Mesoamerica and northern Colombia). *Micrurus ruatanus* is endemic to Isla de Roatán, which lies off the northern coast of Honduras. Only a single attempt has been made to estimate the intrageneric phylogenetic relationships of *Micrurus* (Slowinski, 1995). Slowinski used allozymes and morphology in a combined analysis within a parsimony framework and found *M. ruatanus* and *M. nigrocinctus* to be in the same unresolved clade as *diastema*, *distans*, and a sister group composed of *fulvius* and *alleni*. The clade was supported by a single character which was typical of the result. It seems that coralsnake evolution has been conserved, making shared derived characters difficult to find. Slowinski (1995) did express caution about his phylogeny because many species of coralsnakes were not included in his study.

Leptotyphlopidae

There are eight species of *Leptotyphlops* recorded from the Caribbean region. Of these, *goudotii* and *tenella* are clearly mainland forms. *Leptotyphlops goudotii* is Mesoamerican and *tenella* is South American and is known from only a single record in Antigua, which may be in error (Thomas, 1965b). A third taxon, *columbi*, is found only in the Bahamas and is not apparently related to any of the other Caribbean species. The genus is widely distributed in both the Old and New Worlds, but Thomas (1965b) considered the West Indian radiation (at least the Hispaniolan radiation) monophyletic. However, in a later paper, Thomas *et al.* (1985) were uncertain about the monophyletic status and considered the possibility that the island taxa may represent relictual distributions.

There are no phylogenies for the West Indian species. Thomas *et al.* (1985) discussed in detail all the Hispaniolan taxa and included a key to all the Caribbean forms. They presented a narrative phylogeny for the Hispaniolan taxa which suggested a *calypso-asbolepis-leptepilepta* clade that was separate from *pyrites* and *bilineatus*. With the data avail-

able in Tables 1 and 2 of Thomas *et al.* (1985), I performed a parsimony analysis to estimate the phylogeny of the *bilineatus* group. All the characters from their Table 1 were included but only eye size, color, and rows fused in reduction were used from their Table 2 (the others were uninformative or uncodable). The data were analyzed as unordered and an all zero outgroup was included. An exhaustive search using PAUP 3.1 (Swofford, 1993) yielded four most parsimonious trees with a CI = 1.00. The strict consensus tree showed a ((*pyrites*, *bilineatus*)*calypso*, *asbolepis*, *leptepilepta*) set of relationships. The power of this hypothesis is questionable, given the assumption of monophyly and the absence of true outgroup characters.

Tropidopheidae

This family is represented in the West Indies by the genus *Tropidophis*. There are 16 species in this genus; all but three are found in the West Indies. There have been no phylogenies proposed for the species in the genus, although Stull (1928), Schwartz and Marsh (1960), and Schwartz and Garrido (1975) have made comments about possible species groups in their taxonomic revisions of aspects of *Tropidophis*. Bogert (1968) agreed that *Tropidophis* probably comprised a natural group but was critical of the discussions by Stull (1928) and Schwartz and Marsh (1960). In a recent review of West Indian boas, Tolson and Henderson (1993; they included *Tropidophis* although the genus is not in the Boidae), in their chapter on phylogeny, noted the previous revisers but did not substantially build on those earlier ideas. The genus itself has recently received phylogenetic attention by Kluge (1991), Cundall *et al.* (1993), Heise *et al.* (1995), and Dowling *et al.* (1996) in the context of overall snake phylogeny.

Kluge (1991) used morphology and estimated *Tropidophis* as the sister to *Trachyboa* in a clade with other members of the family. The family in turn was the sister to a clade composed of the bolyeroids, *Acrochordus*, and the colubroids. Cundall *et al.* (1993) also utilized morphological data within a parsimony framework and, in their total evidence estimate, found the family to be the sister to the *Acrochordus*-colubroid clade. The difference between the previous two hypotheses in the placement of *Tropidophis* is simply the relative positions of the bolyeroids and tropidophids. The most recent hypothesis (Heise *et al.*, 1995) is based on mtDNA sequence from the 12S and 16S regions. The mtDNA supported, albeit weakly based on the confidence probabilities at the nodes, *Tropi-*

dophis as the sister to a clade of *Loxocemus*–*Python*. These relationships were quite different from the former two hypotheses and other narrative ideas about snake phylogeny. The differences may be due to the methodological approaches. Kluge (1991) and Cundall *et al.* (1993) worked within a cladistic parsimony framework, whereas Heise *et al.* (1995) employed a neighbor-joining approach with the Jukes-Cantor model. An additional reason why the morphology may differ from the mtDNA is because the Heise *et al.* phylogeny is a gene tree which does not necessarily reflect the species tree.

The phylogenetic estimate by Dowling *et al.* (1996) depicted a most unusual set of relationships for *Tropidophis canus* and *haetianus*. First, *Tropidophis* was not monophyletic. Second, *T. canus* was interested in a clade that contained *Atractaspis* and *Heterodon*, among others (Fig. 8.12). Third, *T. haetianus* was placed unresolved between the Scoleco-*phidia* and the rest of the snakes. Again, the allozyme-based higher level snake phylogeny of Dowling *et al.* presents disconcerting sets of relationships.

Typhlopidae

According to Powell *et al.* (1996a) there are 24 species of *Typhlops* in the Antilles. There have been three attempts at understanding the evolutionary relationships of these taxa. Two studies (Hedges, 1989c; Hedges and Thomas, 1991) used allozyme data in a phylogenetic context to search for cryptic species in Jamaica and Puerto Rico, respectively. Unfortunately, the phylogenetic hypotheses were characterless distance-based estimates which obscure any character support that may or may not be present in the data. Hedges (1989c) concluded that no cryptic species existed on Jamaica. Interestingly, however, an examination of the data shows two populations that each have unique alleles present for two loci. These data may actually suggest the presence of two diagnosable populations on Jamaica. A similar problem exists in the Hedges and Thomas (1991) assessment of Puerto Rican *Typhlops*. In this study, they report three distinct clades among the *richardi* populations, two of which they grant specific status. However, an examination of the allozyme data resulted in zero diagnosable characters for either clade. This would have been obvious in a character-based analysis [which they say they did (Hedges and Thomas, 1991:449) but I could not find]. My own phylogenetic reanalysis of these data did not support either *platycephalus* or *hypomethes*. Dowling *et al.* (1996) found *richardi*, *platycephalus*, and *hy-*

pomethes to form a trichotomy, which was sister to *jamaicensis*. Clearly, the validity of these taxa should be questioned. Thomas (1989) provided a “cladogram of relationships” for all the Antillean *Typhlops* (Fig. 8.15). His hypothesis was based on morphological data and apparently constructed by hand (quite a feat considering there are well over 10^{21} possible trees). As a heuristic exercise, I (and Oscar Flores-Villela) organized Thomas’ data into a matrix of 22 characters and estimated the phylogeny using PAUP 3.1 (Swofford, 1993; with the standard options of a heuristic search). We examined only the large clade and used *caecatus* as the out-group.

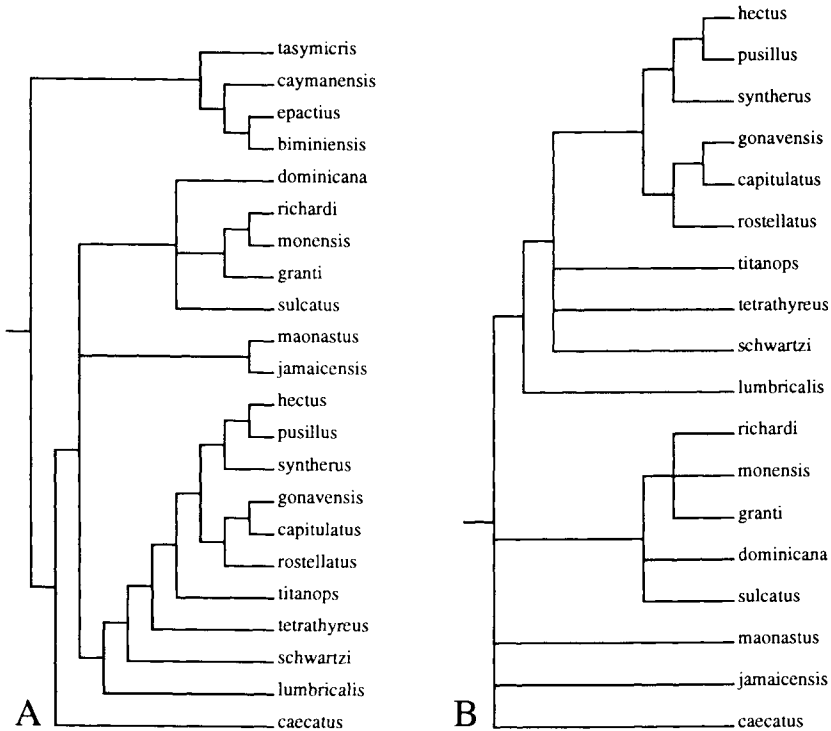


Figure 8.15. Phylogeny estimates of West Indian *Typhlops*. (A) Hand-calculated, morphology based hypothesis (redrawn from Thomas, 1989). (B) Parsimony reanalysis of the same data. See text for analysis details.

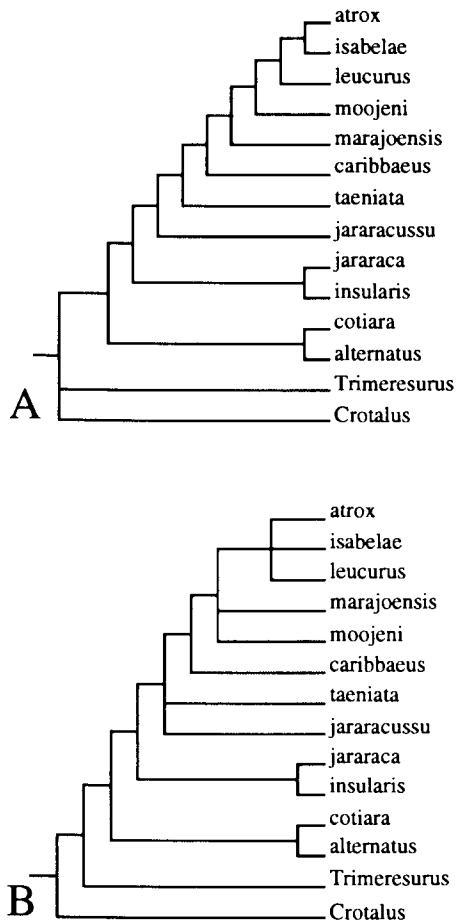


Figure 8.16. Phylogeny estimates of *Bothrops* and *Bothriopsis* based on mitochondrial DNA cytochrome b sequence. (A) 75% majority-rule tree from 11 most parsimonious trees discovered in a parsimony analysis. (B) Semistrict consensus tree of the parsimony consensus tree, a maximum likelihood tree, and two Fitch-Margoliash trees (one with and one without the molecular clock assumption) (redrawn from Salomao *et al.*, 1997).

The analysis resulted in a single most parsimonious tree (24 steps; CI = 0.826; RI = 0.947) that was mostly congruent with Thomas' hypothesis

(Fig. 8.15). The only difference is that the PAUP tree is less resolved (probably because we could not translate all of Thomas' characters into a matrix) and *monastus* and *jamaicensis* are not sister taxa but are placed basally with unresolved relationships. It can be concluded that Thomas' data are fairly robust, but again, more data are needed to solidify Thomas' conclusions.

Viperidae

Two species of *Bothrops*, *caribbaea* and *lanceolatus*, are currently recognized and are restricted to St. Lucia and Martinique, respectively. Historically, these two taxa have been considered as either *B. atrox* or related to *B. atrox* and the recent work supports these concepts. Amaral (1925, 1954) considered *lanceolatus* as an island form of *B. atrox* and sunk the species twice, the second action after Hoge (1952) reelevated the name. Lazell (1964) noted the difficulties in diagnosing the species from the *atrox* complex and admitted that no unique characters existed for either species. Regardless, he found various combinations of characters that allowed for his reelevating the names. Gosner (1987) reexamined a large series of both Antillean species with mainland *atrox/asper* and also concluded that *caribbaea* and *lanceolatus* were probably just *atrox/asper* variants based on the finding that none of the color patterns of the Antillean forms were unique. S. Werman (personal communication, unpublished data) found the island forms to be virtually identical with the *atrox* complex.

The only phylogenies for these taxa are based on 565 (Salomao *et al.*, 1997) and 520 nucleotides (Wüster *et al.*, 1997) of cytochrome *b*. Both studies found *caribbaea* [*lanceolatus* was not included, although Wüster (personal communication) said some DNA sequence was available and that based on that evidence the position of *lanceolatus* remained enigmatic] the sister taxon to the *atrox* complex, but the authors noted that no conclusions could be made about the reality of the species in the *atrox* complex (Fig. 8.16). Curiously, the implication was that *caribbaea* is real. Unfortunately, no characters were shown diagnosing any lineages on the trees, so evaluation of the clades was impossible. In addition, there is no way to ascertain whether or not the sequences used in the two studies are mutually exclusive or from the same region.

The nucleotide data support the notion that the island forms are at least closely related to, if not actually part of, the *atrox* complex. However, clearly a more complete study needs to be undertaken.

Testudines

There are four extant genera of turtles (excluding sea turtles) in the West Indies, and of these only one has endemic species. The taxa *Geochelone carbonaria* and *Pelusios subniger* are considered to be introductions (Pritchard and Trebbau, 1984; Censky, 1988), and *Kinosternon scorpiodes* is found only in the extreme western Caribbean off the shore of Nicaragua where its conspecifics are found throughout Central America into Texas and possibly into South America. Thus, only the relationships of the fourth genus, *Trachemys*, will be considered.

As currently recognized (the history of the contents of this genus has been somewhat tortured, Seidel, 1988a), four species (*terrapen*, *decorata*, *decussata*, *stejnegeri*) are contained in *Trachemys* and the phylogeny of this group appears for the most part to be well supported. Biochemical data (Seidel and Atkins, 1987) and morphological data (Seidel, 1988a) were analyzed as a combined data set within a cladistic framework. The resultant hypothesis (Fig. 8.17) suggested a paraphyletic West Indian *Trachemys* with respect to mainland Neotropical *T. scripta* (Seidel, 1988a).

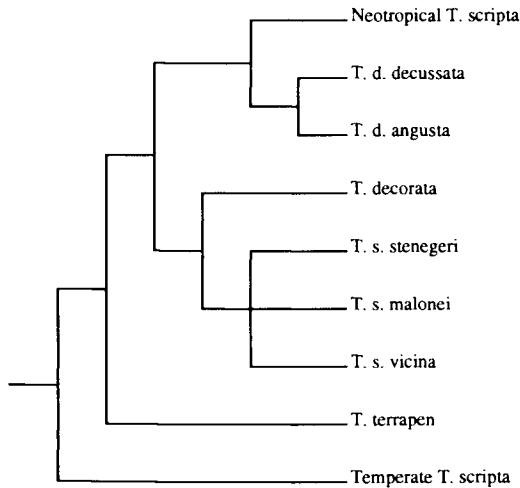


Figure 8.17. Phylogeny estimate of *Trachemys* based on a cladistic analysis of morphology and allozymes (redrawn from Seidel, 1996).

With *Pseudemys* as the root, mainland *T. scripta* fell out as the sister to the W.I. *Trachemys* with the other *scripta*: ((((*scripta*, *decussata*)(*stejnegeri*, *decorata*))*terrapen*)*scripta*). The obvious problem is the status of the taxon “*scripta*” because apparently the label is hiding additional diversity.

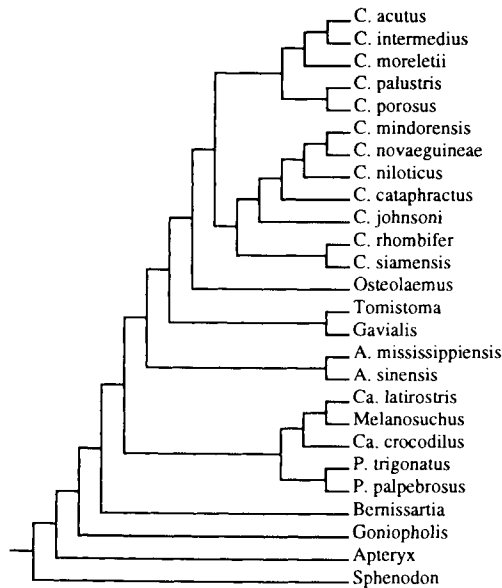


Figure 8.18. Phylogeny estimate of crocodylians based on a parsimony analysis of combined data from rDNA RFLP, mtDNA RFLP, 12S sequence, and morphology. The estimate is the single most parsimonious tree (redrawn from Poe, 1996).

Crocodylia

Four crocodylians are found in the West Indies: *Caiman crocodilus*, *Crocodylus acutus*, *C. intermedius*, and *C. rhombifer*. Of these, one is introduced (Puerto Rico; *Caiman*), two have been recorded as rare vagrants in the Grenadines (*Caiman* and *Crocodylus intermedius*), one is widespread throughout the Greater Antilles and mainland shores of Central America, southeastern North America and northern South America (*C. acutus*), and the fourth is endemic to Cuba (*C. rhombifer*). Gill (1968), in a zoogeographic work, suggested that *rhombifer* and *moreletii* were "certainly closely related" and that any apparent relationship between *acutus* and *intermedius* was only superficial and historical.

Karyotypes were employed by Cohen and Gans (1970) and in a quite convoluted scenario their data indicated an *acutus-johnsoni* group which possibly gave rise to *rhombifer* which was grouped by itself. Also, *rhombifer* may have been derived from *palustris* or *palustris* may have been derived from *rhombifer*. According to their schema, the *acutus* group may have been derived from either *porosus* or a group containing *intermedius*, *niloticus*, and *novaeguineae*. The schema presented by Cohen and Gans was based on certain assumptions about chromosomal change and thus represents what they called the "probable path of change."

Ecological data in the form of nesting habits were brought to bear on the phylogeny of crocodylians by Greer (1970). His phylogeny proposed a large clade which included *acutus*, *intermedius*, *rhombifer*, *niloticus*, *palustris*, *johnsoni*, and *siamensis*. His subdivision of the clade showed an *acutus-intermedius-rhombifer* group. Later, Campbell (1972) suggested that nesting habits do not reflect phylogenetic history but instead reflect ecological similarities.

Brooks (1981) conducted a cladistic analysis on morphological data of crocodile species-specific digenean parasites and built a crocodile phylogeny based on the results. In this classic work utilizing the nonrandom association of hosts and parasites he found an *acutus-rhombifer* clade sister to a *moreletii-intermedius* clade. This study was expanded (Brooks and O'Grady, 1989) to include data from nematode parasites and biogeographic information. In every outcome, the *acutus-rhombifer* sister relationship was supported but the sister relationship to that clade was unresolved.

Densmore (1983) used albumin immunodiffusion data clustered by UPGMA and found *acutus* sister to a clade containing *palustris*, *porosus*,

cataphractus, and *johnsoni*. *Crocodylus rhombifer* was not included in the study. In the same paper, Densmore presented a hypothesis based on the allozyme presence/absence of alleles data and represented by an unrooted Wagner tree. In this case he found *acutus* and *rhombifer* in a clade with *siamensis* and *porosus*. Densmore and White (1991) followed up these data with restriction length fragment data from whole mitochondrial digests and nuclear rDNA digests and analyzed them using phenetic (UPGMA) and compatibility or clique analyses. The compatibility tree of the rDNA data exhibited an *acutus-intermedius* clade and a *moreletii-rhombifer* clade. The relationships of these two clades were unresolved because they were part of a seven branch polytomy. Two different compatibility analyses of the mtDNA data yielded two hypotheses presented as an unrooted networks. One hypothesis indicates an *acutus-moreletii* clade and includes *rhombifer* in a large group composed of *niloticus*, *no-vaeguineae*, *mindorensis*, and *palustris*. The other compatibility network suggested that *acutus* was unique, perhaps sister to *moreletii*, and that *rhombifer* was again part of a larger group but with a different membership: *palustris*, *niloticus*, and *siamensis*. Additionally, Densmore and White combined the data into a phenetic analysis using UPGMA as the clustering algorithm. This result suggested an *acutus-moreletii* clade with *rhombifer* the sister taxon and *intermedius* the sister to the clade composed of *acutus-moreletii-rhombifer*.

Poe (1996) presented a careful combined (total-evidence) analysis of all the previous character sets. He recovered a single most parsimonious tree (Fig. 8.18) from all the discrete characters. *Crocodylus acutus* was found to be the sister of *intermedius*, and *rhombifer* fell out as the sister to *C. siamensis*, but both of these clades had decay indices of only 1. Whereas the *C. acutus* - *C. intermedius* clade is reasonable, the *rhombifer* - *siamensis* relationship stretches the biogeographic imagination. Regardless, I consider the hypothesis of Poe to be the best estimate of crocodylian phylogeny.

Conclusions

A review of phylogeny is, in a way, simple relative to a review of ecology, for example. This is because the boundaries of the goal are defined: find all the figured phylogenies. Although I set out to do just that and have accumulated a large number of phylogenetic hypotheses for the West Indian herpetofauna, I undoubtedly missed some. Even if we as-

sume this as true, the missing phylogenies would not alter my conclusions (nor alter the conclusions of the reader). The main point of summation derived from the review is that only a minuscule fraction of the West Indian herpetofauna is well-known phylogenetically. Few relationships have been estimated with multiple data sets and rigorous phylogenetic methodology, with the result being stability among those relationships (= highly corroborated hypotheses). For example, the taxon *Cricosaura*, although the subject of multiple data sets and phylogenetic analyses, has been unstable in its phylogenetic position and so is not considered to be "known" phylogenetically. I would argue that perhaps *Epicrates*, crocodylians, and *Trachemys* (even with the sticky *scripta* problem) could be considered to have well corroborated phylogenies.

Unfortunately, this pessimistic view of our state of the knowledge of West Indian herpetofaunal phylogeny is not helpful to the nonsystematist who needs a phylogeny to conduct a comparative study. One suggestion would be to use each of the available phylogenetic hypotheses which would lead to multiple testable hypotheses from the comparative study. It is perhaps more work, but certainly would be enlightening and might in fact lead to an empirical reason to choose one of the competing phylogenetic hypotheses over the others (such a study could be reasonably pared down if only philosophically justified approaches to phylogenetic inference are examined). The argument is not tautological but is analogous to using phylogenetic trees to choose among competing (or present novel) geological hypotheses. Another approach, which I think is the more justifiable of the two, would be to include the ecological, behavioral, etc., data with the original data used to estimate the cladogram and then re-analyze the data together. If care is taken in coding the ecological characters in question, concerns of tautology, non-independence, and bias become moot (Luckow and Bruneau, 1997).

The optimistic view is that phylogenetic herpetology in the West Indies is wide open (although I suspect there are those who would argue otherwise) and waiting to be done. To this end, a number of labs have taken up the challenge and are currently working on West Indian problems. The assessment of accuracy of phylogenies is a difficult proposition. I would argue impossible because the *true* phylogeny is unknowable. Instead, we are left with hypotheses that are resilient to falsification. Multiple data sets are a necessity to establishing highly corroborated phylogenies. A priori, no single data set is superior to another (although for methods this is a different matter), and as such both molecular

and morphological based phylogenies need to be estimated. Finally, these multiple data sets should be combined to give the overall best estimate of character covariation (congruence) and thus phylogenetic relationships.

Appendix.

Studies Which Include Figured Estimates of Phylogeny for West Indian Species.

Species	Reference
Anura	
Leptodactylidae	
<i>Eleutherodactylus</i>	Hass and Hedges 1991 Hedges 1989 a,b, 1991 Joglar 1983, 1989 Kaiser <i>et al.</i> 1994 a,b,c, 1996 Lynch 1996 Smith <i>et al.</i> 1981 Heyer 1970
<i>Leptodactylus</i>	
Sauria	
Gekkonidae	
<i>Aristelliger</i>	Russell 1979
<i>Gonatodes</i>	Kluge 1995
<i>Hemidactylus</i>	Kluge 1969 Russell 1979
<i>Phyllodactylus</i>	Russell 1979
<i>Sphaerodactylus</i>	Schwartz and Thomas 1983 Hass 1991, 1996 Kluge 1995 Page and Lydeard 1994
Iguanidae	
<i>Cyclura</i> and <i>Iguana</i>	de Queiroz 1987 Etheridge and de Queiroz 1988 Frost and Etheridge 1989 Hollingsworth 1998 Norell and de Queiroz 1991 Sites <i>et al.</i> 1996

-
- Polychrotidae
 Anolines
- Burnell and Hedges 1990
 Cannatella and de Queiroz 1989
 Case and Williams 1987
 Etheridge 1960
 Frost and Etheridge 1989
 Gorman and Atkins 1969
 Gorman and Kim 1976
 Gorman *et al.* 1980, 1983
 Guyer and Savage 1986, 1992
 Hass *et al.* 1993
 Hedges and Burnell 1990
 Lazell 1966, 1972
 Paull *et al.* 1975
 Peterson 1983
 Poe 1998
 Roughgarden 1995
 Shochat and Dessauer 1981
 Williams 1972, 1977
 Wyles and Gorman 1980
 Yang *et al.* 1974
- Teiidae
Ameiva
Bachia
- Tropiduridae
Leiocephalus
- Xantusiidae
Cricosaura
- Serpentes
 Boidae
Boa
- Corallus*
- Barbour and Noble 1915
 Dixon 1973
 Frost and Etheridge 1989
 Pregill 1992
 Crother *et al.* 1986
 Crother and Presch 1993
 Hedges *et al.* 1991
 Peterson and Bezy 1985
 Schatzinger 1980
 Cadle 1988
 Heise *et al.* 1995
 Underwood 1976
 Henderson and Hedges 1995
 Kluge 1991
 Underwood 1976

<i>Epicrates</i>	Kluge 1989, 1991 Tolson 1987 Underwood 1976
Colubridae	
<i>Alsophis</i>	Cadle 1984, 1988 Crother in review Crother and Hillis 1995 Dowling <i>et al.</i> 1996 Jenner and Dowling 1985 Maglio 1970
<i>Antillophis</i>	Crother in review Crother and Hillis 1995 Dowling <i>et al.</i> 1996 Maglio 1970
<i>Arrhyton</i>	Crother in review Crother and Hillis 1995 Dowling <i>et al.</i> 1996 Maglio 1970
<i>Chironius</i>	Dowling <i>et al.</i> 1996
<i>Clelia</i>	Cadle 1984, 1988 Dessauer <i>et al.</i> 1987 Dowling <i>et al.</i> 1996 Jenner and Dowling 1985
<i>Coniophanes</i>	Cadle 1984, 1988 Crother in review Jenner and Dowling 1985 Myers and Campbell 1981 Pinou and Dowling 1994
<i>Darlingtonia</i>	Crother in review Dowling <i>et al.</i> 1996 Maglio 1970
<i>Hypsirhynchus</i>	Cadle 1984 Crother in review Crother and Hillis 1995 Dowling <i>et al.</i> 1996 Maglio 1970
<i>Ialtris</i>	Crother in review Dowling <i>et al.</i> 1996
<i>Liophis</i>	Dowling <i>et al.</i> 1996 Maglio 1970
<i>Mastigodryas</i>	Dowling <i>et al.</i> 1996

<i>Nerodia</i>	Lawson 1987
<i>Pseudoboa</i>	Cadle 1984 Dessauer <i>et al.</i> 1987 Dowling <i>et al.</i> 1996 Jenner and Dowling 1985
<i>Tretanorhinus</i>	Crother in review Dowling <i>et al.</i> 1996 Pinou and Dowling 1994
<i>Uromacer</i>	Crother in review Crother and Hillis 1995 Dowling <i>et al.</i> 1996 Maglio 1970
Elapidae	
<i>Micrurus</i>	Slowinski 1995
Tropidopheididae	
<i>Tropidophis</i>	Cundall <i>et al.</i> 1993 Dowling <i>et al.</i> 1996 Heise <i>et al.</i> 1995 Kluge 1991
Typhlopidae	
<i>Typhlops</i>	Dowling <i>et al.</i> 1996 Hedges 1989 Hedges and Thomas 1991 Thomas 1989
Viperidae	
<i>Bothrops</i>	Salamao <i>et al.</i> 1997
Testudinata	
Emydidae	
<i>Trachemys</i>	Seidel 1987, 1988, 1996
Crocodylia	
Crocodylidae	
<i>Crocodylus</i>	Brooks 1981 Brooks and O'Grady 1989 Cohen and Gans 1970 Densmore 1983 Densmore and White 1991 Greer 1970 Poe 1996

Ecological and Historical Biogeography of the Caribbean

*Gregory K. Pregill*¹ and *Brian I. Crother*²

¹*Department of Biology, The University of San Diego, 5998
Alcalá Park, San Diego, California 92110-2492*

²*Department of Biology, Southeastern Louisiana University, Hammond, Louisiana
70402*

Introduction

The modern era of biogeography began about 30 years ago consequent to four unrelated but more or less synchronous events: the formalization of plate tectonics and a mobile geology of the earth's crust; advances in radiometric dating techniques; advances in phylogeny reconstruction; and publication of MacArthur and Wilson's (1967) Theory of Island Biogeography. At this point, biogeography went in two directions. One was ecological. Insularity became the focus of study on community evolution and the processes of species interaction. Ecological biogeography thus probes such issues as equilibrium dynamics, taxon cycles, and competition in their capacity to effect species assortment.

The other direction in biogeography was the traditional search for the geographic origin of clades. Historical biogeography depends on phylogeny, levels of endemism, fluctuations in sea level, and past geomorphologies such as connections between a continent and an island, or one island with another. The two approaches to biogeography largely operate independently of one another, although they both attempt to explain the same thing: the geographic distribution of life. Historical biogeography finds no advantage in field experiments and models that strive to reveal ecological relationships. This is because the results obtained usually apply to a narrow set of circumstances from an instant in geological time. Nonetheless, the results of field manipulations and their models accumulate, and the general patterns that emerge from them

make ecological biogeography a successful enterprise. However, ecological biogeography lacks a sturdy historical framework, and often it is difficult to identify which characteristics of a population are opportunistic responses to the environment of the time, and which are the products of history. Historical and ecological biogeography are interwoven, but especially for vertebrates this relationship is not well understood.

The Caribbean biota has been a spectacular inspiration to both approaches. In this chapter we review the herpetological components of each. Of the two, the historical approach is pursued by most herpetologists who study the West Indies. However, the amphibians and reptiles themselves, particularly lizards, are favored subjects of ecologists. Although we make no attempt to construct a unifying set of historical and ecological objectives, we do elaborate on why events in the Caribbean during the past 10,000 years are significant to both.

Historical Biogeography

Thoughts on the origin of the West Indian biota can easily be categorized into two periods: the previcariance era and the vicariance era. The temporal and philosophical boundary between these two periods is a fairly clean paradigm shift in general explanations about distribution. Although herpetologists were mostly minor players in the previcariance era, they took on substantially larger roles afterwards. Williams' (1989b) thorough review of the ideas and participants in Caribbean biogeography consolidated the history for subsequent discussions (e.g., Crother and Guyer, 1996). It is not our intention to cover all of the same ground here; however, some overlap with Williams is inescapable.

Previcariance Era

The central debate during the pre-vicariance era was whether land vertebrates dispersed to the islands over water, or directly by way of land bridges. Although both were dispersalist arguments, they differed in that overwater processes were, of necessity, random or chance events; land bridges implied concordant movement. The debate was constrained by the presumed stability of continents, which demanded the active movement (dispersal) of organisms across the landscape. Additionally, there was a prevalent assumption, steeped in religious views of creation, that taxa arose from centers of origin, further requiring dispersal explanations to account for distributions.

The entrenched foundation of the pre-vicariance attitude left little room for options, and for their part few herpetologists considered possibilities other than overwater dispersal. Indeed, if the continents were fixed then there was no acceptable alternative. However, the land bridge hypothesis had one of its greatest champions in the Harvard herpetologist Thomas Barbour (e.g., 1914, 1915). Barbour argued against the prevalent theory of flotsam and jetsam dispersal as espoused by Matthew (1915) and others. For example, Barbour and Noble (1915:418) noted,

This revision, which is a detailed study of a single genus of strictly terrestrial teid (sic) lizards, shows clearly the close relationship and origin from a common stock of many of the Antillean forms. The data derived from this study seem to argue strongly against the flotsam and jetsam theory.

Approximately 20 years later, Barbour (1937) continued advocating non-overwater dispersal, although in words echoed four decades later by the vicariance proponent Gareth Nelson, he allowed that some random dispersal must have occurred: Barbour (1937:79) stated, "There has been undoubtedly some dispersal by flotsam and jetsam" and Nelson (1974:556) noted, "If so, many episodes of dispersal - which no doubt have occurred." But Barbour (1937:80) went on to say,

However, to conclude that all of the animals of an island, such as any of the Greater Antilles, have been derived by any or all of these causes [various random events] is to support an explanation which is to my way of thinking infinitely less probable than to postulate extensive changes in land form in a region where so much tectonic (sic) movement is evident on every hand. That the separation of the Greater Antilles took place a long time ago is certain, for *Cadea* was not derived from *Amphisbaena* nor *Cricocaura* (sic) differentiated from its Xantusiid forebears except in a very long time.

The confidence Barbour displayed in his 1937 paper was partly attained from Schuchert's (1935) *Historical Geology of the Antillean-Caribbean Region* published two years earlier. Schuchert described the origin of the Greater Antilles in a manner not unlike modern tectonic models, and he concluded that land bridges were the best explanation to account for the West Indian biota.

Barbour's position on the origin of the West Indian biota is remarkable for three reasons: (1) until the vicariance revolution he essentially stood alone against random, overwater dispersal; (2) his reasoning presaged vicariance but without mechanism or method; and (3) he argued mostly from his knowledge of reptile and amphibian distributions and

systematics (at the time most Caribbean zoogeography derived from the study of mammals). As strong as Barbour's arguments were, however, he had little support for them and could offer no geological mechanism to make land bridges credible. The overwater dispersal school was dogma, as expressed, for example, by Darlington (1938:297),

Both of the two independent lines of valid evidence indicate, then, that the fauna of the Greater Antilles has been derived from the mainland across water...so far as I can see, no other hypothesis will fit the facts.

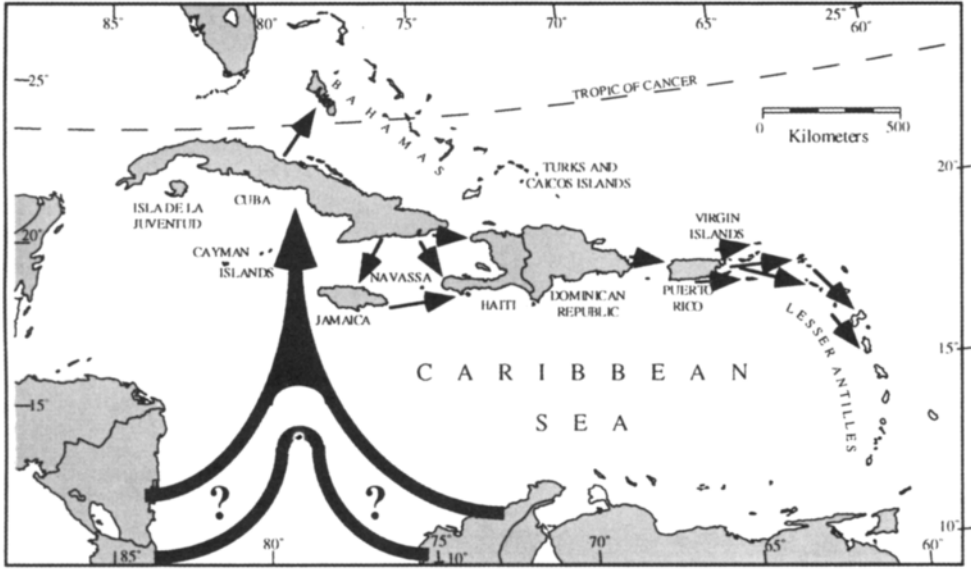
and finished by Simpson (1956:7):

When I first drafted the present paper it seemed useless to reopen a question that had been so thoroughly discussed and had, I felt, been settled.

In the late previcariance era, the tempest over the Antillean biota as a whole spilled onto discussions about the distribution of species among the islands themselves. Various overwater dispersal scenarios proposed complex retrodictions of movement, with taxa making their way from one island to another and sometimes back again. Two examples from herpetology are Gorman and Atkins (1969) and Maglio (1970) (Fig. 9.1). Many of these scenarios were creative efforts at drawing lines and arrows, but their plausibility was difficult if not impossible to test. Ultimately, this problem of testability led to general dissatisfaction with random dispersal as an a priori explanation for Antillean land vertebrates.

Vicariance Era

The shift to a vicariance rationale in biogeography occurred fairly rapidly. In the 1970's, confluence of the independent but essentially simultaneous revolutions in geology and systematics were applied to the then obscure ideas of Leon Croizat's (1958, 1964) panbiogeography (Croizat *et al.*, 1974). Mechanisms of seafloor spreading and plate tectonics replaced the stable geology of the earth's crust with drifting continents and the opening of ocean basins (e.g., Isacks *et al.*, 1968). The first tectonic model for the Caribbean plate soon followed (Malfait and Dinkelman, 1972). At about the same time, the English translation of Willi Hennig's *Phylogenetic Systematics* in 1966 proclaimed a fresh methodology for phylogenetic reconstruction. Its halting but ultimate embrace by systematists is chronicled in a lively book by Hull (1988).



A



B

Figure 9.1. Example dispersal scenarios for West Indian reptiles. (A) Example for xenodontine snakes (redrawn from Maglio, 1970). (B) Example for anoline lizards (redrawn from Gorman and Atkins, 1969).

Croizat devised a simple scheme for plotting the range of species that allowed him to visualize the concordant distribution of diverse taxa. He argued that these "tracks", as he called them, must result from processes that affected entire biotas. Distributions outside the generalized tracks could be explained by specific instances of dispersal.

The vicariance school of historical biogeography began to materialize when Gareth Nelson (1969, 1973, 1974) brought attention to Croizat's ideas. Nelson combined new knowledge on geographic histories based on tectonics, with Hennigian phylogenetics (cladistics). The result was vicariance biogeography. The premise had elegant simplicity: if areas had historical relationships, and if taxa had historical relationships, then when area and taxonomic relationships were congruent the history of the areas explained the distributional history of the taxa. Thus, geographic history is the independent variable and biology the dependent variable. The method's power is its testability, so long as both geographic history and phylogeny are recoverable.

Among the first to apply vicariance methods to real problems was Donn Rosen (1976, 1978, 1985), who in doing so substantially altered the forum for Antillean biogeography. Rosen (1976) argued, based on then current tectonic models (mostly Malfait and Dinkelman, 1972), that a proto-Antilles archipelago was situated between North and South America (SA) from the late Cretaceous to the early Paleocene. This archipelago derived its fauna from the continents to the north and south. As the proto-Antilles moved eastward relative to North and South America in the early Cenozoic, it carried the antecedents of the Antillean biota with it.

Following the publication of Rosen's model, interest in Caribbean biogeography was renewed. Perhaps because amphibians and reptiles are the dominant terrestrial vertebrates (diversity and numbers) in the Antilles, herpetologists have been active contributors. Pregill (1981b) argued that the geophysical evidence for a proto-Antilles was equivocal, and that the fossil record (continental) suggested that many genera of West Indian vertebrates appeared subsequent to the formation of the Greater Antilles. Hedges (1982) rebutted Pregill's (1981b) contention that geologic evidence was insufficient, but did not comment on the fossil record or the composition of the modern fauna, which continued to be points of disagreement (e.g., Williams, 1989b; Crother and Guyer, 1996).

In 1982 Jay Savage published his "Enigma" paper. Savage emphasized the uniqueness of the Mesamerican (MA) herpetofauna and its

relevance to historical biogeographic questions in the region. He modified Rosen's (1976) original model and elaborated on the complex, inter-related histories of the Caribbean and Mesoamerica. He also concluded that there must have been a late Cretaceous-early Paleocene land connection between MA and SA. Savage (1966, 1982) is not alone in arguing for a distinctive MA biota. Cadle (1985), Darwin and Welden (1992), and Crother *et al.* (1992) provided additional support. The reality of a MA-SA connection, although implied in some form by most tectonic models, remains unproven geologically (Crother and Guyer, 1996; Perfit and Williams, 1989). Biologically, however, a MS-SA connection is practically demanded.

Williams (1989b) addressed Savage's (1982) paper in detail. In particular, he disagreed that there was a late Cretaceous-early Paleocene connection between MA and SA, and he also maintained that the Tertiary fossil record was important despite its absence in the region. In tropical continental latitudes, and on most oceanic islands, the vertebrate fossil record is largely silent about events of the Cenozoic before the Quaternary. Tropical soils and climate are not conducive to the long-term preservation of bone, and all but the largest islands lack sufficient deposits in which bones of terrestrial vertebrates have their best chance of being preserved over millions of years. Historical biogeography of the Caribbean is thus disadvantaged by the lack of hard evidence from early periods of the Cenozoic. At issue is whether this gap can be filled in reasonably with assumptions. Minimizing assumptions on the one hand means that negative evidence (the lack of appropriate fossils), however lamentable, is irrelevant to the analysis. On the other hand, a comparatively rich fossil record from temperate North and South America might suggest minimum ages of circum-Caribbean sister taxa. For instance, if passerine birds are unknown before the Miocene and colubrid snakes before the Oligocene, then their appearance in the Caribbean would have postdated the rafting of the proto-Antilles (or whatever the geographic configuration of the islands was in the Paleocene). Still, the reality of such a history can never be demonstrated by corroborating evidence; it is speculation that can only be falsified, in this case by the discovery of older fossils.

In the past 15 or so years numerous herpetofaunal studies have addressed Caribbean biogeography, but there has been no solid consensus to emerge from the effort (e.g., Burnell and Hedges, 1990; Cadle, 1985; Crother, 1989; Crother *et al.*, 1986; Guyer and Savage, 1986; Hass,

1991; Hass and Hedges, 1991; Hass *et al.*, 1993; Hedges, 1989a; Hedges and Burnell, 1990; Hedges *et al.*, 1991; Jenner, 1981; Joglar, 1989; Kluge, 1988, 1995; Page and Lydeard, 1994; Pregill, 1981a, 1992; Roughgarden, 1995; Savage and Lips, 1993; Schwartz and Garrido, 1985; Seidel, 1988a, 1996; Sites *et al.*, 1996; Thomas, 1989; Tolson, 1987; Williams, 1989a). The most recent variables of the vicariance debate were added by Hedges *et al.* (1992b; also in Hedges 1996a). They reported pairwise comparisons of immunological distance (ID) data for selected West Indian and mainland taxa. Based on a molecular clock of those data, they concluded that the taxon divergences were too recent to have been associated with a late Cretaceous vicariance event. Accordingly, they argued for overwater dispersal of the herpetofauna. In rebuttal, Crother and Guyer (1996) combined much of the available data (including nonherpetofaunal phylogenies) into a single parsimony analysis and obtained a consistent result that supported a vicariance hypothesis. But there was more at stake. Hedges *et al.* (1992b) suggested that giant tsunamis – generated by a late Cretaceous bolide impact on the north coast of the Yucatan – scoured the Antilles of all but the most tenacious weeds and arthropods. Hence, land vertebrates must have recolonized later in the Cenozoic.

Page and Lydeard (1994), Crother and Guyer (1996), and Guyer and Crother (1996) argued against the molecular clock/bolide hypothesis [see Hedges *et al.* (1994) and Hedges (1996c,e) for counter arguments]. Page and Lydeard (1994) restated the importance of phylogenetic analysis in recovering biogeographic history. Crother and Guyer (1996) revealed problems with the tsunami hypothesis, the molecular clock calibrations, and the pairwise comparisons. Other studies (e.g., Keller *et al.*, 1997; Stinnesbeck *et al.*, 1997) found little evidence of impact-generated, short-term (over hours or days) tsunami deposits. Even if an impact did occur, physical models dispute the possibility of waves reaching the dimensions (4 km tall) required to inundate the entire West Indies (Gault and Sonett, 1982; see Crother and Guyer, 1996).

The molecular clock is a tantalizing and potentially powerful tool. Unfortunately, it is beset with problems, and is sloppy with respect to calibration. Consider the demonstration by Hillis *et al.* (1996), who idealized a molecular clock with 95% confidence limits and only stochastic variation as a source of error. At an arbitrarily assigned rate of one substitution per 1 million years (my), taxa separated by 15 my could have between 8 and 22 substitutions. However, an age of 5 my could also have

8 substitutions, such that neither age (5 or 15 my) can be rejected. Hillis *et al.* (1996:538) also examined ID data (like that used by Hedges *et al.*, 1992) from birds, and concluded

“The calibration of albumin divergence based on immunological comparisons among birds shows that the confidence limits of new predicted values of time may be so large as to not exclude any reasonable possibility” (emphasis ours).

They illustrated their point (Hillis *et al.*, 1996:538, fig. 14). For an estimated divergence time of approximately 80 my, the ID values within the 95% confidence intervals range from 20 to 90. For a divergence time of 20 my, the range of ID values within the 95% confidence intervals is 0 - 45. Thus, for ID values between 20 and 45, neither 20 nor 80 my ages can be rejected at 95% confidence.

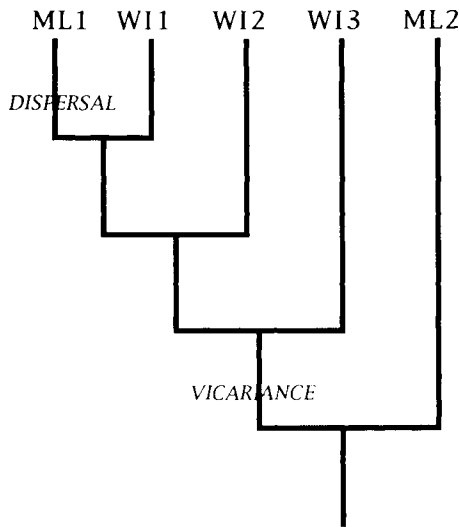


Figure 9.2. The importance of phylogeny for choosing taxa for pairwise comparisons when employing a molecular clock. ML = mainland, WI = West Indies. This “known” hypothetical phylogeny depicts a clade whose origins are in the mainland (ML2), populated the West Indies through vicariance, and subsequently dispersed back to the mainland (ML1). See text for further discussion.

The clock problems notwithstanding, Hedges *et al.* (1992) used nonindependent pairwise comparisons in their analysis. Crother and Guyer (1996) exacerbated the problem by mimicking an expanded data set. This technique fails to account for branch lengths (which imply rates of change), assuming that a phylogeny is correct (Hillis *et al.*, 1996). Branch length estimation itself is a source of error, but that aside, not taking it into account in pairwise comparisons can, by hiding varying rates of change, grossly underestimate calculation of confidence limits (Hillis *et al.*, 1996:534, fig. 9).

Additionally, the choice of taxa in pairwise comparison is critical for molecular clock applications to biogeography. In this sense (and for the moment ignoring branch lengths), a robust estimate of phylogeny is required. Assume a clade has several mainland (ML) and West Indian (WI) taxa. Further assume that the ancestors are ML, that subsequent vicariance led to the WI radiation, and that recent dispersal back to the ML occurred. From Fig. 9.2, if ML1 and any of the WI taxa are compared pairwise, an underestimate of divergence time will result. The closest estimate would come only from comparing ML2 and WI3. The only way to know this *a priori* is through the phylogeny.

Historical biogeography continues to struggle with the exigency of data while it frets over methods and principles. Caribbean biogeography has produced data of lasting value, and important conceptual advances. It has also encouraged remarkably imaginative speculation. We maintain, as have others, that phylogeny remains the basis for tackling the problems.

Ecological Biogeography

By whatever means an island accumulates a fauna – in units (vicariance), incrementally (dispersal) or by a combination of the two – the structural inertia of that fauna has biogeographic implications of its own. Ecological biogeography is the modern synthesis of those implications codified by MacArthur and Wilson (1963, 1967), and consecrated by Lack (1976). Its objective is to seek explanations about the composition of insular floras and faunas that are independent of history and phylogeny. Lack viewed island faunas as stable entities, a result of adaptation along continuous but nonoverlapping resource utilization by individual species. MacArthur and Wilson proposed that the identity of species would change through immigration and extinction, and that the

number of species present on an island was dependent on area and isolation. In the Caribbean, ecological biogeography found its cause célèbre in *Anolis* lizards. Following Etheridge's (1960) pioneer study of their relationships, anoles rapidly ascended to the rank of "model organism." The catapult was Ernest Williams' (1969, 1972; Rand, 1969; Rand and Williams, 1969) concept of the ecomorph, which characterized the repetitive adaptations of anoles to similar habitats on different islands. Subsequently, *Anolis* became the fruit fly of evolutionary ecology (e.g., Roughgarden, 1995). Although the literature is vast and varied (Schwartz and Henderson, 1991), our purpose here is simply to assess how these ecological studies have enriched Caribbean biogeography with respect to land vertebrates, and reptiles and amphibians in particular. We do not consider all ramifications, only the basic tenets of the MacArthur and Wilson model. We are especially interested in the reality of its application in the face of empirical evidence, both historical and contemporary. This line of inquiry is admittedly something of a strawman, a case of comparing apples with oranges. The intent of ecological biogeography is to imagine what could happen rather than to discover what did. On the other hand, we believe that history is inseparable from biogeography. In particular, events of the Holocene left indelible marks on Caribbean island biotas. The most trenchant of these was the arrival of humans.

MacArthur and Wilson and Lack were not the first to consider island biogeography in an ecological context. Island size, distance, and other potential determinants of species number had long been recognized by biogeographers (Gleason, 1922). In the West Indies, Darlington (1957) suggested that the number of amphibian and reptile species tended to increase by a factor of two for each 10-fold increase in island area. Levins and Heatwole (1963) presented statistical formalization of amphibian and reptile distribution throughout the Puerto Rican Bank based on factors of dispersal and species survival (see also Heatwole and MacKenzie, 1966). MacArthur and Wilson wanted to bring area and distance together in a unifying theme. For any island, they reasoned, there should exist an equilibrium number of species (S), based on an island's size and distance from the source pool. The concept of equilibrium supposes that the number of species on an island is constantly subject to increase by newly arriving colonists, but is balanced by concomitant extinction of the residents. Rates of immigration and extinction are specific to the island and taxon. For vagile taxa such as birds and bats, S is achieved more quickly than for less mobile groups such as lizards and frogs. An island also

must remain ecologically constant over a sufficient, but unspecified, period of time for the dynamics of immigration and extinction to attain a stable intersect.

MacArthur and Wilson's model was greeted with unrestrained enthusiasm by some biogeographers, with ridicule by others, and with skeptical curiosity by most. It is intuitively obvious that larger islands would have more species than smaller ones and distant islands fewer species than closer islands of equivalent size. That species number (S) is not attributable to anything more than chance alone was a frequent criticism. In one of the most stinging critiques, Sauer (1969:590) wrote:

In short, the equilibrium model and its derivatives suffer from extreme oversimplification by treating islands as functional units with no attention to internal habitat diversity and by treating species as interchangeable with no allowance for genetic or geographical diversity. This is not even good as a first approximation, because it filters out the interpretable signal instead of the random noise. A model that visualizes various sizes of assemblages of characterless species on various sizes of featureless plains is essentially absurd, since it excludes the very basis of genesis and continued co-existence of multiple species.

Taking the model at face value, few would disagree with Sauer (1969). However, it has kept breathing for over 30 years, sustained not by the reality of an equilibrium number of species per se, but by pursuit of the ecological reasons that explain nonequilibrium (Case and Cody, 1987). Hence, for amphibians and reptiles we would need to know, or at least have a sense of, (1) the capability of an animal to disperse, at what distances, and at what frequency; and (2) the factors that determine successful dispersal—that is, an individual(s) actually becoming established on a new island once it arrived.

Dispersal Capability

Dispersal capability is nearly impossible to determine even generally, and not at all with much precision; very few studies have examined this problem. Amphibians and reptiles have moved between Caribbean islands, no doubt, but there are only limited means for them to do so. Rafts and flotsam are the most obvious vessels available to transport them and their eggs (e.g., Censky *et al.*, 1998; King, 1962a). For amphibians, there are no viable alternatives except by becoming airborne during hurricanes. Lizards and snakes can float, and depending on their tolerance for saltwater, passive drifting on surface currents could make nearby islands accessible (Schoener and Schoener, 1983b). Some anoles

will voluntarily leap into the sea if stranded on wave-swept rocks, and can survive for at least 24 hr before making landfall (Schoener and Schoener, 1984).

Still, a rationale for *continuous* immigration does not hold up for most, if not all, nonvolant terrestrial vertebrates. The frequency of over water dispersal is remote and erratic even for supposedly capable lizards such as anoles. Were dispersal to occur successfully with some regularity, the diversity of West Indian *Anolis* species probably would be lower than it is currently. In the same way, the Lesser Antilles should have more than the present maximum of two species on any island.

Species-Area Relationships

Assuming that a lizard does disperse to a new island, the chances for successful colonization will depend on its health, sex, and the opportunities that await it. Ecological biogeography assesses these chances by extrapolating from the structure of existing communities. The most germane of the "assembly rules" that determine community structure are species per area, habitat complexity, and species interactions (e.g., Diamond, 1975; Haefner, 1988). Tests of the species-area relationship alone have been conducted for numerous taxa and islands. Most often the test is in the manner: $S = c A^z$, where area, A , is the independent variable, c is the y -intercept of the regression line, and z is a value that gives the slope of the regression line fitted to log-transformed data. During the 1970s many studies of insular plants and animals (although practically none on amphibians or reptiles) derived various values for z in hopes of uncovering a meaningful coefficient. The statistical interpretations were explored in detail, as were the assumptions of a log-normal distribution and abundance of species. At best, any unique theoretical basis for the S/A relationship is inconclusive beyond the fact that habitat diversity and area per se typically show a positive relationship to species number (Connor and McCoy, 1979). In a recent example, the incidence of most plant species on small islands surrounding Andros and the Exuma Cays in the Bahamas was positively related to some measure of area and negatively related to distance (Morrison, 1997). Cays with diverse vegetation are significantly higher and closer to mainland islands than cays naturally devoid. In another study, Dodd and Franz (1996) found that 42% of the distributional variation in the herpetofauna of the Exuma Cays Land and Sea Park was explained by island area. However, there was no correlation for lizards alone on the core islands nor with distance between satel-

lite and core islands. They also observed that cats and rats negatively affected local populations and probably their data as well.

The slope (z) and intercept (c) are essentially devoid of biological meaning for the simple reason that the number of species occurring on an island is a function of multiple causes, not simply area alone. No manner of statistical tinkering will consistently reflect the possibilities (Connor and McCoy, 1979). For example, in the British Virgin Islands Lazell (1983) tallied the number of lizard species present on 43 cays ranging in size from 0.1 to 5444 ha. Only two of the cays fit the species-area curve; the rest had "too many" species. Moreover, cays of 10 ha or less should, according to the math, have zero species. Using recomputed values for z , 26% of the cays had too few species, 32% had too many, and the minimum area required to support one species was reduced to 50 m². In terms of percentage error (the actual number of species too few or too many), a better fit was obtained by incorporating elevation into area values, but 33 of 43 islands still did not conform to predictions (Lazell, 1983). In some cases, "too few" species may be an artifact of insufficient collecting, and the island in question might, with additional field work, conform to the equation. Alternatively, the island could simply be so inhospitable that no lizard would survive on it, or perhaps the species never dispersed there in the first place. By contrast, "too many" species requires complex explanations and a bending of the assembly rules. Anoles are capable of occupying almost any piece of land above high tide that supports "more than herb-stage vegetation" (Lazell, 1983:115). Such is nearly true as well for other West Indian lizards such as *Sphaerodactylus*. Hence, species with low vagility but high adaptability inevitably will defy the species-area equation but in opposite directions so that too few or too many species, respectively, will result.

Species Turnover

The equilibrium model supposes that immigration and extinction are continuous and linked. The outcome is a turnover in species composition. Most reputed examples of species turnover on West Indian islands come from short-term studies on birds and Lesser Antillean anoles. Turnover results from the extinction of a resident species as a consequence of an invader becoming established. Turnover, then, is driven by species interaction, especially competition. On smaller islands with fewer species, rates of turnover should be relatively high (e.g., Schoener and Schoener, 1983b; but see Simberloff and Boeklin, 1991). On larger, species-rich

islands invader success should be lower because diverse, strongly interacting communities manifest emergent properties that make them resistant to newcomers (Case, 1991). Lack (1976), by contrast, viewed interaction as important, but in such a way that turnover eventually approaches zero. Short term interactions should give way to stability in the long term because species evolve niche specializations based on opportunity and ecological resources.

Species turnover has its origin in the "taxon-cycle" first conceptualized by Wilson's (1961) studies on Melanesian ants. The concept was applied early on to West Indian birds by assuming that a species progresses through stages of distribution beginning with expansion in range, differentiation, contraction, and finally extinction (Ricklefs, 1970; Ricklefs and Cox, 1972, 1978). That these stages are successional is at best dubious. They are merely a set of distributional patterns that almost any vertebrate in any archipelago will fall into, and there are several possible explanations to account for each without ever going through such a "cycle" (Pregill and Olson, 1981).

For reptiles, a taxon cycle was proposed for two species of *Sphaerodactylus* lizards on St. Croix (MacLean and Holt, 1979). Distributional information gathered there in the 1930s was compared with the results of field work 40 years later. During that interval the range of the endemic *S. beattyi* had contracted, whereas that of an invading species, *S. macrolepis*, had expanded. The two lizards are similar in size and habitat preferences, but their distributions are sharply defined on the island. They segregate in part by different physiological tolerances for water loss; *S. beattyi* is better adapted to xeric microhabitats (MacLean and Holt, 1979). If the range of either species would be greater on the island in the absence of the other, then competitive interaction is probably operating. However, this is not a taxon cycle in the sense of Wilson (1961) or Ricklefs and Cox (1972, 1978), which would require that the species actually passed through the stages from expansion to extinction.

In the West Indies the taxon cycle achieved notoriety in studies of Lesser Antillean anoles. The distribution of these lizards is characterized by the repetition of size morphs on either one- (intermediate body size) or two-species (large and small body size) islands. A variant of the taxon cycle explains this pattern in the form of an invasion/coevolution model (Pacala and Roughgarden, 1982; Roughgarden and Pacala, 1989; Roughgarden, 1995). A larger, invading species outcompetes a smaller resident species and eventually pushes it to extinction. The invader then stabi-

lizes at the intermediate solitary size, whence, potentially, the cycle repeats itself. This model has received considerable scrutiny, and other authors have argued for character displacement or size assortment within the constraints of phylogeny (Losos, 1992b; Miles and Dunham, 1996; Giannasi *et al.*, 1997). For example, two species in sympatry may evolve in different directions to avoid competition, viz size displacement. Conversely, perhaps only species of different size could coexist on the island in the first place and therefore must have evolved allopatrically.

We add that mean body size of insular lizards varies over time in response to changing circumstances of habitat quality and predation that are independent of interactions with other species of lizards. There is ample fossil evidence, for example, demonstrating a reduction in body size during the Holocene for species in the Caribbean and elsewhere. This reduction correlates with human settlement and associated declines in environmental quality, and introductions of nonnative predators (Pregill, 1986; Pregill and Dye, 1989). Proof of a taxon cycle must come from data showing that a species actually went through the stages, whether the stages are defined as changing distribution patterns or as evolutionary changes in size. Given the time involved, a fossil record presumably would be required, and thus far the record reveals no such pattern.

Holocene Extinctions

We noted earlier that historical evidence has minimal relevance in ecological biogeography, which is why the Holocene fossil record is an under-utilized resource in such studies (but see Case *et al.*, 1992). Events of the past 10,000 years, however, have altered the ecological complexion of most all islands through extinction and landscape transformations. As a result, whatever natural processes of species composition remain are not easily identified. The West Indian Holocene record, although far from complete, now lists hundreds of vertebrate populations and species that disappeared locally, regionally, or completely. About 80% of the West Indian land mammal fauna no longer exists (Morgan and Woods, 1986; MacPhee and Marx, 1997), and many of the 20 orders and 64 families of resident birds also include extinct representatives (Olson and Hilgartner, 1982; Cuello, 1988). Large tortoises, freshwater turtles, snakes, and lizards vanished as well (e.g., Olson *et al.*, 1990). This comparatively recent loss of diversity means that present ecological associations do not reflect a full roster of participating species. It means

that caution is called for when interpreting current distributions, especially those that appear relictual, or are incongruous with respect to a broader pattern. In many instances current distributions have resulted from recent extinctions on intervening islands. Extinction is of course predicted by the equilibrium hypotheses as a consequence of new immigrants colonizing an island. The difference, however, is that none of the Holocene extinctions is attributable to species turnover. For example, *Cyclura pinguis* is known historically only from tiny Anegada Island, but until the late Holocene this large rock iguana ranged throughout much of the Puerto Rican Bank (Pregill, 1981b). On a broader front, at least some of the current Greater Antillean vertebrate fauna actually extended well into the Lesser Antilles during most of the Holocene. Paleofaunas from the Leeward Islands of Anguilla, Antigua, and Barbuda record several extinct vertebrates (species and genera) that today are confined to the Greater Antilles. Among these is the Puerto Rican Parrot *Amazona vittata* that today survives in small numbers in Puerto Rico's Luquillo forest and is regarded as endemic to that island. Other examples include a thick-knee (*Burhinus* sp.), a crow (*Corvus* sp.), the bats *Mormoops blainvillei*, *Macrotus waterhousii*, and the extinct *Phyllonycteris major*, and lizards of the genera *Cyclura* and *Leiocephalus* (Steadman *et al.*, 1984a; Pregill *et al.*, 1988, 1994).

Much of the evidence for a faunal collapse during the Holocene occurs near the time of first human contact. We do not know, however, if the Holocene extinctions of West Indian vertebrates were relatively abrupt for some or most taxa, were gradual over thousands of years for others, or the extent to which humans were the proximate cause in either case (Steadman *et al.*, 1984a; Morgan and Woods, 1986; MacPhee *et al.*, 1989; Pregill *et al.*, 1994). Nevertheless, that the losses post-date human arrival is, by itself, strong evidence of direct or indirect human involvement (Martin and Steadman, 1998). There are, unfortunately, very few well-dated, fossil-bearing strata available to stabilize the chronology. Some extinctions may be related to changes in climate, vegetation, land area, sea level or other non-anthropogenic effects associated with the glacial/interglacial transition 10,000-12,000 years ago, or with other natural phenomena later in the Holocene. The late Pleistocene climate of the West Indies appears to have been characterized by aridity that gave way to more mesic conditions at the beginning of the Holocene. The fossil record includes a high proportion of xeric-adapted species that became extinct at that time, especially in the Greater Antilles (Pregill and

Olson, 1981). As in other parts of the globe, there may have been mid-Holocene climatic fluctuations of sufficient magnitude that the environmental changes brought on by people were exacerbated. For example, intense late Holocene aridity is indicated in Madagascar that acted in concert with human hunting and fire regimes. The combined effects were disastrous for native vertebrates, especially birds and mammals (Burney, 1993). Sediment cores from the Persian Gulf also indicate two to three centuries of severe drought in this region about 2000-3000 years ago (Kerr, 1998).

The West Indian Holocene record derives from both paleontological (non-cultural) and archaeological (cultural) contexts. Non-cultural fossils are almost exclusively the accumulated remains of owl pellets deposited in caves and sinkholes. Bone records from archaeological sites derive from the "kitchen middens" of Amerindians. Compared with the Greater Antilles, there are far fewer non-cultural sites in the Lesser Antilles, although archaeological sites abound (Pregill *et al.*, 1994). The most desirable vertebrate record would be one from a site spanning three key horizons: (1) Prehuman – deposits in place prior to the arrival of people to the Antilles; (2) Prehistoric (pre-Columbian) – deposits dating from the appearance of indigenous people up to the time that Europeans and Africans arrived, about 500 years ago; and (3) Historic (post-Columbian) – deposits that date from the time of European/African contact. For any island, calibrating the prehuman/human interface is critical in order to establish a minimum age for anthropogenic effect. Next, it is important to ascertain which species persisted into the Historic period (and there were some) but disappeared soon after Europeans began unloading their ships. Besides their technology, Europeans brought exotic predators and competitors both domestic (goat, pig, dog, cat) and commensal (black rat). Historic settlement may have accelerated extirpation that began in the Prehistoric period, or in some cases may have initiated it.

Dated, bone-bearing strata directly associated with first human contact are rare in the Antilles. However, the chronology of human arrival itself is retrievable from archaeological sites that demonstrate an established cultural presence. Still, earliest cultural evidence may lag by several or more centuries the initial arrival of small human cohorts. Even these small, early groups could have modified the landscape through repeated burning. For example, in Puerto Rico sediment core analysis from Lake Tortuguero on the north central coast revealed a spike in charcoal stratigraphy indicative of human presence at ca. 5500 years before

present (ybp), approximately two millennia earlier than the oldest archaeological dates from that island (Burney *et al.*, 1994). This puts the occupation of Puerto Rico in synchrony with Hispaniola and Cuba, which have archaeological sites dated to around 6000 ybp (Moore, 1991). This is a minimum age and people may well have arrived in the Greater Antilles (except Jamaica) as early as 7000 ybp (Rouse, 1989).

In the Lesser Antilles, the colonization by Amerindian peoples from coastal regions of northeast South America occurred by 2000 BC (or perhaps even earlier) in the period known as the Archaic Age of that region (Keegan and Diamond, 1987; Rouse, 1992). Lesser Antillean Archaic Age sites are usually coastal and often near mangrove stands (Davis, 1982). These people were neither agrarian nor potters. Faunal remains from their occupation sites are poorly studied, and indicate only that the first Amerindians primarily exploited marine and estuarine animals, especially mollusks (Pregill *et al.*, 1994). Their impact on the terrestrial vertebrate fauna is not known. By about 500-400 BC, another northward migration began, but of ceramic-producing people that for the next thousand years developed agricultural practices, and exploited terrestrial as well as marine resources. They also transported exotic species from South America (e.g., the agouti *Dasyprocta* sp.), and shuttled native ones (e.g., *I. iguana*, some parrots, *Amazona*) among the islands (Steadman *et al.*, 1984b). Within about 200 years following European settlement (late 1700s) most aboriginal peoples in the West Indies had been extirpated through warfare, disease, enslavement, and interbreeding, although remnant groups have persisted on a few islands even to the present.

In both the Greater and Lesser Antilles there were several millennia of human-engendered changes to the landscape and fauna before biologists arrived to collect plants and animals, and to study ecosystems. Native vertebrates on smaller islands were especially vulnerable from the beginning. In the Lesser Antilles, the Holocene fossil record shows that the current vertebrate fauna is not nearly as rich as it was several thousand years ago. From Sombrero to Grenada human activities of recent millennia resulted in the loss of indigenous rice rats, birds, lizards, snakes, and tortoises. At least 79 indigenous populations or species of vertebrates became extinct on islands from which fossil specimens of them are known (Pregill *et al.*, 1994). The extinct and extirpated herpetofauna, for instance, is represented by 18 species from 10 islands and in multiple trophic levels (Table 9.1). Because of poor chronologies, it is

Table 9.1. Extinct and Extirpated Amphibians and Reptiles of the Lesser Antilles^a.

Species (location)	Extinct, historic record	Extinct, no historic record	Extinct, known only from fossils
<i>Leptodactylus fallax</i> (St. Kitts, Guadeloupe, St. Lucia)	X		
<i>Geochelone</i> sp. (Barbados)			X
<i>Geochelone</i> <i>sombrenensis</i> (Sombbrero)			X
<i>Anolis bimaculatus</i> giant morph ^b (Antigua, Barbuda)			X
<i>Anolis watsi</i> (Anguilla)	X		
cf. <i>Cyclura</i> sp. (Barbuda)			X
<i>Iguana delicatissima</i> (St. Kitts)		X	
<i>Iguana iguana</i> (Barbados)		X	
<i>Iguana</i> sp. (Marie Galante)		X	
<i>Leiocephalus cuneus</i> (Antigua, Barbuda)			X
<i>Leiocephalus</i> cf. <i>cuneus</i> (Anguilla, Guadeloupe)			X
<i>Leiocephalus herminieri</i> * (Martinique)	X		
<i>Ameiva cineracea</i> (Guadeloupe)	X		
<i>Ameiva major</i> * (Martinique)	X		
<i>Boa constrictor</i> (Antigua)		X	
Boidae sp. (Antigua)			X
<i>Alsophis</i> sp. (Barbuda)		X	
cf. <i>Clelia clelia</i> (Barbuda)		X	

*Taxa with an asterisk have no fossil record and have become extinct historically (from Pregill *et al.*, 1994).

^bPregill *et al.* (1988).

difficult to estimate which and how many species persisted into the Historic Age on a given island. Only two of the 15 extinct or extirpated species of reptiles with a fossil record are known historically by specimens. Six others are extant elsewhere in the Lesser Antilles, and seven are known by fossils only.

Holocene faunal losses are not peculiar to the West Indies. World-wide, islands experienced various degrees of extinction of native species coincident with first human contact (Burney, 1993). Pacific island ecosystems from Hawaii to New Zealand were profoundly altered by the people who first settled them, sometimes within a few generations (Kirch and Hunt, 1997). Overall, the pattern for oceanic islands follows that of the peopling of the New World and Australia, which suffered substantial losses in their megafaunas at and during the time humans colonized them. There is not universal agreement about the causes of the continental extinctions (Martin and Klein, 1984) or that in North America native peoples who arrived there around 12,000 years ago (Reitz *et al.*, 1996) had anything to do with them. The point, however, is that the European's "New World" of 500 years ago, both continental and insular, had already been modified by people to varying degrees.

For equilibrium biogeography there is an important message: the distribution and abundance of plants and animals in the West Indies (and on other islands) cannot be viewed principally as the outcome of natural ecological processes. Contemporary faunas and populations are merely a fragmented subset of what existed a few thousand or even hundreds of years ago. Habitat utilization and trophic structure have been rearranged, and if this is not taken into account attempts to model island biotas realistically will fall short. Anoles, for example, may be more abundant today because of their ability to exploit patchy, discontinuous habitats resulting from human land use (Rand and Williams, 1969). MacArthur, Wilson, and Lack are deservedly among the greatest names in twentieth-century biogeography. At the time of their syntheses on island biotas, there was no knowledge of the Holocene and its consequences. Today, without applying this knowledge, ecological biogeography founders in its attempt to elucidate species origins and distributional patterns. In a practical sense, it will remain stuck in the subjunctive so long as history is ignored.

Coda

When we back away from Caribbean biogeography the most solvent contributions in view are the careful documentation of species on each and every island, now and in the past. When practiced with sound taxonomy, survey and inventory represent scholarship of lasting value. Albert Schwartz appreciated this better than anyone. In the West Indies new records of taxa are still being made, albeit at a slower pace than several decades ago (see Chapter 1). Still, surprising discoveries await, especially paleontological ones. Contemporary herpetologists and paleontologists are in a foot-race to find them before they are lost to ever-expanding human land use. The effort, expense, and skill required often go unrecognized while disproportionate celebrity is extended to speculative ventures in ecological theory. Without detailed knowledge of distributions and variation, biogeography is stuck at ground zero. Put another way, the Schwartz and Thomas (1976) checklist is the most valuable contribution to the biogeography of West Indian amphibians and reptiles in the past 50 years.

The West Indies and Middle America: Contrasting Origins and Diversity

William E. Duellman

*Natural History Museum and Department of Systematics and Ecology
The University of Kansas, Lawrence, Kansas 66045*

Introduction

The geography of North and South America is unique in having two large continents attached to one another on one side by a narrow connection and partially bridged on the other side by a series of islands. Historically, the two continents never have been closer, because they were widely separated since the breakup of Pangaea and have drifted closer throughout the millions of years since the fragmentation of Gondwanaland. Geological evidence, albeit not always consistent, suggests that what is now Central America was an archipelago in the late Cretaceous and subsequently some of these islands moved eastward to form what is now the Greater Antilles. By the Miocene another chain of volcanic islands emerged between nuclear Central America and South America; continued orogeny and erosion eventually formed a long, narrow peninsula which in the late Pliocene connected with South America. Also during the Cretaceous, another chain of islands, the Lesser Antilles, was inserted between the Greater Antilles and South America. These historical events and their biogeographic significance have been discussed at length by various authors in two volumes—Stehli and Webb (1985) and Woods (1989).

Although the herpetofaunas of Middle America and the West Indies have been studied intensively for more than half a century, no critical comparisons of the two faunas have been made. This is especially compelling because both regions received many lineages of their biotas from the same source areas—North America and South America—yet their compositions are extremely different. Thus, the biogeography of these two areas provides a fascinating challenge for retrodiction. This challenge is especially intriguing because no single set of hierarchical expla-

nations seems to apply to the West Indian herpetofauna, a problem addressed eloquently by Williams (1989b).

Materials and Methods

For purposes of this analysis, Middle America is defined as the Central American countries and southeastern Mexico east of the Isthmus of Tehuantepec; therefore, the rest of Mexico is part of North America. The West Indies exclude the Netherlands Antilles (Aruba, Bonaire, and Curaçao) and the continental islands of Trinidad and Tobago. Areas of regions were obtained from various atlases, and that for Mexico east of the Isthmus of Tehuantepec was determined by use of a Micro-Plan II image analysis system (Laboratory Computer Systems, Inc., Cambridge, Ma.). Composition and taxonomy of the West Indian herpetofauna was updated from Schwartz and Henderson (1991). For Middle America, these data were summarized from Villa *et al.* (1988).

Origins of the Herpetofaunas

Mathewsian biogeographers (e.g., Dunn, 1931; Darlington, 1938) believed in the fixity of continental platforms and interpreted biogeographic patterns entirely by the mechanism of dispersal. With respect to Middle America and the West Indies, this complacency was shattered by Rosen's (1976) application of revolutionary geological interpretations of plate tectonics and elaborated upon by Rosen's (1978) broader ramifications on historical biogeography. Rosen's biogeographic scenario was based primarily on Malfait and Dinkelman's (1972) plate tectonic model that postulated a series of islands, the proto-Antilles, forming in the region that is now lower Central America and drifting eastward to become the Greater Antilles; subsequently, another series of islands formed, expanded, and interconnected to form the present-day lower Central America, while the basement of most of the Lesser Antilles was exposed in the upper Cretaceous, and the islands were formed principally by volcanism in the Tertiary (Speed *et al.*, 1979). A major fault zone lies between Dominica and Martinique; this Dominican Fault and associated geological evidence strongly suggests independent histories of the northern and southern Lesser Antilles (Bouysson, 1984; Speed, 1985). The geological evidence has been expanded and reinterpreted in a variety of ways (e.g., Donnelly, 1989; Perfit and Williams, 1989; Pindell and Barrett, 1990; Pitman *et al.*, 1993); according to Pittman *et al.*, the complexity of the plates and their associated tectonic events obviates a consensus of interpretation.

This geological evidence necessitated re-evaluation of biogeographic data. Conflicting hypotheses of dispersal versus vicariance have been proposed for both West Indian and Middle American phyletic lineages of the biota. The vicariance model involves the separation (vicariance event) of a continuous distribution of a taxon by the formation of a barrier, whereas the dispersal model involves the dispersal of organisms across a pre-existing barrier. The implications relative to the proposed proto-Antilles and the lower Central American archipelago are obvious. Savage (1982) championed the vicariance model for the Middle American and West Indian herpetofaunas, whereas others, notably Pregill (1981c), Williams (1989b), and Hedges (1996b) supported the dispersal model for the West Indian herpetofauna. Williams (1989b:22), a staunch dispersalist, argued

Between these two competing historical hypotheses [dispersal and vicariance] I do not see any a priori grounds for choice. Drifting islands do not make vicariance per se more plausible or dispersal less so. In fact, in the whole history of the West Indies, as now understood, I do not see how dispersal on any present evidence can be ruled out. A moving Antillean complex alters distances between islands and mainlands and among the islands. It makes dispersal easier or more difficult. It says, however, nothing per se about faunas or how they got where they are. Nearness assists dispersal, distance makes dispersal difficult and very improbable. Nearness is not enough for vicariance, and distance makes it, by definition, impossible. To falsify dispersal in the Caribbean even in the earliest Tertiary will require evidence that we do not now have.

More recent and an entirely different kind of geological evidence now pervades the controversy. A presumed huge bolide impact off the coast of Yucatan is estimated to have created a huge crater (300 km across) at the Cretaceous/Tertiary boundary (Hildebrand and Boynton, 1990; Sharpton *et al.*, 1993). Hedges *et al.* (1992b) speculated that the proto-Antilles were the closest islands to the impact site and likely could have been devastated by the effects of this impact, especially giant tsunamis—waves estimated to have been four or five km in height at the point of impact (Hildebrand and Boynton, 1990). In this scenario, the biota on these Noah's arks (*sensu* McKenna, 1973) might have been obliterated, or the diversity might have been greatly reduced. However, because the impact site was in shallow water (<200 m), the wave height would have been no more than 200 m at the point of impact and greatly diminishing peripherally (Ward *et al.*, 1985). Thus, Crother and Guyer (1996) argued that the bolide impact would not have obliterated the biotas on islands having major physical relief and thereby discarded the hy-

pothesis of Hedges *et al.* (1992b) that the biota of the Greater Antilles was mostly eliminated at Cretaceous/Tertiary boundary.

The major problem in dealing with the biogeography of the herpetofaunas of Middle America and the West Indies is a lack of evidence defining monophyletic lineages and the absence of phylogenetic analyses of taxa within those lineages. The two largest genera, *Anolis* and *Eleutherodactylus*, inhabiting both regions have been subjected to at least partial analyses. Guyer and Savage (1987) subjected data on *Anolis* to phylogenetic analyses resulting in the recognition of five genera of species formerly placed in *Anolis*, and they proposed a biogeographic hypothesis based on vicariance. Their methodology was evaluated critically by Cannatella and de Quieroz (1989), who concluded that Guyer and Savage's "proposed taxonomy is neither demanded by, nor consistent with, the results of phylogenetic analysis." Williams (1989a) rigorously criticized Guyer and Savage's work on the basis of "serious errors and confusions" and recommended that neither the databases nor the taxonomy be accepted without reexamination of the primary evidence (but see rebuttal by Guyer and Savage, 1992). Roughgarden (1990, 1995) reviewed the biogeography of *Anolis* in Puerto Rico and the Lesser Antilles; he concluded that the *Anolis oculatus*, *cuvieri*, and *crisatellus* groups (with the exception of the Bahaman *A. scriptus* in the latter group) are restricted to islands on the Puerto Rican bank, whereas the *Anolis bimaculatus* group is restricted to the Lesser Antilles north of the Dominica Channel and the *Anolis roqueti* group inhabits the Lesser Antilles to the south of that channel.

Investigations by Lynch (1986), Hedges (1989a), and Joglar (1989) have resulted in the recognition of five subgenera of *Eleutherodactylus*. Some major discrepancies exist between Hedges' (1989a) results based on immunological distances and Joglar's (1989) results based on morphology. The subgenera *Euhyas* and *Pelorius* are endemic to the West Indies, and the phylogenetically composite subgenus *Eleutherodactylus* occurs in the West Indies and Middle America and is widespread in South America. The subgenus *Syrrhophus* is principally Mexican, whereas the subgenus *Craugastor* is widespread in Middle America and also extends into northwestern South America. Of these supposed five lineages, only *Craugastor* is demonstrably monophyletic (Lynch and Duellman, 1997).

Only a few other genera have been subjected to significant phylogenetic analyses. According to Pregill (1981a), the West Indian toads are a monophyletic group (*Peltophryne*), but this lineage is derived from within the South American *Bufo granulatus* group (Hedges, 1996b). Hylid frogs are highly diverse in Middle America (*Agalychnis*, *Anotheca*,

Duellmanohyla, *Plectrohyla*, *Ptychohyla*, and *Triprrion* endemic or nearly so) and far less diverse in the West Indies (five species of *Hyla* and the endemic genera *Calyptahyla* and *Osteopilus*, the latter with three species) (Trueb and Tyler, 1974). However, Hedges (1996b) remarked that molecular data indicate that, with the exception of *Hyla heilprini*, the West Indian hylids form a monophyletic group. The relationships of most of the lineages of hylids within Central America remain obscure, but a phylogeny of *Plectrohyla* is available (Duellman and Campbell, 1992; Wilson *et al.*, 1994), as is one for the genera *Duellmanohyla* and *Ptychohyla* (Campbell and Smith, 1992). The *Eleutherodactylus rostralis* group of *Eleutherodactylus* was subjected to a phylogenetic analysis by Savage (1987). Although detailed phylogenetic analyses of the highly diverse Middle American salamander fauna have not been published, many studies (e.g., Wake, 1987; Wake and Lynch, 1976; Wake and Elias, 1983; Wake and Johnson, 1989) of the bolitoglossines reveal relationships with North America and separate radiations in the Mexican highlands, nuclear Central America, and lower Central America.

Among lizards, the gekkonid genus *Sphaerodactylus* occurs on the mainland in Middle and South America but is especially diverse in the West Indies. With two possible exceptions, the mainland species represent a monophyletic lineage (Harris and Kluge, 1984), and Hass (1991) postulated that the West Indian species are a monophyletic lineage. The iguanian lizard genus *Leiocephalus* is demonstrably monophyletic (Pregill, 1992). However, its relationships remain controversial. Although placed in its own subfamily as a sister group to the South American Tropicurinae by Frost and Etheridge (1989), Hedges *et al.* (1992b) suggested that *Leiocephalus* is more closely related to North American crotaphytids and phrynosomatids. On the other hand, recent evidence places *Leiocephalus* as the sister group to Madagascan oplurines (D. R. Frost, personal communication). Xantusiid lizards are distributed in southwestern North America and Middle America with *Cricosaura typica* endemic to Cuba. Analyses by Crother *et al.* (1986) placed *Cricosaura* as the sister group to the Middle American *Lepidophyma*, but Hedges and Bezy (1994) concluded that *Cricosaura* is the basal lineage in the family. The systematic relationships among West Indian and mainland species of *Celestus* and *Diploglossus* are in such disarray that generic assignment is controversial for some species, but *Sauresia* and *Wetmorena* are part of a radiation of West Indian *Celestus* (Campbell and Camarillo, 1994; Hedges *et al.*, 1992b; Hedges, 1996b). The mainland gerrhonotine anguids were analyzed by Good (1988), but his results were challenged by Campbell and Frost (1993) in their analysis of *Abrotonia*.

Among the snakes in the West Indies, the speciose genera *Epicrates* and *Tropidophis* are represented on the South American mainland by only one and three species, respectively; the West Indian lineages are monophyletic (Kluge, 1989; Hedges *et al.*, 1992b). Within the morass of the Colubridae, six "alsophine" (Xenodontinae) genera (*Antillophis*, *Arhryton*, *Darlingtonia*, *Hypsirhynchus*, *Ialtris*, and *Uromacer*) are endemic to the Greater Antilles, and *Alsophis* occurs in the Lesser Antilles and South America. Although there is immunological evidence for relationships of the West Indian genera with South American xenodontines (Cadle, 1984a, 1985), the monophyly of the alsophines remains questionable. Cadle (1984a,b,c, 1985) identified two major clades of xenodontine colubrids—a South American clade and a Middle American clade. Of the six West Indian endemic *Leptotyphlops*, all but *L. columbi* in the Bahamas represent a single radiation (Thomas *et al.*, 1985), but their relation with mainland American taxa is unknown. The majority of New World *Typhlops* occur in the West Indies; Thomas (1989) identified two groups, one with African and the other with New World species, but Hedges *et al.* (1992) indicated that the West Indian taxa form a monophyletic group. Mainland taxa are even less well-known; the only substantive phylogenetic analyses of Middle American genera are those of the colubrid *Adelphicos* by Campbell and Ford (1982) and Campbell and Brodie (1988), and of the viperid *Bothriechis* by Crother *et al.* (1992).

Savage (1982) made a strong argument for the recognition of a distinctive Mesoamerican herpetofauna, the earliest origins of which were from the ancestral tropical American herpetofauna that was fragmented by the inundation of the land connection between Central and South America in the early Tertiary; this biota was enhanced by subsequent isolation of stocks of northern affinities. Thus, ancestral stocks of South American and North American origin differentiated *in situ* through vicariance, and these vicariants speciated throughout most of the Tertiary. The result was a distinctive Mesoamerican herpetofauna amounting to more than 70 genera and hundreds of species, including essentially all bolitoglossine salamanders; frog genera such as *Atelophryniscus*, *Plectrohyla*, and *Ptychohyla*; lizard genera such as *Corytophanes*, *Laemancetus*, and *Lepidophyma*; snake genera such as *Conopsis*, *Drymobius*, and *Scaphiodontophis*; and turtle genera such as *Claudius* and *Staurotypus*. The distinctiveness of the Mesoamerican biota was obscured somewhat by the biotic interchange across the Panamanian isthmus beginning in the Late Pliocene, when South American elements dispersed into Central America; these invaders include centrolenid, dendrobatid, and hemiphractine frogs, gymnophthalmid lizards, and diverse kinds of snakes (e.g., *Boa*, *Bothrops*, *Epicrates*, *Liophis*, and *Xenodon*). Likewise, a few

North American groups dispersed into Central America; these included *Rana*, *Sceloporus*, *Coluber*, *Kinosternon*, and natricine snakes. Some of these North American groups (e.g., *Rana*, *Sceloporus*, and *Kinosternon*) had undergone major speciation in the southwestern parts of their ranges, principally on the Mexican Plateau. Likewise, many Mesoamerican groups dispersed into South America, including *Bolitoglossa*, *Oedipina*, *Agalychnis*, *Smilisca*, *Basiliscus*, *Coniophanes*, and colubrine snakes.

Whereas the evolution and biogeography of the Mesoamerican herpetofauna is reasonably well substantiated (albeit there are some controversial groups), Savage's (1982) attempt to explain the origin of the Greater Antillean herpetofauna by a vicariance model involving vicariance of the archaic Mesoamerican elements on the proto-Antilles suffers from more controversial evidence. Hedges (1996b) estimated divergence times from albumin immunological data (see Chapter 9) of most lineages of the West Indian herpetofauna and concluded that most lineages came from South American sources (supported by patterns of ocean currents). Furthermore, he postulated that stocks dispersed into the West Indies throughout the Cenozoic. According to Hedges (1996a), the *Euhyas* stock of *Eleutherodactylus* and the xantusiid lizard *Cricosaura typica* are the only examples of Middle America/Greater Antillean vicariance that possibly were involved with the proto-Antilles. However, Crother and Guyer (1996) recalibrated Hedges' "clocks" and revised divergence dates based on geological evidence; they concluded that with the exception of lineages endemic to Jamaica, more than 60% of the lineages could have been shared by the mainland and the Greater Antilles.

Savage (1982) and Hedges (1996b) agreed that the herpetofauna of the more recent Lesser Antilles was derived from South America by dispersal; in fact, many of the species inhabiting these islands are distributed on the South American mainland. However, there is geological evidence that there was continuity between northern South America and the southern Antillean plate in the Cretaceous (Speed, 1985); there is a distinct difference between the herpetofaunas in the northern and southern Lesser Antilles (Roughgarden, 1995). Thus, some of the lineages in the Lesser Antilles may be the result of vicariance.

Most of the West Indian herpetofauna seems to have been derived from South America, although many lineages (e.g., *Eleutherodactylus unistrigatus* group, *Sphaerodactylus*, *Gonatodes*, *Mabuya*, *Ameiva*) are diverse in South America, Middle America, and the West Indies. Only two genera, *Celestus* and *Tretanorhinus*, are shared exclusively by the Greater Antilles and Middle America. If the Greater Antillean herpetofauna was derived by vicariance from the Mesoamerican herpetofauna in the early Tertiary, one might expect more lineages or pairs of lineages to

be common to the two regions. Whereas the North American contribution to the Middle American herpetofauna was great and included early stocks that differentiated in Middle America (e.g., bolitoglossine salamanders, sceloporine lizards, *Eumeces*, *Thamnophis*, and *Crotalus*) and later dispersalists, North America made an insignificant contribution to the herpetofauna of the West Indies—only turtles of the genus *Trachemys* and the natricine snake *Nerodia clarki*.

Diversity of the Herpetofaunas

To anyone contemplating the composition of the herpetofaunas of the West Indies and Middle America, immediately obvious are the absence of caecilians and especially salamanders in the West Indies and the diversity and abundance of some genera, especially *Eleutherodactylus*, *Anolis*, and *Sphaerodactylus*, in the West Indies. As emphasized by Williams (1989b), the herpetofauna of the West Indies is depauperate in the number of genera (lineages) represented. However, contrary to a commonly held misconception, the islands support a herpetofauna rich in species. The native, non-marine herpetofauna of Middle America consists of 793 species in an area of 692,886 km², whereas that of the West Indies consists of 613 species within a land area of 232,392 km². Because of the much greater area of Middle America, the density of species per area (1.14 per square km⁴) is less than one-half that in the West Indies (2.64 per square km⁴).

Within Middle America, the largest number of species occur in Guatemala and Costa Rica (with Panama a close third), whereas in the West Indies, the largest numbers are in Hispaniola and Cuba (Table 10.1). The herpetofaunal diversity in Guatemala (366 species with a density of 3.4 per square km⁴) and Costa Rica (361 species with a density of 7.1 per square km⁴) is associated with the highlands in both countries, whereas that in Panama (350 species with a density of 4.5 per square km⁴) is enhanced by the continuation of the Cordillera de Talamanca from Costa Rica into Panama, as well as numerous South American taxa (e.g., *Pleurodema*, *Pipa*, *Enyalioides*, and *Atractus*) that enter eastern Panama. In contrast, the primarily lowland countries, Belize and El Salvador, have the smallest herpetofaunas—131 species with a density of 5.7 per square km⁴ and 120 species with a density of 5.6 per square km⁴, respectively. Densities for other Middle American units are as follows: southeastern Mexico, 1.3; Honduras, 2.1; and Nicaragua, 1.6 species per square km⁴. The low species density in southeastern Mexico reflects the large area of the Yucatan Peninsula, in which relatively few

Table 10.1. Comparative Composition of the Native, Non-Marine Herpetofaunas in Middle America and the West Indies. Areas are in square kilometers; numbers are species.

Geographic Unit	Area	Anura	Amphibians	Reptiles	Total
SE Mexico	239,555	73	94	216	310
Belize	22,965	27	29	102	131
Guatemala	108,889	95	133	233	366
Honduras	112,088	57	74	162	236
El Salvador	21,393	25	31	89	120
Nicaragua	130,000	52	58	152	210
Costa Rica	50,900	120	149	212	361
Panama	77,096	127	156	194	350
Middle America	692,886	246	337	456	793
Cuba	114,524	50	50	101	151
Hispaniola	76,484	63	63	138	201
Jamaica	10,962	23	23	31	54
Puerto Rico	8,897	20	20	47	67
Bahama Bank	14,368	2	2	41	43
Lesser Antilles	7,157	10	10	84	94
West Indies	232,392	167	167	446	613

species occur; also, the low density in Nicaragua is a result of the absence of extensive highland regions.

Among the Greater Antilles, Hispaniola has the largest herpetofauna (201 species) and a higher density of species per area (2.63 per square km⁴) than Cuba with 151 species and a density of 1.3 per square km⁴. However, Puerto Rico has 67 species with a density of 7.5 per square km⁴, and Jamaica has 54 species with a density of 4.9 per square km⁴. The large number of species (94) and high density (13.1 per square km⁴) in the Lesser Antilles has two causal factors: (1) the high degree of endemism (especially of *Anolis* and *Sphaerodactylus*) on particular islands and (2) the large number of South American taxa (e.g., *Bachia*, *Kentropyx*, *Thecadactylus*, *Corallus*, and *Bothrops*) present on the islands. In contrast, the Bahama Bank has only 43 species with a density of 2.9 per square km⁴. The low density is owing to the fact that only two

amphibians are present, and many of the species of reptiles are widespread on the islands.

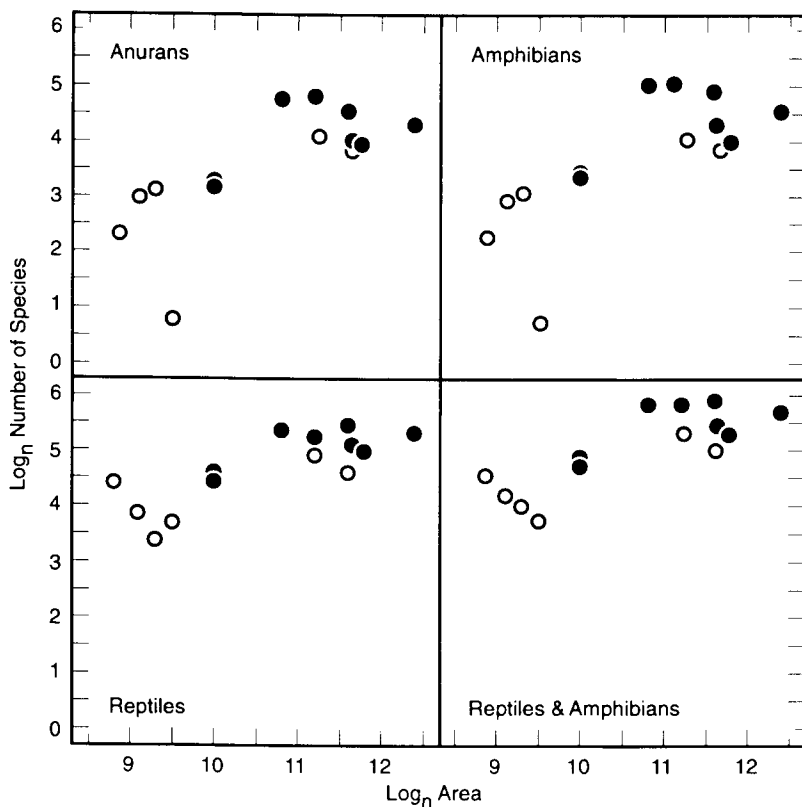


Figure 10.1. Log-log plots of numbers of species and areas of geographic units (see Table 10.1 for data). Solid dots are countries in Middle America; open circles are islands in the West Indies. Correlation analysis reveals only reptiles in Middle America are significant ($r = 0.766$, $p = <0.05$).

For any given continuous land mass, the highest densities of species are in Puerto Rico (7.5 per square km^4) and Costa Rica (7.1 per square km^4), but the actual numbers of species are vastly different—67 in Puerto Rico and 361 in Costa Rica. In comparing West Indian islands with continental areas of like size, there is a great difference in species richness. For example, Cuba (114,524 km^2) has 151 species, whereas Honduras (112,088 km^2) has 236 species. Likewise, Hispaniola (76,484 km^2) has 201 species, whereas Panama (77,096 km^2) has 350 species.

These differences would not be nearly so great if the lowlands on the mainland were excluded and the uplands were treated as islands. Even so, there is remarkable diversity in the uplands of Middle America; the highlands of nuclear Central America contain 126 species of amphibians, whereas there are 138 species in the highlands of Costa Rica and western Panama (J. A. Campbell, personal communication).

The theory of island biogeography contends that larger islands harbor more species (MacArthur and Wilson, 1963). However, this does not hold for the Greater Antillean herpetofauna. Cuba is about twice the size of Hispaniola but has only 75% as many species as Hispaniola. Jamaica is slightly larger than Puerto Rico but has only 80% as many species. A correlation analysis of numbers of species of anurans, amphibians, reptiles, and amphibians and reptiles in areas within these geographic units in the West Indies and Middle America in Table 10.1 revealed only one significant correlation ($r = 0.766$, $P < 0.05$)—area vs. number of species of reptiles in Middle America (Fig. 10.1).

As expected, because of insularity, endemism is higher in the West Indies than in Middle America. In the West Indies, 98% of the amphibians and 93% of the reptiles are endemic, whereas in Middle America, 73% of the amphibians and 65% of the reptiles are endemic. In Middle America, endemism is highest in the highlands of nuclear Central America (Chiapas, Mexico, eastward to western Honduras) and in the Central Highlands and Cordillera de Talamanca in Costa Rica and Panama.

The faunal unbalance between Middle America and the West Indies is indicative of independent histories of their biotas. Many significant elements in the Middle American herpetofauna are absent in the West Indies, including caecilians, salamanders, centrolenid, microhylid, ranid, and most genera of hylid frogs. Although the West Indian reptile fauna is diverse, it is missing most genera of iguanid and teiid lizards and colubrid snakes that occur in Middle America.

On the other hand, some genera are far more diverse in the West Indies than they are in Middle America. For example, amphisbaenians are represented by 14 species in the West Indies but only three species in Middle America and *Typhlops* by 23 in the West Indies and only three in Middle America. By far the most striking examples of speciation in the West Indies are frogs of the genus *Eleutherodactylus* and lizards of the genera *Anolis* and *Sphaerodactylus* (Fig. 10.2). Of the Middle American geographic units, Costa Rica has the highest densities of all three genera: *Eleutherodactylus*, 0.61 per square km⁴, *Anolis*, 0.51 per square km⁴, and *Sphaerodactylus*, 0.10 per square km⁴. Hispaniola has the highest numbers of these species among single islands in the West Indies:

Eleutherodactylus, 0.73 per square km⁴, *Anolis*, 0.54 per square km⁴, and *Sphaerodactylus*, 0.45 per square km⁴. The densities in Hispaniola are not that much higher than in Costa Rica.

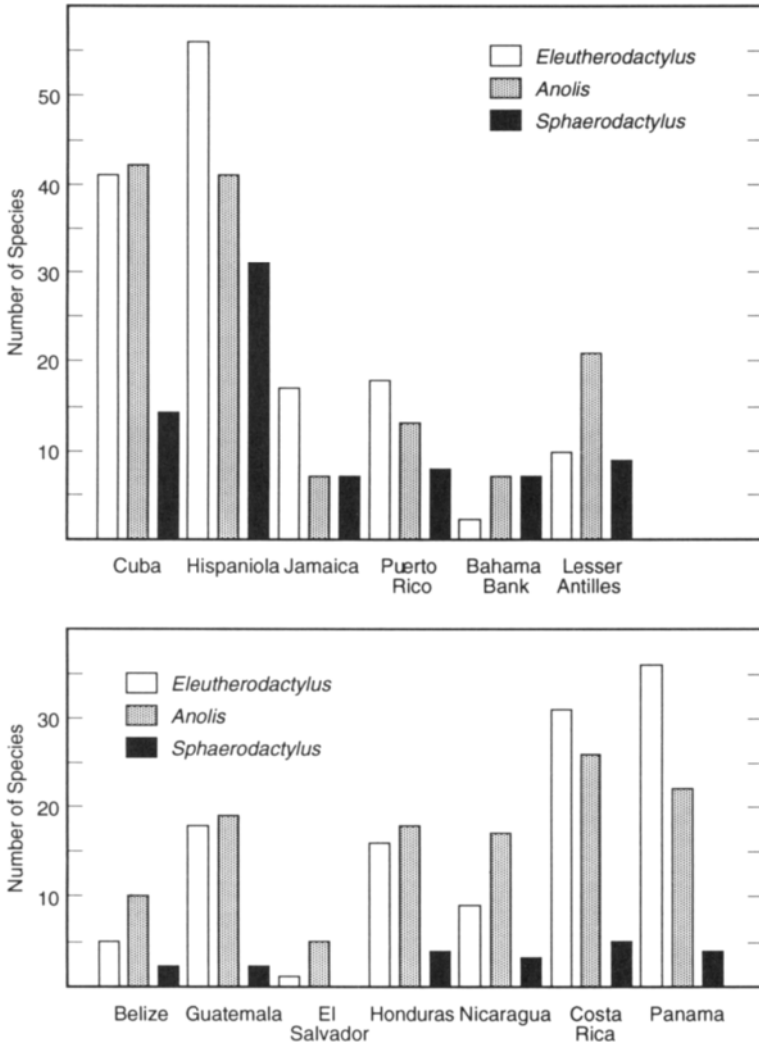


Figure 10.2. Comparative numbers of species in three genera in Middle American countries and on islands in the West Indies.

On the other hand, for the much smaller island of Puerto Rico the densities are as follows: *Eleutherodactylus*, 2.02 per square km⁴, *Anolis*, 1.46 per square km⁴, and *Sphaerodactylus*, 0.80 per square km⁴. The highest densities—*Eleutherodactylus* (1.40 per square km⁴), *Anolis* (2.93 per square km⁴), and *Sphaerodactylus* (1.26 per square km⁴)—for a geographic unit are in the Lesser Antilles, where many species are endemic to one of many islands.

Discussion

The patterns of distribution and speciation in the West Indian herpetofauna involve far fewer lineages than those in Middle America. Has the absence of certain lineages “permitted” the great diversity of some lineages (e.g., *Eleutherodactylus*, *Anolis*, and *Sphaerodactylus*) in the West Indies? Unfortunately, few comparative ecological data are available. Most ecological studies in the West Indies have involved acquisition of data to demonstrate potential interspecific competition (e.g., Schoener, 1968; Schoener and Gorman, 1968), although Williams (1969), Schoener and Schoener (1971a,b), and Roughgarden (1995) presented data on habitat utilization by *Anolis*. Most studies on Middle American anoles have dealt with single species (e.g., Andrews, 1971, on *A. polylepis*; Fitch 1972, on *A. tropidolepis*).

On the other hand, in the West Indies there are no studies comparable to the thorough investigations of communities, such as at Barro Colorado Island, Panama (Rand and Myers, 1990) and La Selva, Costa Rica (Guyer, 1990), which were compared by Duellman (1990). Because the total numbers of species of amphibians and reptiles at these two sites (133 and 134 species, respectively) approach the total for the entire island of Cuba (150), it seems likely that communities at given sites in the West Indies will be significantly smaller than on the mainland of Middle America or South America.

Knowledge of the Middle American and West Indian herpetofaunas is approaching puberty. It will reach maturity only when the historical geology is clarified precisely, when the fauna is known taxonomically, when biogeographic patterns have been established on the basis of congruent cladograms of unrelated taxa, and when comparable ecological studies reveal the composition, patterns of resource utilization, and reproductive biology within communities. I hope that the foregoing essay will provide inspiration and initiative for workers to undertake these tasks so that sometime in the not too distant future a more precise comparison of two fascinating herpetofaunas will be possible.

This Page Intentionally Left Blank

References

- Adams, W. E., K. W. Pickersgill, and G. Underwood. 1957. The development of the carotid arch in *Sphaerodactylus argus* Gosse, with some remarks on "Inselbildungen" in the aortic arches. *J. Morphol.* 121: 399-423.
- Adler, K. 1989. Herpetologists of the Past, pp. 5-125 in *Contributions to the History of Herpetology*, edited by K. Adler. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.
- Adolph, S. C., and J. Roughgarden. 1983. Foraging by passerine birds and *Anolis* lizards on St. Eustatius (Neth. Antilles): implications for interclass competition and predation. *Oecologia* 56: 313-317.
- Ahrenfeldt, R. H. 1953. Two British anatomical studies on American reptiles (1650-1750) – I. Hans Sloane: comparative anatomy of the American crocodile. *Herpetologica* 9: 79-86.
- Ahrenfeldt, R. H. 1954. Identification of the Amphibia and Reptilia recorded in Jamaica by Hans Sloane (1688-89). *Copeia* 1954: 105-111.
- Amaral, A. 1925. On the differentiation of the species *Bothrops atrox* (Linné, 1758), *B. jararaca* (Wied, 1824), and *B. jararacussu* Lacerda, 1884. *Contrib. Harvard Inst. Trop. Biol. Med.* 2: 22-43.
- Amaral, A. 1954. Contribuição ao conhecimento dos ofídios neotrópicos XXXV. A propósito de revalidação de *Coluber lanceolatus* Lacépède, 1789. *Mem. Inst. Butantan* 26: 207-214.
- Anderson, K. 1996. A karyological perspective on the monophyly of the hylid genus *Osteopilus*, pp. 157-168 in *Contributions to West Indian Herpetology: A Tribute to Albert Schwartz*, edited by R. Powell, and R. W. Henderson. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.
- Andrews, E. A. 1892. Notes on the fauna of Jamaica. *Johns Hopkins Univ. Circ.* 97: 72-77.
- Andrews, R. 1971. Structural habitat and time budget of a tropical *Anolis* lizard. *Ecology* 52: 262-270.

- Andrews, R. 1976. Growth rate in island and mainland anoline lizards. *Copeia* 1976: 477-482.
- Andrews, R. 1979. Evolution of life histories: a comparison of *Anolis* lizards from matched island and mainland habitats. *Breviora* 454: 1-51.
- Anonymous. 1943. Notes from members. *Nat. Hist. Notes Nat. Hist. Soc. Jamaica* 1: 7.
- Anonymous. 1950. Corrections and comments. *Nat. Hist. Notes Nat. Hist. Soc. Jamaica* 4: 66.
- Anonymous. 1967. An annotated checklist of the Amphibia of Jamaica. *Caribbean Biol. Center Newsletter* 2: 15-16.
- Anonymous. 1978. Opinion 1104/relative precedence of *Cornufer* Tschudi, 1838, and *Platymantis* Günther, 1858 (Amphibia Salientia). *Bull. Zool. Nomencl.* 34: 222-233.
- Anonymous. 1989. Biggest little islands in the hemisphere. *The Nature Conservancy* 39: 32.
- Arnold, D. L. 1980. Geographic variation in *Anolis brevirostris* (Sauria: Iguanidae) in Hispaniola. *Breviora* 461: 1-31.
- Asprey, G. F., and R. G. Robbins. 1953. The vegetation of Jamaica. *Ecol. Monogr.* 23: 359-412.
- Auffenberg, W. 1958. A small fossil herpetofauna from Barbuda, Leeward Islands, with the description of a new species of *Hyla*. *Q. J. Florida Acad. Sci.* 21: 248-254.
- Auffenberg, W. 1967. Notes on West Indian tortoises. *Herpetologica* 31: 34-44.
- Avila-Pires, T. C. S. 1995. Lizards of Brazilian Amazonia (Reptilia: Squamata). *Zool. Verhand.* 299.
- Bailey, J. R. 1937. A review of some recent *Tropidophis* material. *Proc. New England Zool. Club* 16: 41-52.

- Barbour, T. 1910. Notes on the herpetology of Jamaica. Bull. Museum Comp. Zool 52: 273-301.
- Barbour, T. 1914. A contribution to the zoogeography of the West Indies, with especial reference to amphibians and reptiles. Mem. Museum of Comp. Zool. 44: 205-359.
- Barbour, T. 1915. Recent notes regarding West Indian reptiles and amphibians. Proc. Biol. Soc. Washington 27: 71-78.
- Barbour, T. 1916. Additional notes on West Indian reptiles and amphibians. Proc. Biol. Soc. Washington 29: 215-220.
- Barbour, T. 1917. A new Antillean *Sphaerodactylus*. Proc. Biol. Soc. Washington 30: 163-164.
- Barbour, T. 1921. *Sphaerodactylus*. Mem. Museum Comp. Zool. 47: 217-278.
- Barbour, T. 1922. An annotated list of the Amphibia and Reptilia of Jamaica, pp. 666-669 in Handbook of Jamaica for 1922.
- Barbour, T. 1923. West Indian investigations of 1922. Occ. Pap. Museum Zool. Univ. Michigan 132: 1-9.
- Barbour, T. 1925. New Neotropical lizards. Proc. Biol. Soc. Washington 38: 101-102.
- Barbour, T. 1930a. A list of Antillean reptiles and amphibians. Zoologica (New York) 11: 61-116.
- Barbour, T. 1930b. Some faunistic changes in the Lesser Antilles. Proc. New England Zool. Club 11: 73-85.
- Barbour, T. 1933. Notes on *Scolecosaurus*. Copeia 1933: 74-77.
- Barbour, T. 1935. A second list of Antillean reptiles and amphibians. Zoologica 19: 75-141.
- Barbour, T. 1937. Third list of Antillean reptiles and amphibians. Bull. Museum Comp. Zool. 82: 73-166.
- Barbour, T. 1942. Two preoccupied names. Copeia 1942: 179.

- Barbour, T., and A. F. Carr. 1940. Antillean terrapins. Mem. Museum Comp. Zool. 54: 381-415.
- Barbour, T., and G. K. Noble. 1915. A revision of the lizards of the genus *Ameiva*. Bull. Museum Comp. Zool. 59: 417-479.
- Barbour, T., and G. K. Noble. 1916. A revision of the lizards of the genus *Cyclura*. Bull. Museum Comp. Zool. 60: 139-164.
- Barbour, T., and G. K. Noble. 1920. Some amphibians from northwestern Peru, with revision of the genera *Phyllobates* and *Telmatobius*. Bull. Museum Comp. Zool. 63: 304-427.
- Barbour, T., and C. T. Ramsden. 1919. The herpetology of Cuba. Mem. Museum Comp. Zool. 47: 71-213.
- Barbour, T., and B. J. Shreve. 1937. Novitates cubanae. Bull. Museum Comp. Zool. 80: 377-387.
- Baskin, J. N., and E. E. Williams. 1966. The Lesser Antillean *Ameiva* (Sauria, Teiidae). Re-evaluation, zoogeography, and effects of predation. Studies on the Fauna of Curacao and other Caribbean Islands 23: 144-177.
- Bauer, A. M., and A. P. Russell. 1993. *Aristelliger*. Cat. Am. Amphib. Rept. 565: 1-565.4.
- Bell, H. J. 1889. Obeah; Witchcraft in the West Indies. Sampson Low, Marston, Searle and Rivington.
- Bennett, A. F., and G. C. Gorman. 1979. Population density and energetics of lizards on a tropical island. *Oecologia* 42: 339-358.
- Bezy, R. L. 1972. Karyotypic variation and evolution of the lizards in the family Xantusiidae. *Contr. Sci. Nat. Hist. Museum Los Angeles Co.* 227: 1-29.
- Bezy, R. L., and J. A. Peterson. 1988. The microstructure of scale surfaces in the xantusiid lizard genus *Lepidophyma*. *Herpetologica* 44: 281-289.
- Birdsey, R. A., and P. L. Weaver. 1987. Forest area trends in Puerto Rico. U.S. Forest Service Res. Note 331: 1-13.

- Blake, J. A. 1986. Complex chromosomal variation in natural populations of the Jamaican lizard *Anolis grahami*. *Genetica* 69: 3-17.
- Bocourt, M.-F. 1870. Description de quelques sauriens nouveaux originaires de l'Amérique méridionale. *Nouv. Arch. Museum Hist. Nat. Paris* 6: 11-18.
- Bocourt, M.-F. 1870-1900. Etudes sur les reptiles et les batraciens. *Miss. Sci. Mex. et Am. cent.* 3: 1-1012.
- Bogart, J. P. 1974. A karyosystematic study of frogs in the genus *Leptodactylus* (Anura: Leptodactylidae). *Copeia* 1974: 728-737.
- Bogart, J. P. 1981. Chromosome studies in *Sminthillus* from Cuba and *Eleutherodactylus* from Cuba and Puerto Rico (Anura: Leptodactylidae). *Life Sci. Contrib., Royal Ontario Museum* 129: 1-22.
- Bogart, J. P., and S. B. Hedges. 1995. Rapid chromosome evolution in Jamaican frogs of the genus *Eleutherodactylus* (Leptodactylidae). *J. Zool. London* 235: 9-31.
- Bogert, C. M. 1968. The variations and affinities of the dwarf boas of the genus *Ungaliophis*. *Novitates* 2340: 1-26.
- Böhme, W. 1984. Erstfund eines fossilen Kugelfingergeckos (Sauria: Gekkonidae) aus dominikanischem Bernstein (Oligozän von Hispaniola, Antillen). *Salamandra* 20: 212-220.
- Bonnaterre, P.-J. 1789. *Erpétologie*, in *Encyclopédie Méthodique. Tableau Encyclopédique et Méthodique de Trois Règnes de la Nature*. Panckoucke, Paris.
- Boulenger, G. A. 1882. Catalogue of the Batrachia Gradientia s. Caudata and Batrachia Apoda in the Collection of the British Museum, Trustees of the British Museum, London.
- Boulenger, G. A. 1883-1896. Catalogue of the Snakes in the British Museum (Natural History). Trustees of the British Museum, London.
- Boulenger, G. A. 1885. Catalogue of the Specimens of Lizards in the Collection of the British Museum (Natural History). Trustees of the British Museum, London.

- Boulenger, G. A. 1887. Reptilia and Batrachia. Zool. Rec. 24: 1-34.
- Boulenger, G. A. 1889. Catalogue of the Chelonians, Rhychocephalians, and Crocodiles in the British Museum (Natural History). Trustees of the British Museum, London.
- Boulenger, G. A. 1891. On reptiles, batrachians, and fishes from the lesser West Indies. Proc. Zool. Soc. London 1891: 351-357.
- Boulenger, G. A. 1893. Catalogue of the snakes of the British Museum (Natural History). Volume I. Trustees of the British Museum, London.
- Boulenger, G. A. 1894. Catalogue of the snakes of the British Museum (Natural History). Volume II. Trustees of the British Museum, London.
- Boulenger, G. A. 1906. Reptiles and Batrachians, pp. 517-531 in The History of the Collections contained in the Natural History Departments of the British Museum. Trustees of the British Museum, London.
- Bourgeois, R. W. 1995. Herpetological publications of Albert Schwartz, pp. 21-27 in Contributions to West Indian Herpetology: A Tribute to Albert Schwartz, edited by R. Powell, and R. W. Henderson. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.
- Bouysee, P. 1984. The Lesser Antilles island arc: structure and geodynamic evolution. Initial Reports, Deep Sea Drilling Project, 78A: 83-103. Government Printing Office, Washington DC., U.S.
- Bowersox, S. R., S. Calderón, G. Cisneros, R. S. Garcia, C. Huntington A. Lathrop, L. Lenart, J. S. Parmerlee, Jr., R. Powell, A. Queral, D. D. Smith, S. P. Sowell, and K. C. Zippel. 1994a. Miscellaneous natural history notes on amphibians and reptiles from the Dominican Republic. Bull. Chicago Herpetol. Soc. 29: 54-55.
- Bowersox, S. R., S. Calderón, R. Powell, J. J. Parmerlee, D. D. Smith, and A. Lathrop. 1994b. Nahrung eines Riesenanolis, *Anolis barahonae*, von Hispaniola, mit einer Zusammenfassung des Nahrungsspektrums westindischer Riesenanolis-Arten. Salamandra 30: 155-160.
- Bowin, C. 1975. The geology of Hispaniola, pp. 501-522 in Ocean Basins and Margins. Vol. 3, Gulf Coast, Mexico, and the Caribbean, edited by A. E. M. Nairn, and F. G. Stehli. Plenum, New York.

- Braker, H. E., and H. W. Greene. 1994. Population biology: life histories, abundance, demography, and predator-prey interactions, pp. 244-255 in *La Selva: Ecology and Natural History of a Neotropical Rain Forest*, edited by L. A. McDade, S. Bawa, H. A. Hespdenheide and G. S. Hartshorn. Univ. Chicago Press, Chicago.
- Brongersma, L. D. 1937. The types of *Psammophis antillensis* Schlegel. *Zool. Meded.* 20: 1-5.
- Brooks, D. R. 1981. Hennig's parasitological method: a proposed solution. *Syst. Zool.* 30: 229-249.
- Brooks, D. R., and D. A. McLennan. 1991. *Phylogeny, Ecology and Behavior: a Research Program*. Univ. of Chicago Press, Chicago.
- Brooks, D. R. and R. T. O'Grady. 1989. Crocodylians and their helminth parasites: macroevolutionary considerations. *Am. Zool.* 29:873-883.
- Brooks, E. S. J. 1954. *Sir Hans Sloane. The Great Collector and His Circle*. Batworth Press, London.
- Brooks, G. R., Jr. 1982. An analysis of prey consumed by the anuran, *Leptodactylus fallax*, from Dominica, West Indies. *Biotropica* 24: 209-214.
- Brooks, G. R. 1983. *Gymnophthalmus pleei* Bocourt: an addition to the lizard fauna of Dominica, West Indies. *Herpetol. Review* 14: 31-32.
- Browne, P. 1789. *The Civil and Natural History of Jamaica with Complete Linnaean Indices*, London.
- Buden, D. W. 1966. An evaluation of Jamaican *Dromicus* (Serpentes, Colubridae) with the description of a new species. *Breviora* 238: 1-10.
- Buide, M. S. 1967. Lista de los anfidios y reptiles de cuba. *Torreia* 1:60.
- Bullock, D. J., and P. G. H. Evans. 1990. The distribution, density and biomass of terrestrial reptiles in Dominica, West Indies. *J. Zool. London* 222: 421-443.
- Bullock, D. J., and H. M. Jury. 1990. A note on the feeding habits of *Ameiva fuscata* from Dominica, Lesser Antilles. *Herpetol. J.* 1: 532-534.

- Bullock, D. J., H. M. Jury, and P. G. H. Evans. 1993. Foraging ecology in the lizard *Anolis oculatus* (Iguanidae) from Dominica, West Indies. *J. Zool. London* 230: 19-30.
- Burke, K. 1988. Tectonic evolution of the Caribbean. *Annu. Rev. Earth Planet. Sci.* 16: 201-230.
- Burnell, K. L., and S. B. Hedges. 1990. Relationships of West Indian *Anolis* (Sauria: Iguanidae): An approach using slow-evolving protein loci. *Carib. J. Sci.* 26: 7-30.
- Burney, D. A. 1993. Recent animal extinctions: recipes for disaster. *Am. Sci.* 81: 530-541.
- Burney, D. A., L. P. Burney, and R. D. E. MacPhee. 1994. Holocene charcoal stratigraphy from Laguna Tortuguero, Puerto Rico, and the timing of human arrival on the island. *J. Archaeol. Sci.* 21:273-281.
- Burns, J. K., C. A. Cunningham, R. A. Dupuis, M. N. Trask, J. S. Tulloch, R. Powell, J. S. Parmerlee, Jr., K. L. Kopecky, and M. L. Jolley. 1992. Lizards of the Cayos Siete Hermanos, Dominican Republic, Hispaniola. *Bull. Chicago Herpetol. Soc.* 27: 225-232.
- Buskirk, R. E. 1985. Zoogeographic patterns and tectonic history of Jamaica and the northern Caribbean. *J. Biogeogr.* 12: 445-461.
- Buth, D. G. 1984. The application of electrophoretic data in systematic studies. *Annu. Rev. Ecol. Syst.* 15: 501-522.
- Cadle, J. E. 1984a. Molecular systematics of neotropical xenodontine snakes: I. South American xenodontines. *Herpetologica* 40: 8-20.
- Cadle, J. E. 1984b. Molecular systematics of neotropical xenodontine snakes: II. Central American xenodontines. *Herpetologica* 40: 21-30.
- Cadle, J. E. 1984c. Molecular systematics of neotropical xenodontine snakes. III. Overview of xenodontine phylogeny and history of New World snakes. *Copeia* 1984: 641-652.
- Cadle, J. E. 1985. The neotropical colubrid snake fauna (Serpentes: Colubridae): lineage components and biogeography. *Syst. Zool.* 34: 1-20.

- Cadle, J. E. 1988. Phylogenetic relationships among advanced snakes: A molecular perspective. Univ. California Publ. Zool. 119: i-x, 1-77.
- Cadle, J. E., and H. W. Greene. 1993. Phylogenetic patterns, biogeography, and ecological structure of neotropical snake assemblages, pp. 281-293 in Species Diversity in Ecological Communities: Historical and Geographical Perspectives, edited by R. E. Ricklefs, and D. Schleter. Univ. Chicago Press, Chicago.
- Campbell, H. W. 1972. Ecological or phylogenetic interpretations of crocodylian nesting habits. Nature 238: 404-405.
- Campbell, J. A. 1994. A new lizard of the genus *Diploglossus* (Anguidae: Diploglossinae) from Mexico, with a review of Mexican and northern Central American species. Herpetologica 50: 193-209.
- Campbell, J. A., and J. E. D. Brodie. 1988. A new colubrid snake of the genus *Adelphicos* from Guatemala. Herpetologica 44: 416-422.
- Campbell, J. A., and L. S. Ford. 1982. Phylogenetic relationships of the colubrid snakes of the genus *Adelphicos* in the highlands of Middle America. Occas. Pap. Museum Nat. Hist. Univ. Kansas 100: 1-22.
- Campbell, J. A., and D. R. Frost. 1993. Anguid lizards of the genus *Abronia*: revisionary notes, descriptions of four new species, and phylogenetic analysis, and key. Bull. Am. Museum Nat. Hist. 216: 1-121.
- Campbell, J. A., and E. N. Smith. 1992. A new frog of the genus *Ptychohyala* (Hylidae) from the Sierra de Santa Cruz, Guatemala, and description of a new genus of Middle American stream-breeding treefrogs. Herpetologica 48: 153-167.
- Campbell, J. A., and J. P. Vannini. 1989. Distribution of amphibians and reptiles in Guatemala and Belize. Proc. West. Found. Vert. Zool. 4: 1-21.
- Cannatella, D. C. 1986. A new genus of Bufonid (Anura) from South America, and phylogenetic relationships of the neotropical genera. Herpetologica 42: 197-205.
- Cannatella, D. C., and K. de Queiroz. 1988. Phylogenetic systematics of the anoles: is a new taxonomy warranted? Syst. Zool. 38: 57-69.

- Capote, R. P., R. E. Ricardoi, and A. V. González. 1989. 1. Vegetación Actual, 1:1,000,000. X. Flora and Vegetación. Nuevo Atlas Nacional de Cuba: Academia de Ciencias de Cuba.
- Carey, W. M. 1975. The rock iguana, *Cyclura pinguis*, on Anegada, British Virgin Islands, with notes on *Cyclura ricordi* and *Cyclura cornuta* of Hispaniola. Bull. Florida State Museum, Biol. Sci. 19: 189-233.
- Case, J. E., T. L. Holcombe, and R. G. Martin. 1984. Map of geological provinces in the Caribbean region, pp. 1-30 in The Caribbean-South American Plate Boundary and Regional Tectonics, edited by W. E. Bonini, R. B. Hargrave, and R. Shagam. Geol. Soc. Am. Mem., Boulder, CO.
- Case, S. M., and E. E. Williams. 1984. A study of a contact zone in the *Anolis distichus* complex in the central Dominican Republic. Herpetologica 40: 118-137.
- Case, S. M., and E. E. Williams. 1987. The cybotoid anoles and *Chamaelinops*: evidence of mosaic evolution. Zool. J. Linnaean Soc. 91: 325-341.
- Case, T. J. 1975. Species numbers, density compensation, and colonizing ability of lizards on islands in the Gulf of California. Ecology 56: 3-18.
- Case, T. J. 1991. Invasion resistance, species build-up and community collapse in metapopulation models with interspecies competition. Biol. J. Linnean Soc. 42: 239-266.
- Case, T. J., and D. T. Bolger. 1991. The role of introduced species in shaping the distribution and abundance of island reptiles. Evol. Ecol. 5: 272-290.
- Case, T. J., D. T. Bolger, and A. D. Richman. 1992. Reptilian extinctions: the last ten thousand years, pp. 91-125 in Conservation Biology: the Theory and Practice of Nature Conservation, Preservation, and Management, edited by P. L. Fiedler, and S. K. Jain. Chapman and Hall, New York.
- Case, T. J., and M. L. Cody. 1987. Testing theories of island biogeography. Am. Sci. 75: 402-410.
- Case, T. J., M. E. Gilpin, and J. M. Diamond. 1979. Overexploitation, interference competition, and excess density compensation in insular faunas. Am. Nat. 113: 843-854.

- Cei, J. M., V. Erspamer, and M. Roseghini. 1972. Biogenic amines, pp. 233-242 in *Evolution in the genus Bufo*, edited by W. F. Blair. Univ. Texas Press, Austin, TX.
- Censky, E. J. 1988. *Geochelone carbonaria* (Reptilia: Testudines) in the West Indies. *Florida Sci.* 50: 108-114.
- Censky, E. J. 1995a. Mating strategy and reproductive success in a teiid lizard, *Ameiva plei*. *Behaviour* 132: 529-557.
- Censky, E. J. 1995b. Reproduction in two Lesser Antillean populations of *Ameiva plei* (Teiidae). *J. Herpetol.* 29.
- Censky, E. J. 1996. The evolution of sexual size dimorphism in the teiid lizard *Ameiva plei*: A test of alternative hypotheses, pp. 277-289 in *Contributions to West Indian Herpetology: A Tribute to Albert Schwartz*, edited by R. Powell, and R. W. Henderson. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.
- Censky, E. J., and K. Hodge. 1997. Geographic distribution: *Ramphotyphlops braminus*. *Herpetol. Rev.* 28: 210.
- Censky, E. J., K. Hodge, and J. Dudley. 1998. Over-water dispersal of lizards due to hurricanes. *Nature* 395: 556.
- Censky, E. J., and K. Lindsay. 1997. Geographic distribution: *Gymnophthalmus underwoodi*. *Herpetol. Re.* 28: 210.
- Censky, E. J., and D. R. Paulson. 1992. Revision of the *Ameiva* (Reptilia: Teiidae) of the Anguilla Bank, West Indies. *Ann. Carnegie Museum* 61: 177-195.
- Chandler, C. R., and P. J. Tolson. 1990. Habitat use by a boid snake, *Epicrates monensis*, and its anoline prey, *Anolis cristatellus*. *J. Herpetol.* 24: 151-157.
- Cheke, A. S. 1984. Lizards of the Seychelles, pp. 331-360 in *Biogeography and Ecology of the Seychelles Islands*, edited by D. R. Stoddart. W. Junk Publ., The Hague.
- Cheng, H. Y. 1983. Predation, tail regeneration and reproduction in two sympatric geckos (*Sphaerodactylus*) in Hispaniola. 26th Annu. Meetg Soc. Stud. Amphib. Rept.: 52.

- Cisek, J. M., C. A. Cunningham, R. A. Dupuis, R. P. Seibolt, and R. Powell. 1990. *Wetmorena haetiana* (NCN). Food Habits. Herpetol. Rev. 21: 62.
- Cochran, D. M. 1923a. A new species of *Eleutherodactylus* from the Dominican Republic [sic]. Proc. Biol. Soc. Washington 36: 93-94.
- Cochran, D. M. 1923b. A new frog of the genus *Leptodactylus*. J. Washington Acad. Sci. 13: 184-185.
- Cochran, D. M. 1923c. A new *Anolis* from Haiti. J. Washington Acad. Sci. 13: 225-226.
- Cochran, D. M. 1924. Notes on the herpetological collections made by Dr. W. L. Abbott on the island of Haiti. Proc. U.S. Natl. Museum 66: 1-15.
- Cochran, D. M. 1927. A new genus of anguid lizards from Haiti. Proc. Biol. Soc. Washington 40: 91-92.
- Cochran, D. M. 1928a. A new species of *Chamaelinorops* from Haiti. Proc. Biol. Soc. Washington 41: 45-47.
- Cochran, D. M. 1928b. A new genus and species of lizard, *Hispaniolus pratensis*, from the Haitian Republic. Proc. Biol. Soc. Washington 41: 49-51.
- Cochran, D. M. 1928c. The herpetological collections made in Haiti and its adjoining islands by Walter J. Eyerdam. Proc. Biol. Soc. Washington 41: 53-59.
- Cochran, D. M. 1931a. New reptiles from Beata Island, Dominican Republic. Proc. Biol. Soc. Washington 44: 89-92.
- Cochran, D. M. 1931b. A new lizard from Haiti, *Sphaerodactylus stejnegeri*. Copeia 1931: 89-91.
- Cochran, D. M. 1932a. Two new subspecies of lizards of the genus *Leiocephalus* from Hispaniola. Proc. Biol. Soc. Washington 45: 177-182.
- Cochran, D. M. 1932b. Two new lizards from Hispaniola. Proc. Biol. Soc. Washington 45: 183-187.
- Cochran, D. M. 1932c. A new snake, *Ialtris parishii*, from the Republic of Haiti. Proc. Biol. Soc. Washington 45: 189-190.

- Cochran, D. M. 1932d. A new frog, *Eleutherodactylus wetmorei*, from the Republic of Haiti. Proc. Biol. Soc. Washington 45: 191-193.
- Cochran, D. M. 1933. A new gecko from Haiti, *Aristelliger expectatus*. Proc. Biol. Soc. Washington 46: 33-35.
- Cochran, D. M. 1934a. A new lizard, *Leiocephalus personatus lunatus*, from the Dominican Republic. Occas. Pap. Boston Soc. Nat. Hist. 8: 153-156.
- Cochran, D. M. 1934b. Herpetological collections made in Hispaniola by the Utowana Expedition, 1934. Occas. Pap. Boston Soc. Nat. Hist. 8: 163-188.
- Cochran, D. M. 1935. New reptiles and amphibians collected in Haiti by P. J. Darlington. Proc. Boston Soc. Nat. Hist. 40: 367-375.
- Cochran, D. M. 1937. A necessary change in an amphibian name. J. Washington Acad. Sci. 27: 312.
- Cochran, D. M. 1938a. A new species of frog from Haiti. Proc. Biol. Soc. Washington 51: 93-94.
- Cochran, D. M. 1938b. Reptiles and amphibians from the Lesser Antilles collected by Dr. S. T. Danforth. Proc. Biol. Soc. Washington 51: 147-156.
- Cochran, D. M. 1939. Diagnoses of three new lizards and a frog from the Dominican Republic. Proc. New England Zool. Club 18: 1-3.
- Cochran, D. M. 1941. The herpetology of Hispaniola. Bull. U.S. Nat. Museum 177: vii + 398.
- Cochran, D. M. 1961. Type specimens of reptiles and amphibians in the United States National Museum. U.S. Nat. Museum Bull. 220: xv + 291.
- Cockerell, T. D. A. 1894. A little known Jamaican naturalist, Dr. Anthony Robinson. Am. Nat. 28: 775-790.
- Cocteau, J. T., and G. Bibron. 1843. Reptiles. Hist. Fis. Pol. Nat. Cuba 4: 1-143.
- Cohen, M. M., and C. Gans. 1970. The chromosomes of the order Crocodylia. Cytogenetics 9: 81-105.

- Collette, B. B. 1961. Correlations between ecology and morphology in anoline lizards from Havana, Cuba and southern Florida. *Bull. Museum Comp. Zool.* 125: 137-162.
- Connor, E. F., and E. D. McCoy. 1979. The statistics and biology of the species-area relationship. *Am. Nat.* 113: 791-833.
- Cope, E. D. 1861. Notes and descriptions of anoles. *Proc. Acad. Nat. Sci. Philadelphia* 13: 208-215.
- Cope, E. D. 1861 (1862). On the genera *Panolopus*, *Centropyx*, *Aristelliger* and *Sphaerodactylus*. *Proc. Acad. Nat. Sci. Philadelphia* 13: 494-500.
- Cope, E. D. 1862a. Synopsis of the species of *Holcosus* and *Ameiva*, with diagnoses of new West Indian and South American Colubridae. *Proc. Acad. Nat. Sci. Philadelphia* 14: 60-82.
- Cope, E. D. 1862b. Contributions to Neotropical saurology. *Proc. Acad. Nat. Sci. Philadelphia* 14: 176-188.
- Cope, E. D. 1863a. On *Trachycephalus*, *Scaphiopus* and other American Batrachia. *Proc. Acad. Nat. Sci. Philadelphia* 15: 43-54.
- Cope, E. D. 1863b. In Errata and addenda. *Proc. Acad. Nat. Sci. Philadelphia* 14: 594.
- Cope, E. D. 1864. Contributions to the herpetology of tropical America. *Proc. Acad. Nat. Sci. Philadelphia* 16: 166-181.
- Cope, E. D. 1868. An examination of the Reptilia and Batrachia obtained by the Orton Expedition to Equador and the Upper Amazon, with notes on other species. *Proc. Acad. Nat. Sci. Philadelphia* 20: 96-140.
- Cope, E. D. 1869. Seventh contribution to the herpetology of tropical America. *Proc. Am. Philos. Soc.* 11: 147-169.
- Cope, E. D. 1871. Ninth contribution to the herpetology of tropical America. *Proc. Acad. Nat. Sci. Philadelphia* 23: 200-224.
- Cope, E. D. 1879. Eleventh contribution to the herpetology of tropical America. *Proc. Am. Philos. Soc.* 18: 261-277.

- Cope, E. D. 1885. The large iguanas of the Greater Antilles. *Am. Nat.* 19: 1005-1006.
- Cope, E. D. 1886. On the species of Iguaninae. *Proc. Am. Philos. Soc.* 23: 261-271.
- Cope, E. D. 1895. The Batrachia and Reptilia of the Univ. of Pennsylvania West Indian Expedition of 1890 and 1891. *Proc. Acad. Nat. Sci. Philadelphia* 46: 429-442.
- Corke, D. 1987. Reptile conservation on the Maria Islands (St. Lucia, West Indies). *Biol. Conserv.* 40: 263-279.
- Corke, D. 1992. The status and conservation needs of the terrestrial herpetofauna of the Windward Islands (West Indies). *Biol. Conserv.* 62: 47-58.
- Cousens, P. N. 1956. Notes on the Jamaican and Cayman Island lizards of the genus *Celestus*. *Breviora* 56: 1-6.
- Croizat, L. 1958. Panbiogeography. Published by the author. Caracas, Venezuela.
- Croizat, L. 1964. Space, Time and Form: the biological synthesis. Published by the author, Caracas, Venezuela.
- Croizat, L., G. Nelson, and D. E. Rosen. 1974. Centers of origin and related concepts. *Syst. Zool.* 23: 265-287.
- Crombie, R. I. 1973. Comment on the proposed suppression of *Hyla crucialis* (Amphibia). *Z.N.(S.)* 1982. *Bull. Zool. Nomencl.* 30: 4-6.
- Crombie, R. I. 1974. The ecology, behavior and systematics of Jamaican hylid frogs. *Yearbook Am. Philos. Soc.* 1973: 304-305.
- Crombie, R. I. 1977. A new species of frog of the genus *Eleutherodactylus* (Amphibia: Leptodactylidae) from the Cockpit Country of Jamaica. *Proc. Biol. Soc. Washington* 90: 194-204.
- Crombie, R. I. 1985. Another new forest-dwelling frog (Leptodactylidae: *Eleutherodactylus*) from the Cockpit Country of Jamaica. *Trans. San Diego Soc. Nat. Hist.* 21: 145-153.

- Crombie, R. I., D. W. Steadman, and J. C. Barber. 1984. A preliminary survey of the vertebrates of Cabarita Island, St. Mary Parish, Jamaica. *Atoll. Res. Bull.* 280: 1-12.
- Crother, B. I. 1990. Is 'some better than none' or do allele frequencies contain phylogenetically useful information. *Cladistics* 6(2): 277-281.
- Crother, B. I. 1999. Phylogenetic relationships among West Indian xenodontine snakes (Serpentes; Colubridae) with comments on the phylogeny of some mainland xenodontines. *Contemporary Herpetology*.
- Crother, B. I., J. A. Campbell, and D. M. Hillis. 1992. Phylogeny and historical biogeography of the palm-pitvipers, genus *Bothriechis*: biochemical and morphological evidence, pp. 1-20 in *Biology of the Pitvipers*, edited by J. A. Campbell, and J. E. D. Brodie. Selva, Tyler. TX.
- Crother, B. I., and C. Guyer. 1996. Caribbean historical biogeography: was the dispersal-vicariance debate eliminated by an extraterrestrial bolide? *Herpetologica* 52: 440-465.
- Crother, B. I., and D. M. Hillis. 1995. Nuclear ribosomal DNA restriction sites, phylogenetic information, and the phylogeny of some xenodontine (Colubridae) snakes. *J. Herpetol.* 29:316-320.
- Crother, B. I., M. M. Miyamoto, and W. F. Presch. 1986. Phylogeny and biogeography of the lizard family Xantusiidae. *Syst. Zool.* 35: 37-45.
- Crother, B. I., and W. F. Presch. 1992. The phylogeny of xantusiid lizards: The concern for analysis in the search for a best estimate of phylogeny. *Mol. Phylogenet. Evol.* 1: 289-294.
- Crother, B. I., and W. Presch. 1994. Xantusiid lizards, concern for analysis, and the search for a best estimate of phylogeny: further comments. *Mol. Phylo. Evol.* (3)3: 272-275.
- Crozier, R. H. 1992. Genetic diversity and the agony of choice. *Biol. Conserv.* 61: 11-15.
- Crozier, R. H. 1997. Preserving the information content of species: genetic diversity, phylogeny, and conservation worth. *Annu. Rev. Ecol. Syst.* 28: 243-68.

- Cuello, J. P. 1988. Lista de las aves fósiles de al región neotropical y de las islas Antillanas. *Paula-Coutianas Porto alegre*. 2: 3-79.
- Cullen, D. J., and R. Powell. 1994. A comparison of food habits of a montane and lowland population of *Anolis distichus* (Lacertilia: Polychrotidae) from the Dominican Republic. *Bull. Maryland Herpetol. Soc.* 30: 62-66.
- Cullom, S. J., and A. Schwartz. 1980. Variation in the Hispaniolan giant anole *Anolis barahonae* with the description of two new subspecies. *Herpetologica* 36: 93-98.
- Cundall, D., V. Wallach, and D. A. Rossman. 1993. The systematic relationships of the snake genus *Anomochilus*. *Zool. J. Linnean Soc.* 109: 275-299.
- Cunningham, C. A., R. Powell, and S. B. Hedges. 1998. *Eleutherodactylus chlorophenax*. *Cat. Am. Amphib. Rept.* 648: 1-2.
- Cunningham, C. A., J. S. Tulloch, and R. Powell. 1993. Food habits of four Hispaniolan species of *Sphaerodactylus* (Lacertilia: Gekkonidae). *Herpetol. Nat. Hist.* 1: 91-93.
- Curtis, A. 1947. Prevalence of snakes in Haiti. *Herpetologica* 3: 224.
- Cuvier, G. 1807. Sur les différentes espèces de crocodiles vivans et sur leurs caractères distinctifs. *Annu. Natl. Museum Hist. Nat. Paris* 10: 8-66.
- Darlington, P. J. 1938. The origin of the fauna of the Greater Antilles, with discussion of dispersal of animals over water and through the air. *Q. Rev. Biol.* 13: 274-300.
- Darlington, P. J. 1957. *Zoogeography: The Geographical Distribution of Animals*. J. Wiley & Sons, Inc., New York.
- Darwin, S. P., and A. L. Welden (Editors). 1992. *Biogeography of Mesoamerica*, Special Publ. No. 1. Tulane Studies Zool. Bot., New Orleans, LA.
- Daudin, F.-M. 1802a. *Histoire Naturelle, Générale et Particuliere, des Reptiles*. L'Imprimerie de F. Dufart, Paris.
- Daudin, F.-M. 1802b. *Histoire Naturelle des Rainettes, des Grenouilles et des Crapauds*. Lerrault, Paris.

- Daudin, F.-M. 1803. *Histoire Naturelle des Reptiles*. Lerrault, Paris.
- Davis, D. D. 1982. Archaic settlement and resource exploitation in the Lesser Antilles: preliminary information from Antigua. *Carib. J. Sci.* 17: 107-122.
- de Beer, G. R. 1953. *Sir Hans Sloane and the British Museum*. Oxford Univ. Press, New York.
- de Queiroz, K. 1987. Phylogenetic systematics of iguanine lizards: a comparative osteological study. *Univ. California Publ. Zool.* 118: 1-203.
- Densmore, L. D. 1983. Biochemical and immunological systematics of the order Crocodylia. pp397-465 in *Evolutionary Biology*, Vol. 16, edited by M. K. Hecht, B. Wallace, and G. H. Prance. Plenum, New York.
- Densmore, L. D., and R. D. Owen. 1989. Molecular systematics of the order Crocodylia. *Am. Zool.* 29: 831-841.
- Densmore, L. D., and P. S. White. 1991. The systematics and evolution of the Crocodylia as suggested by restriction endonuclease analysis of mitochondrial and nuclear ribosomal DNA. *Copeia* 1991: 602-615.
- Dessauer, H. C., J. E. Cadle, and R. Lawson. 1987. Patterns of snake evolution suggested by their proteins. *Fieldiana Zool.* 1376: 1-34.
- Dial, R., J. Roughgarden, and S. C. Tobin. 1994. Notes on the absolute abundance of canopy anoles, *Anolis cuvieri*, *A. stratulus*, and *A. evermanni* (Lacertilia: Polychridae) in the Luquillo Forest, Puerto Rico. *Carib. J. Sci.* 30: 278-279.
- Diamond, J. 1975. Assembly of species communities, pp. 342-444 in *Ecology and Evolution of Communities*, edited by M. L. Cody and J. Diamond. Harvard Univ. Press, Cambridge.
- Diaz, L. M., A. R. Estrada, and L. V. Moreno. 1996. A new species of *Anolis* (Sauria: Iguanidae) from the Sierra de Trinidad, Sancti Spiritu, Cuba. *Carib. J. Sci.* 32(1): 54-58.
- Dickerson, M. C. 1916. Description of a new amphisbaenian collected by the late Charles S. Mead in 1911, on the Isle of Pines, Cuba. *Bull. Am. Museum Nat. Hist.* 35: 659-662.

- Diesel, R., G. Baurle, and P. Vogel. 1995. Cave breeding and froglet transport: a novel pattern of anuran brood care in the Jamaican frog, *Eleutherodactylus cundalli*. *Copeia* 2: 354-360.
- Dixon, J. R. 1962. The leaf-toed geckos, genus *Phyllodactylus*, of northeastern South America. *Southwest. Nat.* 7: 211-226.
- Dixon, J. R. 1973a. *Phyllodactylus*. *Cat. Am. Amphib. Rept.* 141: 1-2.
- Dixon, J. R. 1973b. A systematic review of the teiid lizards, genus *Bachia*, with remarks on *Heterodactylus* and *Anotosaura*. *Univ. Kansas Museum Nat. Hist. Misc. Publ.* 57: 1-47.
- Dixon, J. R. 1980. The neotropical colubrid snake genus *Liophis*. The generic concept. *Milwaukee Publ. Museum Contr. Biol. Geol.* 31: 1-40.
- Dixon, J. R. 1981. The neotropical colubrid snake genus *Liophis*: The eastern Caribbean complex. *Copeia* 1981: 296-304.
- Dixon, J. R., and R. B. Huey. 1970. Systematics of the lizards of the gekkonid genus *Phyllodactylus* of mainland South America. *Los Angeles County Museum Contrib. Sci.* 192: 1-78.
- Dixon, J. R., and P. Soini. 1986. The Reptiles of the Upper Amazon Basin, Iquitos Region, Peru. *Milwaukee Public Museum, Milwaukee, WI.*
- Dobson, A. P., S. V. Pacala, J. D. Roughgarden, E. R. Carper, and E. A. Harris. 1992. The parasites of *Anolis* lizards in the northwestern Lesser Antilles. I. Patterns of distribution and abundance. *Oecologia* 91: 110-117.
- Dodd, C. K., and R. Franz. 1996. Species richness and biogeography of the herpetofauna in the Exuma Cays Land and Sea Park, Bahamas, pp. 175-194 in *Contributions to West Indian Herpetology: A Tribute to Albert Schwartz*, edited by R. Powell, and R. W. Henderson. *Society for the Study of Amphibians and Reptiles Contributions in Herpetology*, Ithaca, NY.
- Donnelly, M. A., and C. Guyer. 1994. Patterns of reproduction and habitat use in an assemblage of neotropical frogs. *Oecologia* 98(3-4): 291-302.
- Donnelly, T. W. 1989. History of marine barriers and terrestrial connections: Caribbean paleogeographic inference from pelagic sediment analysis, pp. 103-118 in *Biogeography of the West Indies: Past, Present, and Future*, edited by C. A. Woods. *Sandhill Crane Press, Gainesville.*

- Dowling, H. G., R. Highton, G. C. Maha, and L. R. Maxson. 1983. Biochemical evaluation of colubrid snake phylogeny. *J. Zool. London* 201: 309-329.
- Dowling, H. G., C. A. Hass, S. B. Hedges, and R. Highton. 1996. Snake relationships revealed by slow-evolving proteins: a preliminary survey. *J. Zool. London* 240: 1-28.
- Downer, A., and R. Sutton. 1990. *Birds of Jamaica - A Photographic Field Guide*. Cambridge Univ. Press., Cambridge, UK.
- Drewry, G. E., and K. Jones. 1976. A new ovoviviparous frog, *Eleutherodactylus jasper* (Amphibia, Anura, Leptodactylidae) from Puerto Rico. *J. Herpetol.* 10: 161-165.
- Duellman, W. E. 1970. The hylid frogs of Middle America. *Monogr. Museum Nat. Hist. Univ. Kansas* 1: 1-748.
- Duellman, W. E. 1978. The biology of an equatorial herpetofauna in Amazonian Ecuador. *Misc. Publ. Museum Nat. Hist., Univ. Kansas* 65: 1-352.
- Duellman, W. E. 1987. Lizards in an Amazonian rain forest community: resource utilization and abundance. *Nat. Geog. Res.* 3: 489-500.
- Duellman, W. E. 1989. Tropical herpetofauna communities: patterns of community structure in neotropical rainforests, pp. 61-88 in *Vertebrates in Complex Tropical Systems*, edited by M. L. Harmelin-Vivien, and F. Bourlière. Springer-Verlag, New York.
- Duellman, W. E. 1990. Herpetofaunas in neotropical rainforests: comparative composition, history, and resource use, pp. 455-505 in *Four Neotropical Rainforests*, edited by A. H. Gentry. Yale Univ. Press, New Haven, CT.
- Duellman, W. E. 1993. Amphibian species of the world: additions and corrections. *Univ. Kansas Museum Nat. Hist. Spec. Publ.* 21: 1-372.
- Duellman, W. E., and J. A. Campbell. 1992. Hylid frogs of the genus *Plectrohyla*: systematics and phylogenetic relationships. *Misc. Publ. Museum Zool. Univ. Michigan* 181: 1-32.

- Duellman, W. E., and J. R. Mendelson, III. 1995. Amphibians and reptiles from northern Departamento Loreto, Peru: taxonomy and biogeography. Univ. Kansas Sci. Bull. 55: 329-376.
- Duellman, W. E., and L. Trueb. 1986. Biology of Amphibians. McGraw-Hill, New York.
- Duer, C. K., J. M. Cisek, and R. Powell. 1992. Food habits of *Osteopilus dominicensis* (Anura: Hylidae). Carib. J. Sci. 28: 226-228.
- Duméril, A. M. C. 1853. Prodrome de la classification des reptiles ophidiens. Troisième ordre de la classe des reptiles. Les ophidiens. Mem. Acad. Sci. France 23: 399-535.
- Duméril, A. M. C. 1861. Reptiles et poissons de l'Afrique occidentale. Arch. Museum d'Hist. Nat. 10: 137-240.
- Duméril, A. M. C., and G. Bibron. 1834. Érpétologie Générale ou Histoire Naturelle Complète des Reptiles. Imprimerie et Fonderie de Fain, Paris.
- Duméril, A. M. C., and G. Bibron. 1836. Érpétologie Générale ou Histoire Naturelle Complète des Reptiles. Vol. 3. Librairie Encyclopédique de Roret, Paris.
- Duméril, A. M. C., and G. Bibron. 1837. Érpétologie Générale ou Histoire Naturelle Complète des Reptiles. Vol. 4. Librairie Encyclopédique de Roret, Paris.
- Duméril, A. M. C., and G. Bibron. 1839. Érpétologie Générale ou Histoire Naturelle Complète des Reptiles. Vol. 5. Librairie Encyclopédique de Roret, Paris.
- Duméril, A. M. C., and G. Bibron. 1841. Érpétologie Générale ou Histoire Naturelle Complète des Reptiles. Vol. 8. Librairie Encyclopédique de Roret, Paris.
- Duméril, A. M. C., and G. Bibron. 1844. Érpétologie Générale ou Histoire Naturelle Complète des Reptiles. Librairie Encyclopédique de Roret, Paris.
- Duméril, A. M. C., and A. H. A. Duméril. 1851. Catalogue Méthodique de la Collection des Reptiles du Muséum d'Histoire Naturelle du Paris. Gide et Baudry, Paris.

- Duméril, A. M. C., G. Bibron, and A. H. A. Duméril. 1854. *Érpetologie Générale ou Histoire Naturelle Complète des Reptiles*. Vol. 7. Libraire Encyclopédique Roret, Paris.
- Dunn, E. R. 1920a. A new lizard from Haiti. *Proc. New England Zool. Club* 7: 33-34.
- Dunn, E. R. 1920b. On the Haitian snakes of the genera *Leimadophis* and *Uromacer*. *Proc. New England Zool. Club* 7: 37-44.
- Dunn, E. R. 1925. A new tree-toad from Jamaica. *Occ. Pap. Boston Soc. Nat. Hist.* 5: 161-162.
- Dunn, E. R. 1926a. The frogs of Jamaica. *Proc. Boston Soc. Nat. Hist.* 38: 111-130.
- Dunn, E. R. 1926b. Additional frogs from Cuba. *Occ. Pap. Boston Soc. Nat. Hist.* 5(2):209-215.
- Dunn, E. R. 1927. Key to Jamaican frogs and notes on their habits, pp. 532-538 in *Handbook of Jamaica*.
- Dunn, E. R. 1928. Two new frogs from Jamaica. *Proc. U.S. Natl. Museum* 74: 1-2.
- Dunn, E. R. 1929. On the trail of the snorer. *Nature* 13: 110-112.
- Dunn, E. R. 1931. The herpetological fauna of the Americas. *Copeia* 1931: 106-119.
- Dunn, E. R. 1932. The colubrid snakes of the Greater Antilles. *Copeia* 1932: 89-92.
- Dunn, E. R. 1934. Notes on *Iguana*. *Copeia* 1934: 1-4.
- Dunn, E. R. 1936. Notes on American Mabuyas. *Proc. Acad. Natl. Sci. Philadelphia* 87: 533-557.
- Dunn, E. R. 1939. Mainland forms of the snake genus *Tretanorhinus*. *Copeia* 1939: 212-217.

- Du Tertre, J. 1654. Histoire générale, des Isles de S. Christophe, de la Guade-
loupe, de la Martinique, et autres dans l'Amérique, Paris.
- ENCAMP. 1980. Survey of conservation priorities in the Lesser Antilles. Car-
ibbean Conservation Association. School of Natural Resources, Univ. Michigan.
- Eggleton, P., and D. Vane-Wright. 1994. Pattern and Process: Phylogenetic
Approaches to Ecological Problems. Academic Press, New York.
- Espeut, W. B. 1882. On the acclimitization of the Indian mungoos in Jamaica.
Proc. Zool. Soc. London 1882: 712-714.
- Estrada, A. R. 1992. Lista de anfibios y reptiles de Cayo Saetia, Costa N. de
Mayari, Holguín, Cuba. Comun. Breves Zool. 14.
- Estrada, A. R. 1993a. Herpetofauna del Archipiélago de los Canarreos, Cuba.
Poeyana 431: 1-19.
- Estrada, A. R. 1993b. Anfibios y reptiles de Cayo Coco, Archipiélago de Sa-
bana-Camagüey, Cuba. Poeyana 432: 1-21.
- Estrada, A. R. 1994. Herpetofauna de la cuenca Banao-Higuanojo, Sancti
Spiritus, Cuba. Rev. Acad. Colombiana Cien. 73.
- Estrada, A. R., and R. Alonso. 1997. Nueva especie del grupo *limbatus* (Lepto-
dactylidae: *Eleutherodactylus*) de la región oriental de Cuba. Carib. J. Sci. 33:
41-44.
- Estrada, A. R., and L. F. de Armas. 1998. Apuntes ecológicos sobre *Cri-
cosaura typica*. Carib. J. Sci, in press.
- Estrada, A. R., and O. H. Garrido. 1990. Nueva subespecie de *Anolis jubar*
(Lacertilia, Iguanidae) para Cayo Coco y la Loma de Cunagua, Ciego de Avila,
Cuba. Biología IV(1): 71-79.
- Estrada, A. R., and O. H. Garrido. 1991. Dos nuevas especies de *Anolis* (Lac-
ertilia: Iguanidae) de la Región Oriental de Cuba. Carib. J. Sci. 27(3-4): 146-
161.
- Estrada, A. R., and S. B. Hedges. 1991. Nueva especie de *Eleutherodactylus*
(Anura: Leptodactylidae) de la Región Oriental de Cuba. Carib. J. Sci. 27(3-4):
139-145.

- Estrada, A. R., and S. B. Hedges. 1996a. A new frog of the genus *Eleutherodactylus* from eastern Cuba (Anura: Leptodactylidae). *Herpetologica* 52(3): 435-439.
- Estrada, A. R., and S. B. Hedges. 1996b. A new diminutive frog of the genus *Eleutherodactylus* from Cuba (Anura, Leptodactylidae). *Copeia* 1996 (4): 852-859.
- Estrada, A. R., and S. B. Hedges. 1997a. Nueva especie de *Eleutherodactylus* (Anura: Leptodactylidae) del macizo Sagua-Baracoa, Cuba. *Carib. J. Sci.* 33(3-4): 222-226.
- Estrada, A. R., and S. B. Hedges. 1997b. A new species of frog from Meseta de Cabo Cruz, eastern Cuba (Leptodactylidae: *Eleutherodactylus*). *Carib. J. Sci.* 33(3-4): 227-236.
- Estrada, A. R., and S. B. Hedges. 1998. Sistemática de las rana ribereñas de Cuba (Leptodactylidae: *Eleutherodactylus*) con la descripción de una nueva especie. *Carib. J. Sci.*, in press.
- Estrada, A. R., and J. Novo. 1985a. A claraciones sobre la distribución de *Eleutherodactylus pinarensis* Dunn (Amphibia: Anura: Leptodactylidae) en la occidente de Cuba. *Poeyana* 302: 1-6.
- Estrada, A. R., and J. Novo. 1985b. Nueva especie de *Eleutherodactylus* del grupo *ricordi* (Anura:Leptodactylidae) del occidente de Cuba. *Poeyana* 303: 1-10.
- Estrada, A. R., and A. S. Rodriguez. 1984. Análisis de la ecomorfología de 23 especies de lagartos cubanos del género *Anolis*. *Ciencias Biológicas* 12: 91-104.
- Estrada, A. R., J. Novo, and L. V. Moreno. 1989. Redescrpción de *Eleutherodactylus symingtoni* Schwartz (Anura:Leptodactylidae) del occidente de Cuba. *Biología* III(2): 155-165.
- Etheridge, R. 1960. The relationships of the anoles (Reptilia: Sauria: Iguaniidae): an interpretation based on skeletal morphology. PhD. Dissertation, Univ. of Michigan, Ann Arbor.
- Etheridge, R. E. 1965. Fossil lizards from the Dominican Republic. *Q. J. Florida Acad. Sci.* 28: 83-105.

- Etheridge, R. E. 1966. The systematic relationships of West Indian and South American lizards referred to the iguanid genus *Leiocephalus*. *Copeia* 1966: 79-91.
- Etheridge, R. E. and K. de Queiroz. 1988. A phylogeny of iguanidae, pp. 283-368 in *Phylogenetic Relationships of the Lizard Families: Essays Commemorating Charles L. Camp*, edited by R. Estes and G. Pregill. Stanford Univ. Press, Stanford.
- Evans, L. T. 1938. Cuban field studies on territoriality of the lizard, *Anolis sagrei*. *Comp. Psychol.* 25: 97-125.
- Evans, P. 1989. Herpetofauna of the Commonwealth of Dominica (Windward I., Lesser Antilles), West Indies. *British Herpetol. Soc. Bull.* 28: 5-7.
- Ewel, J. J., and J. L. Whitmore. 1973. The ecological life zones of Puerto Rico and the U.S. Virgin Islands. *Forest Service Res. Paper* 18: 1-72.
- Faith, D. P. 1992. Conservation evaluation and phylogenetic diversity. *Biol. Conserv.* 61: 1-10.
- Faith, D. P. 1994. Phylogenetic diversity: a general framework for the prediction of feature diversity. *Philos. Trans. R. Soc. London Ser. B* 345:45-48.
- Fischer, J. G. 1856. Neue Schlangen des Hamburgischen Naturhistorischen Museums. *Abh. Naturwiss. Ver. Hamburg* 3: 79-116.
- Fischer, J. G. 1883. Beschreibungen neuer Reptilien. *Oster-Progr. Akad. Gymn. Hamburg* 1883: 1-17.
- Fischer, J. G. 1888. Ueber eine Kollektion Reptilien und Amphibien von Hayti. *Jahrb. Wiss. Anst. Hamburg* 5: 23-45.
- Fitch, H. S. 1972. Ecology of *Anolis tropidolepis* in Costa Rican cloud forest. *Herpetologica* 28: 10-21.
- Fitch, H. S. 1987. The sin of anecdotal writing. *Herpetol. Rev.* 18: 68.
- Fitch, H. S., and R. W. Henderson. 1987. Ecological and ethological parameters in *Anolis bahorucoensis*, a species having rudimentary development of the dewlap. *Amphib.-Rept.* 8: 69-80.

- Fitch, H. S., R. W. Henderson, and H. Guarisco. 1989. Aspects of the ecology of an introduced anole: *Anolis cristatellus* in the Dominican Republic. *Amphib.-Rept.* 10: 307-320.
- Fitzinger, L. 1843. *Systema Reptilium. Fasciculus primus. Amblyglossae.* Braumüller et Seidel, Vindobonae.
- Fleishman, L. J., E. R. Loew, and M. Leal. 1993. Ultraviolet vision in lizards. *Nature* 365: 397.
- Flores, G. 1984. Comparative osteology, relationships, and evolution in Jamaican frogs of the genus *Eleutherodactylus*, pp. 122. Harvard Univ., Cambridge, MA.
- Flores, G., J. H. Lenzycki, and J. J. Palumbo. 1994. An ecological study of the endemic Hispaniolan anoline lizard, *Chamaelinorops barbouri* (Lacertilia: Iguanidae). *Breviora* 499: 1-23.
- Floyd, H. B., and T. A. Jenssen. 1983. Food habits of the Jamaican lizard, *Anolis opalinus*: resource partitioning and seasonal effects examined. *Copeia* 1983: 319-331.
- Fobes, T. M., R. Powell, J. J. S. Parmerlee, A. Lathrop, and D. D. Smith. 1992. Natural history of *Anolis cybotes* (Sauria: Polychridae) from an altered habitat in Barahona, Dominican Republic. *Carib. J. Sci.* 28: 200-207.
- Forcart, L. 1951. Nomenclature remarks on some generic names of the snake family Boidae. *Herpetologica* 7: 197-199.
- Forey, P. I., C. J. Humphries, and R. I. Vane-Wright. 1994. *Systematics and Conservation Evaluation.* Clarendon, Oxford, UK.
- Forsgaard, K. 1983. The axial skeleton of *Chamaelinorops*, pp. 284-295 in *Advances in Herpetology and Evolutionary Biology*, edited by A. G. J. Rhodin, and K. Miyata. *Museum Comp. Zool.*, Cambridge, MA.
- Fowler, H. W. 1918. Some amphibians and reptiles from Porto Rico and the Virgin Islands. *Publ. Carnegie Inst. Washington* 252: 1-16.
- Franz, R., and C. A. Woods. 1983. A fossil tortoise from Hispaniola. *J. Herpetol.* 17: 79-81.

- Frazer, H., S. Carrington, A. Forde, and J. Gilmore. 1990. A-Z of Barbados Heritage. Heinemann, Jamaica.
- Frost, D. R., and R. Etheridge. 1989. A phylogenetic analysis and taxonomy of iguanian lizards (Reptilia: Squamata). Univ. Kansas Museum Nat. Hist. Misc. Publ. 81: 1-65.
- Frost, D. R., and D. M. Hillis. 1990. Species in concept and practice: Herpetological applications. *Herpetologica* 46: 87-104.
- Gali, F., and A. Schwartz. 1982. A new subspecies of *Leiocephalus personatus* from the República Dominicana. *J. Herpetol.* 16: 177-179.
- Gali, F., A. Schwartz, and A. Suarez. 1988. A new subspecies of *Leiocephalus personatus* (Sauria: Iguanidae) from Haiti. *Proc. Biol. Soc. Washington* 101: 1-3.
- Gallagher, D. S., and J. R. Dixon. 1980. A new lizard (Sauria: Teiidae: *Kentropyx*) from Brazil. *Copeia* 1980: 616-620.
- Gans, C. 1964. *Amphisbaena schmidti*, a third species of the genus from Puerto Rico (Amphisbaenia: Reptilia). *Breviora* 198: 1-11.
- Gans, C. 1967. The diphyletism of the amphisbaenia (Reptilia): a re-evaluation based upon chromosome counts. *Copeia* 1967: 485-487.
- Gans, C. 1978. The characteristics and affinities of the Amphisbaenia. *Trans. Zool. Soc. London* 34:347-416.
- Gans, C. 1990. Patterns in amphisbaenian biogeography: a preliminary analysis, pp. 133-143 in *Vertebrates in the Tropics*, edited by G. Peters, and R. Hut-terer. Museum Alexander Koenig, Bonn.
- Gans, C., and A. A. Alexander. 1962. Studies on amphisbaenids (Amphisbaenia, Reptilia). 2. On the amphisbaenids of the Antilles. *Bull. Museum Comp. Zool.* 128: 67-158.
- Garman, S. 1887a. On West Indian Gekkonidæ and Anguidæ. *Bull. Essex Inst.* 19: 17-24.

- Garman, S. 1887b. On West Indian reptiles. Iguanidae. Bull. Essex Inst. 19: 25-30.
- Garman, S. 1887c. West Indian Batrachians in the Museum of Comparative Zoology. Bull. Essex Inst. 19: 13-16.
- Garman, S. 1887d. On West Indian reptiles and batrachians in the Museum of Comparative Zoology at Cambridge, Mass. Bull. Essex Inst. 19: 1-16.
- Garman, S. 1887e. On West Indian reptiles in the Museum of Comparative Zoology at Cambridge, Mass. Proc. Am. Philos. Soc. 24: 278-286.
- Garrick, L. D., R. L. Sutton, and J. W. Lang. 1985. Observations on the largest Jamaican tree frog, *Calyptahyla crucialis*. Carib. J. Sci. 21: 159-162.
- Garrido, O. H. 1972. *Anolis bremeri* Barbour (Lacertilia: Iguanidae) en el occidente de Cuba Isla de Pinos. Carib. J. Sci. 12(1-2): 59-77.
- Garrido, O. H. 1973a. Nueva especie de *Leiocephalus* (Lacertilia, Iguanidae) para Cuba. Poeyana (116): 1-19.
- Garrido, O. H. 1973b. Distribución y variación de *Anolis homolechis* Cope (Lacertilia: Iguanidae) en Cuba. Poeyana 120: 1-68.
- Garrido, O. H. 1975a. Nuevos reptiles del archipiélago cubano. Poeyana 141: 1-58.
- Garrido, O. H. 1975b. Distribución y variación de *Anolis argillaceus* Cope (Lacertilia: Iguanidae) en Cuba. Poeyana 142: 1-28.
- Garrido, O. H. 1975c. Distribución y variación del complejo *Anolis cyanopleurus* (Lacertilia: Iguanidae) en Cuba. Poeyana 143: 1-60.
- Garrido, O. H. 1975d. Variación de *Anolis argillaceus* Cope (Lacertilia: Iguanidae) en el occidente de Cuba y la Isla de Pinos. Poeyana 144: 1-18.
- Garrido, O. H. 1981. Nueva subespecie de *Anolis equestris* (Sauria: Iguanidae) para Cuba, con comentarios sobre la distribución y afinidades de otras poblaciones del complejo. Poeyana 232: 1-15.

- Garrido, O. H. 1982a. Descripción de una nueva especie cubana de *Chamaeleolis* (Lacertilia: Iguanidae), con notas sobre su comportamiento. *Poeyana* 236: 1-25.
- Garrido, O. H. 1982b. Nueva especie de *Anolis* (Lacertilia: Iguanidae) para Cuba. *Doñana Acta Vert.* 9: 31-37.
- Garrido, O. H. 1983. Nueva especie de *Anolis* de la Sierra del Turquino. *Carib. J. Sci.* 19(3-4): 71-76.
- Garrido, O. H. 1985. Nueva subespecie de *Anolis isolepis* (Lacertilia: Iguanidae) para Cuba. *Doñana Acta Vert.* 12(1): 41-49.
- Garrido, O. H. 1988. Nueva especie para la ciencia de *Anolis* (Lacertilia: Iguanidae) de Cuba perteneciente al complejo *argillaceus*. *Doñana Acta Vert.* 15(1): 45-57.
- Garrido, O. H. 1990. Nueva especie de *Anolis* de la sección beta (Lacertilia: Iguanidae) para Cuba. *Biología* 4: 157-162.
- Garrido, O. H., and S.B. Hedges. 1992. Three new grass anoles from Cuba (Squamata: Iguanidae). *Carib. J. Sci.* 28(1-2): 21-29.
- Garrido, O. H., and M. L. Jaume. 1984. Catálogo descriptivo de los Anfibios y Reptiles de Cuba. *Doñana Acta Vert.* 11(2): 5-128.
- Garrido, O. H., O. Pérez-Beato, and L. V. Moreno. 1991. Nueva especie de *Chamaeleolis* (Lacertilia: Iguanidae) para Cuba. *Carib. J. Sci.* 27(3-4): 162-168.
- Garrido, O. H., and A. Schwartz. 1972. The Cuban *Anolis spectrum* complex (Sauria, Iguanidae). *Proc. Biol. Soc. Washington* 85(45): 509-522.
- Gault, D. E., and C. P. Sonnet. 1982. Laboratory simulation of pelagic asteroidal impact: atmospheric injection, benthic topography, and the surface wave radiation field, pp. 69-92 in *Geological Implications of Impacts of Large Asteroids and Comets on the Earth*, edited by L. T. Silver, and P. H. Schultz. *Geol. Soc. Am. Spec. Pap.* 190. Boulder, CO.
- Gauthier, J. A., R. E. Estes, and K. de Queiroz. 1988. A phylogenetic analysis of Lepidosauromorpha, pp. 15-98 in *Phylogenetic Relationships of the Lizard Families: Essays Commemorating Charles L. Camp*, edited by R. Estes and G. Pregill. *Stanford Univ. Press*, Stanford, CA.

- Giannasi, N., R. S. Thorpe, and A. Malhorta. 1997. Introduction of *Anolis* species to the island of St. Lucia, West Indies: testing for hybrids using multivariate morphometrics. *J. Herpetol.* 31: 586-589.
- Gleason, H. A. 1922. On the relation between species and area. *Ecology* 3: 158-162.
- Goin, C. J. 1950. Color pattern inheritance in some frogs of the genus *Eleutherodactylus*. *Bull. Chicago Acad. Sci.* 9: 1-15.
- Goin, C. J. 1953. Rediscovery of the frog *Litoria luteola* Gosse in Jamaica. *Occ. Pap. Inst. Jamaica* 7: 1-4.
- Goin, C. J. 1954. Remarks on the evolution of color pattern in the *gossei* group of the frog genus *Eleutherodactylus*. *Ann. Carnegie Museum* 33: 185-195.
- Goin, C. J. 1958. Further studies on color pattern inheritance in the frog, *Eleutherodactylus nubicola*. *Yearbook Am. Philos. Soc.*: 248-250.
- Goin, C. J. 1959a. Notes on the maxillary dentition of some frogs of the genera *Eleutherodactylus* and *Leptodactylus*. *Herpetologica* 15: 134-136.
- Goin, C. J. 1959b. A synonym and a homonym in the frog genus *Hyla*. *Copeia* 4: 340-341.
- Goin, C. J. 1960. Pattern variation in the frog *Eleutherodactylus nubicola* Lynn. *Bull. Florida State Museum* 5: 243-258.
- Goin, C. J., and B. W. Cooper. 1950. Notes on a collection of amphibians from Jamaica. *Occ. Pap. Inst. Jamaica* 4: 1-9.
- Goldwasser, L., and J. Roughgarden. 1993. Construction and analysis of a large Caribbean food web. *Ecology* 74: 1216-1233.
- González-Bermúdez, F., and L. Rodríguez-Schettino. 1982. Datos etoecológicos sobre *Anolis vermiculatus* (Sauria: Iguanidae). *Poeyana* 245: 1-18.
- Good, D. A. 1988. Phylogenetic relationships among gerrhonotine lizards. An analysis of external morphology. *Univ. California Publ. Zool.* 121: 1-139.

- Gorman, G. C., and L. Atkins. 1968a. Natural hybridization between two sibling species of *Anolis* lizards: chromosome cytology. *Science* 159: 1358-1360.
- Gorman, G. C., and L. Atkins. 1968b. New karyotypic data for 16 species of *Anolis* (Sauria: Iguanidae) from Cuba, Jamaica, and the Cayman Islands. *Herpetologica* 24: 13-21.
- Gorman, G. C., and L. Atkins. 1969. The zoogeography of Lesser Antillean *Anolis* lizards – an analysis based upon chromosomes and lactic dehydrogenase. *Bull. Museum Comp. Zool.* 138: 53-80.
- Gorman, G. C., D. G. Buth, M. Soulé, and S. Y. Yang. 1980a. The relationships of the *Anolis cristatellus* species group: Electrophoretic analysis. *J. Herpetol.* 14: 269-278.
- Gorman, G. C., D. Buth, M. Soulé, and S. Y. Yang. 1983. The relationships of the Puerto Rican *Anolis*: Electrophoretic and karyotypic studies, pp. 626-642 in *Advances in Herpetology and Evolutionary Biology*, edited by A. G. J. Rhodin, and K. Miyata. *Museum Comp. Zool.*, Cambridge.
- Gorman, G. C., D. G. Buth, and J. S. Wyles. 1980b. *Anolis* lizards of the Eastern Caribbean: A case study of evolution. III. A cladistic analysis of albumin immunological data, and the definition of species groups. *Syst. Zool.* 29: 143-158.
- Gorman, G. C., and H. C. Dessauer. 1965. Hemoglobin and transferrin electrophoresis and relationships of island populations of *Anolis* lizards. *Science* 150: 1454-1455.
- Gorman, G. C., and H. C. Dessauer. 1966. The relationships of *Anolis* of the *roquet* species group (Sauria: Iguanidae) - I. Electrophoretic comparison of blood proteins. *Comp. Biochem. Physiol.* 19: 845-853.
- Gorman, G. C., and R. Harwood. 1977. Notes on population density, vagility, and activity patterns of the Puerto Rican grass lizard, *Anolis pulchellus* (Reptilia, Lacertii, Iguanidae). *J. Herpetol.* 11: 363-368.
- Gorman, G. C., and Y. J. Kim. 1976. *Anolis* lizards of the eastern Caribbean: a case study in evolution. II. Genetic relationships and genetic variation of the *bi-maculatus* group. *Syst. Zool.* 25: 62-77.

- Gosner, K. L. 1987. Observations on Lesser Antillean pit vipers. *J. Herpetol.* 21: 78-80.
- Gosse, P. H. 1847. *The Birds of Jamaica*. John Van Voorst, London.
- Gosse, P. H. 1848a. On the habits of *Ameiva dorsalis*. *Proc. Zool. Soc. London* 16: 24-27.
- Gosse, P. H. 1848b. On the habits of *Cyclura lophoma*, an iguaniform lizard. *Proc. Zool. Soc. London* 16: 99-104.
- Gosse, P. H. 1848c. On the habits of *Mabuya agilis*. *Proc. Zool. Soc. London* 16: 59-62.
- Gosse, P. H. 1850. Description of a new genus and six new species of Saurian reptiles. *Ann. Mag. Nat. Hist.* 6: 344-448.
- Gosse, P. H. 1851. *A Naturalist's Sojourn in Jamaica*. Longman, Brown, Green, & Longmans, London.
- Grant, C. 1931a. A new species of *Aristelliger* from Navassa. *J. Dept. Agric. Puerto Rico* 4: 399-400.
- Grant, C. 1931b. A new species and two new subspecies of the genus *Anolis*. *J. Dept. Agric. Puerto Rico* 15: 219-222.
- Grant, C. 1931c. Reestablishment of a scincid lost since 1837. *J. Dept. Agric. Porto Rico* 15: 217-218.
- Grant, C. 1932. The hemidactyls of the Porto Rico region. *J. Dept. Agric. Puerto Rico* 16: 51-57.
- Grant, C. 1938. The Jamaican *Dromicus funereus* re-established. *Copeia* 1938: 83-86.
- Grant, C. 1939a. Additional data on Jamaican snakes of the genus *Dromicus*. *Copeia* 1939: 105-106.
- Grant, C. 1939b. Two new sphaerodactyls from Jamaica. *Copeia* 1939: 7-13.
- Grant, C. 1940a. Notes on the reptiles and amphibians of Jamaica, with diagnoses of new species and subspecies. *Jamaica To-Day* 15: 151-157.

- Grant, C. 1940b. The herpetology of Jamaica II. The reptiles. Bull. Inst. Jamaica Sci. Ser. 1: 61-148.
- Grant, C. 1940c. The herpetology of the Cayman Islands. Bull. Inst. Jamaica Sci. Ser. 2: iv + 65.
- Grant, C. 1944a. New sphaerodactyls from Cuba and the Isle of Pines. Herpetologica 2(6):118-125.
- Grant, C. 1944b. Scale structure in Jamaican lizards of the genus *Celestus*. Copeia 1944: 109-111.
- Grant, C. 1946. Note from Major Chapman Grant. Nat. Hist. Notes Nat. Hist. Soc. Jamaica 3: 78.
- Grant, C. 1951. The specific characters of the Celesti, with description of a new species of *Celestus* (Sauria, Anguidae). Copeia 1951: 67-69.
- Grant, C. 1958. A new *Gymnophthalmus* (Reptilia, Teiidae) from Barbados. Herpetologica 14: 227-228.
- Grant, C. 1959. Herpetology of Barbados, B.W.I. Herpetologica 15: 97-101.
- Grant, C., and C. R. D. Sola. 1934. Antillean tortoises and terrapins: distribution, status, and habits of *Testudo* and *Pseudemys*. Copeia 1934: 73-79.
- Gravenhorst, J. L. C. 1829. Reptilia Musei Zoologici Vratislaviensis Recensita et Descriptiva. Fasciculus Primus continens Chelonids et Batrachia. Sumptibus Leopoldi Vossi, Lipsiae.
- Gravenhorst, J. L. C. 1837/1838. Beiträge zur genaueren Kenntniss einiger Eidechsen-gattungen. Nova Acta Acad. Caes. Leop.-Carol. 18: 712-784.
- Gray, J. E. 1827. A description of a new genus and some new species of saurian reptiles; with a revision of the species of chameleons. Philos. Mag. 2: 207-214.
- Gray, J. E. 1830. Specilegia Zoologica, or Original Figures and Short Systematic Descriptions of New and Unfigured Animals. Part II. Trustees of the British Museum, London.

- Gray, J. E. 1831. A synopsis species of the class Reptilia, pp. 1-110 in *The Class Reptilia, Arranged by Baron C. Cuvier, with Specific Descriptions. The Animal Kingdom Arranged in Conformity with its Organization by the Baron Cuvier, with Additional Descriptions of all the Species Hitherto Named, and of Many Others.* Edited by E. Griffiths and E. Pidgeon. Vol. 9., Whitaker, London.
- Gray, J. E. 1838a. Catalog of the slender-tongued Saurians, with descriptions of many new genera and species. *Annot. Nat. Hist.* 1: 274-283.
- Gray, J. E. 1838b. Catalogue of the slender-tongued Saurians, with descriptions of many new genera and species. *Annu. Mag. Nat. Hist.* 2: 287-293.
- Gray, J. E. 1840. Catalogue of the species of reptiles collected in Cuba by W. S. MacLeay, Esp. – with some notes on their habits extracted from his MS. *Annu. Mag. Nat. Hist.* 5: 108-115.
- Gray, J. E. 1842. Synopsis of the species of prehensile-tailed snakes, or family Boidae. *Zool. Misc.* 41-46.
- Gray, J. E. 1845. Catalogue of the Specimens of Lizards in the Collection of the British Museum. Newman, London.
- Gray, J. E. 1852. Description of *Sauresia*, a new genus of Scincidae from St. Domingo. *Annu. Mag. Nat. Hist.* 10: 281-282.
- Graybeal, A., and D. C. Cannatella. 1995. A new taxon of Bufonidae from Peru, with descriptions of two new species and a review of the phylogenetic status of supraspecific Bufonid taxa. *Herpetologica* 51: 105-131
- Greene, H. W. 1988. Species richness in tropical predators, pp. 259-280 in *Tropical Rainforests: Diversity and Conservation*, edited by C. F. Almeda, and M. Pringle. California Acad. Sci. and Pacific Div. American Assoc. Adv. Sci., San Francisco.
- Greene, H. W. 1994. Systematics and natural history, foundations for understanding and conserving biodiversity. *Am. Zool.* 34: 48-56.
- Greer, A. E. 1965. A new subspecies of *Clelia clelia* (Serpentes: Colubridae) from the island of Grenada. *Breviora* 223: 1-6.
- Greer, A. E. 1967. Notes on the mode of reproduction in anguid lizards. *Herpetologica* 23:94-99.

- Greer, A. E. 1970. Evolutionary and systematic significance of crocodylian nesting habits. *Nature* 227: 523-524.
- Griffiths, I. 1959. The phylogeny of *Sminthillus limbatus* and the status of the Brachycephalidae (Amphibia: Salientia). *Proc. Zool. Soc., London* 132: 457-487.
- Groombridge, B. 1987. The distribution and status of world crocodylians, pp. 9-21 in *Wildlife Management: Crocodiles and Alligators*, edited by G. Webb, C. Manolis and P. Whitehead. Surrey Beatty & Sons, Chipping Norton, Australia.
- Gundlach, J. 1867. Revista y catalogo de los reptiles cubanos. *Rep. Fisico-Natural Isla Cuba* 2: 102-119.
- Gundlach, J. 1880. Contribución a la Erpetología Cubana. G. Montiel, La Habana.
- Günther, A. C. L. G. 1858. Catalogue of Colubrine Snakes in the Collection of the British Museum. Trustees of the British Museum, London.
- Günther, A. C. L. G. 1858 (1859). Catalogue of the Batrachia Salientia in the Collection of the British Museum. Trustees of the British Museum, London.
- Günther, A. C. L. G. 1861. On a new species of the family Boidae. *Proc. Zool. Soc. London* 1861: 142.
- Günther, A. C. L. G. 1862. On new species of snakes in the collection of the British Museum. *Annu. Mag. Nat. Hist.* 9: 124-132.
- Günther, A. C. L. G. 1865. Fourth account of new species of snakes in the collection of the British Museum. *Annu. Mag. Nat. Hist.* 15: 89-98.
- Günther, A. 1888. Notes on reptiles and frogs from Dominica, West Indies. *Annu. Mag. Nat. Hist.* 2: 362-366.
- Günther, A. E. 1980. *The Founders of Science at the British Museum, 1753-1900*. Halesworth, Suffolk, UK.
- Guyer, C. 1988a. Food supplementation in a tropical mainland anole, *Norops humilis*: demographic effects. *Ecology* 69: 350-361.

- Guyer, C. 1988b. Food supplementation in a tropical mainland anole, *Norops humilis*: effects on individuals. *Ecology* 69: 362-369.
- Guyer, C. 1990. The herpetofauna of La Selva, Costa Rica, pp. 371-385 in *Four Neotropical Rainforests*, edited by A. H. Gentry. Yale Univ. Press, New Haven, CT.
- Guyer, C., and B. I. Crother. 1996. Additional comments on the origin of the West Indian herpetofauna. *Herpetologica* 52: 620-622.
- Guyer, C., and J. M. Savage. 1986. Cladistic relationships among anoles (Sauria: Iguanidae). *Syst. Zool.* 35: 509-531.
- Guyer, C., and J. M. Savage. 1992. Anole systematics revisited. *Syst. Biol.* 41: 89-120.
- Haefner, J. W. 1988. Assembly rules for Greater Antillean *Anolis* lizards. *Oecologia* 74: 551-565.
- Hager, J., and T. A. Zanoni. 1993. La vegetación natural de la República Dominicana: una nueva clasificación. *Moscoso* 7: 39-81.
- Hallowell, E. 1857. Notes on the reptiles in the collection of the Academy of Natural Sciences of Philad'a. *Proc. Acad. Nat. Sci. Philadelphia* 8: 221-238.
- Hardy, J.D. 1967. Geographic variation in the West Indian lizard, *Anolis angusticeps*, with the description of a new form, *Anolis angusticeps paternus*, subsp. nov., from the Isle of Pines, Cuba (Reptilia: Iguanidae). *Carib. J. Sci.* 6(1-2): 23-31.
- Harlan, R. 1824/1825. Description of two species of Linnæan *Lacerta*, not before described, and construction of the new genus *Cyclura*. *J. Acad. Nat. Sci. Philadelphia* 4: 242-251.
- Harlan, R. 1826a. Descriptions of several new species of batrachian reptiles, with observations on the larvae of frogs. *Am. J. Sci. Arts* 10: 53-65.
- Harlan, R. 1826b. A new species of *Hyla*. *Am. J. Sci. Arts* 10: 64-65.
- Harris, D. M., and A. G. Kluge. 1984. The *Sphaerodactylus* (Sauria: Gekkonidae) of Middle America. *Occas. Pap. Museum Zool. Univ. Michigan* 706: 1-59.

- Harvey, P. H., and M. D. Pagel. 1991. *The Comparative Method in Evolutionary Biology*. Oxford Univ. Press, Oxford, UK.
- Hass, C. A. 1991. Evolution and biogeography of West Indian *Sphaerodactylus* (Sauria: Gekkonidae): a molecular approach. *J. Zool. (London)* 225: 525-561.
- Hass, C. A. 1996. Relationships among West Indian geckos of the genus *Sphaerodactylus*: a preliminary analysis of mitochondrial 16S ribosomal RNA sequences, pp. 175-194 in *Contributions to West Indian Herpetology: A Tribute to Albert Schwartz*, edited by R. Powell, and R. W. Henderson. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.
- Hass, C. A., and S. B. Hedges. 1991. Albumin evolution in West Indian frogs of the genus *Eleutherodactylus* (Leptodactylidae): Caribbean biogeography and a calibration of the albumin immunological clock. *J. Zool. London* 225: 413-426.
- Hass, C. A., and S. B. Hedges. 1992. Karyotype of the Cuban lizard *Cricosaura typica* and its implications for Xantusiid phylogeny. *Copeia* 1992: 563-565.
- Hass, C. A., S. B. Hedges, and L. R. Maxson. 1992. The relationships of West Indian anguid lizards. 1992 ASIH/HL Joint Meetg. Progr.: 115. (abstract).
- Hass, C. A., S. B. Hedges, and L. Maxson. 1993. Molecular insights into the relationships and biogeography of West Indian anoline lizards. *Biochem. Syst. Ecol.* 21: 97-114.
- Heatwole, H., R. Levins, and M. D. Byer. 1981. Biogeography of the Puerto Rican Bank. *Atoll. Res. Bull.* 251: 1-55.
- Heatwole, H., and F. MacKenzie. 1966. Herpetogeography of Puerto Rico. IV. Paleogeography, faunal similarity and endemism. *Evolution* 21: 429-438.
- Heatwole, H., and O. J. Sexton. 1966. Herpetofaunal comparisons between two climatic zones in Panama. *Am. Midl. Natur.* 75: 45-60.
- Heatwole, H., and F. Torres. 1967. Distribution and geographic variation of the ameivas of Puerto Rico and the Virgin Islands. *Studies Fauna Curaçao Carib. Isl.* 24: 64-111.

- Hecht, M. K. 1951. Fossil lizards of the West Indian genus *Aristelliger* (Gekkonidae). *Am. Museum Novit.* 1538: 1-33.
- Hecht, M. K. 1952. Natural selection in the lizard genus *Aristelliger*. *Evolution* 6: 112-124.
- Hedges, S. B. 1982. Caribbean biogeography: implications from recent plate tectonic studies. *Syst. Zool.* 31: 518-522.
- Hedges, S. B. 1987. Vocalization and habitat preference of the Jamaican tree-frog, *Hyla marianae* (Anura: Hylidae). *Carib. J. Sci.* 23: 380-384.
- Hedges, S. B. 1988. A new diminutive frog from Hispaniola (Leptodactylidae: *Eleutherodactylus*). *Copeia* 1988: 636-641.
- Hedges, S. B. 1989a. Evolution and biogeography of West Indian frogs of the genus *Eleutherodactylus*: slow-evolving loci and the major groups, pp. 305-370 in *Biogeography of the West Indies: Past, Present, and Future*, edited by C. A. Woods. Sandhill Crane Press, Gainesville, FL.
- Hedges, S. B. 1989b. An island radiation: allozyme evolution in Jamaican frogs of the genus *Eleutherodactylus* (Anura, Leptodactylidae). *Carib. J. Sci.* 25: 123-147.
- Hedges, S. B. 1989c. Geographic protein variation in the Jamaican blind snake, *Typhlops jamaicensis* (Serpentes: Typhlopidae). *Carib. J. Sci.* 25: 93-96.
- Hedges, S. B. 1991. Electrophoretic and morphologic variation in *Eleutherodactylus glaphycompus* (Anura: Leptodactylidae) of Hispaniola. *J. Herpetol.* 25: 10-17.
- Hedges, S. B. 1993. Global amphibian declines: a perspective from the Caribbean. *Biodiver. Conserv.* 2: 290-303.
- Hedges, S. B. 1996a. Historical biogeography of West Indian vertebrates. *Annu. Rev. Ecol. Syst.* 27: 163-196.
- Hedges, S. B. 1996b. The origin of West Indian amphibians and reptiles, pp. 95-128 in *Contributions to West Indian Herpetology: A Tribute to Albert Schwartz*, edited by R. Powell, and R. W. Henderson. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.

- Hedges, S. B. 1996c. Vicariance and dispersal in Caribbean biogeography. *Herpetologica* 52: 466-473.
- Hedges, S. B. 1996d. The Hispaniolan frog *Eleutherodactylus neodreptus* Schwartz (Anura: Leptodactylidae) is a synonym of *E. audanti* Cochran. *Carib. J. Sci.* 32: 248.
- Hedges, S. B. 1996e. More on West Indian zoogeography. *Herpetologica* 52: 622-624.
- Hedges, S. B. 1999. Distribution patterns of amphibians in the West Indies, pp. 000-000 in *Regional Patterns of Amphibian Distribution: A Global Perspective*, edited by W. E. Duellman. Johns Hopkins Univ. Press, Baltimore, Maryland.
- Hedges, S. B. and R. L. Bezy. 1993. Phylogeny of xantusiid lizards: concern for data and analysis. *Mol. Phylogen. Evol.* 2:76-87.
- Hedges, S. B., and R. L. Bezy. 1994. Xantusiid lizards and phylogenetic inference. *Mol. Phylogen. Evol.* 3: 275-278.
- Hedges, S. B., R. L. Bezy, and L. R. Maxson. 1991. Phylogenetic relationships and biogeography of xantusiid lizards, inferred from mitochondrial DNA sequences. *Mol. Biol. Evol.* 8: 767-780.
- Hedges, S. B., and K. L. Burnell. 1990. The Jamaican radiation of *Anolis* (Sauria: Iguanidae): an analysis of relationships and biogeography using sequential electrophoresis. *Carib. J. Sci.* 26: 31-44.
- Hedges, S. B., A. R. Estrada, and R. Thomas. 1992a. Three new species of *Eleutherodactylus* from eastern Cuba, with notes on vocalizations of other species (Anura:Leptodactylidae). *Herpetol. Monograph*, 6:68-83.
- Hedges, S. B. and O. H. Garrido. 1992a. A new species of *Tropidophis* from Cuba (Serpentes: Tropidophidae). *Copeia* 1992(3): 820-825.
- Hedges, S. B., and O. H. Garrido. 1992b. Cuban snakes of the genus *Arrhyton*: two new species and a reconsideration of *A. redimitum* Cope. *Herpetologica* 48: 168-177.
- Hedges, S. B., and O. H. Garrido. 1993. A new species of gecko (*Sphaerodactylus*) from Central Cuba. *J. Herpetol.* 27(3): 300-306.

- Hedges, S. B., C. A. Hass, and T. K. Mauge. 1989. Physiological color change in snakes. *J. Herpetol.* 23: 450-455.
- Hedges, S. B., L. González, and A. R. Estrada. 1995. Rediscovery of the Cuban frog *Eleutherodactylus cubanus* and *E. turquinensis* (Anura: Leptodactylidae). *Carib. J. Sci.* 31(3-4): 327-332.
- Hedges, S. B., C. A. Hass, and L. R. Maxson. 1992b. Caribbean biogeography: molecular evidence for dispersal in West Indian terrestrial vertebrates. *Proc. Natl. Acad. Sci. (USA)* 89: 1909-1913.
- Hedges, S. B., C. A. Hass, and L. R. Maxson. 1994. Reply: towards a biogeography of the Caribbean. *Cladistics* 10: 43-55.
- Hedges, S. B., and R. Thomas. 1987. A new burrowing frog from Hispaniola with comments on the *inoptatus* group of the genus *Eleutherodactylus* (Anura: Leptodactylidae). *Herpetologica* 43: 269-279.
- Hedges, S. B., and R. Thomas. 1989a. Supplement to West Indian amphibians and reptiles: a check-list. *Milwaukee Publ. Museum Contr. Biol. Geol.* 77: 1-11.
- Hedges, S. B., and R. Thomas. 1989b. A new species of *Anolis* (Sauria: Iguanidae) from the Sierra de Neiba, Hispaniola. *Herpetologica* 45: 330-336.
- Hedges, S. B., and R. Thomas. 1991a. The importance of systematic research in the conservation of amphibian and reptile populations, pp. 56-61 in *Status y Distribución de los Reptiles y Anfibios de la Región de Puerto Rico*, edited by J. A. Moreno. Dept. Recur. Nat. Puerto Rico.
- Hedges, S. B., and R. Thomas. 1991b. Cryptic species of snakes (Typhlopidae: *Typhlops*) from the Puerto Rico Bank detected by protein electrophoresis. *Herpetologica* 47: 448-459.
- Hedges, S. B., and R. Thomas. 1992a. A new marsh-dwelling species of *Eleutherodactylus* from Haiti (Anura: Leptodactylidae). *J. Herpetol.* 26: 191-195.
- Hedges, S. B., and R. Thomas. 1992b. Two new species of *Eleutherodactylus* from remnant cloud forest in Haiti (Anura, Leptodactylidae). *Herpetologica* 48: 351-358.

- Hedges, S. B., and R. Thomas. 1998. A new anguid lizard from Cuba. *Copeia* 1998: 97-103.
- Hedges, S. B., R. Thomas, and R. Franz. 1987. A new species of *Eleutherodactylus* (Anura, Leptodactylidae) from the Massif de la Hotte, Haiti. *Copeia* 1987: 943-949.
- Heise, P. J., L. R. Maxson, H. G. Dowling, and S. B. Hedges. 1995. Higher-level snake phylogeny inferred from mitochondrial DNA sequences of 12S rRNA and 16S rRNA genes. *Mol. Biol. Evol.* 12(2): 259-265.
- Henderson, R. W. 1974. Aspects of the ecology of the neotropical vine snake, *Oxybelis aeneus* (Wagler). *Herpetologica* 30: 19-24.
- Henderson, R. W. 1984. The diet of the Hispaniolan snake *Hypsirhynchus ferox* (Colubridae). *Amphib.-Rept.* 5: 367-371.
- Henderson, R. W. 1988a. *Diploglossus anelpistus*. *Cat. Am. Amphib. Rept.* 424: 1.
- Henderson, R. W. 1988b. *Diploglossus carraui*. *Cat. Am. Amphib. Rept.* 425: 1.
- Henderson, R. W. 1990. A new subspecies of *Alsophis antiquae* (Parker) from Great Bird Island (Antigua), Lesser Antilles. *Carib. J. Sci.* 25: 119-122.
- Henderson, R. W. 1992. Consequences of predator introductions and habitat destruction on amphibians and reptiles in the post-Columbus West Indies. *Carib. J. Sci.* 28: 1-10.
- Henderson, R. W. 1993a. Foraging and diet in West Indian *Corallus enydris* (Serpentes: Boidae). *J. Herpetol.* 27: 24-28.
- Henderson, R. W. 1993b. On the diets of some arboreal boids. *Herpetol. Nat. Hist.* 1: 91-96.
- Henderson, R. W., M. H. Binder, and R. A. Sajdak. 1982. Ecological relationships of the tree snakes *Uromacer catesbyi* and *U. oxyrhynchus* (Colubridae) on Isla Saona, República Dominicana. *Amphib.-Rept.* 2: 153-163.
- Henderson, R. W., and R. W. Bourgeois. 1993. Notes on the diets of West Indian *Liophis* (Serpentes: Colubridae). *Carib. J. Sci.* 29: 253-254.

- Henderson, R. W., and B. I. Crother. 1989. Biogeographic patterns of predation in West Indian colubrid snakes, pp. 479-517 in *Biogeography of the West Indies: Past, Present, and Future*, edited by C. A. Woods. Sandhill Crane Press, Inc., Gainesville.
- Henderson, R. W., B. I. Crother, T. A. Noeske-Hallin, A. Schwartz, and C. R. Dethloff. 1987a. The diet of the Hispaniolan snake *Antillophis parvifrons* (Colubridae). *J. Herpetol.* 21: 330-334.
- Henderson, R. W., A. Delatte, and T. J. McCarthy. 1993. *Gekko gecko* (Sauria: Gekkonidae) established on Martinique, French West Indies. *Carib. J. Sci.* 29: 128-129.
- Henderson, R. W., and G. T. Haas. 1993. Status of the West Indian snake *Chironius vincenti*. *Oryx* 27: 181-184.
- Henderson, R. W., and S. B. Hedges. 1995. Origin of West Indian populations of the geographically widespread boa *Corallus enydris* inferred from mitochondrial DNA sequences. *Mol. Phylo. Evol.* 4: 88-92.
- Henderson, R. W., and K. F. Henderson. 1995. Altitudinal variation in body temperature in foraging tree boas (*Corallus enydris*) on Grenada. *Carib. J. Sci.* 31: 73-76.
- Henderson, R. W., T. A. Noeske-Hallin, B. I. Crother, and A. Schwartz. 1988a. The diets of Hispaniolan colubrid snakes II. Prey species, prey size, and phylogeny. *Herpetologica* 44: 55-70.
- Henderson, R. W., T. A. Noeske-Hallin, J. A. Ottenwalder, and A. Schwartz. 1987b. On the diet of the boa *Epicrates striatus* on Hispaniola, with notes on *E. fordi* and *E. gracilis*. *Amphib.-Rept.* 8: 251-258.
- Henderson, R. W., R. Powell, J. C. Daltry, and M. L. Day. 1996. *Alsophis antiguae*. *Cat. Amer. Amphib. Rept.* (632):1-3.
- Henderson, R. W., and R. A. Sajdak. 1986. West Indian racers: a disappearing act or a second chance? *Lore, Milwaukee Publ. Museum* 36: 13-18.
- Henderson, R. W., and R. A. Sajdak. 1996. Diets of West Indian racers (Colubridae: *Alsophis*): Composition and biogeographic implications, pp. 327-338 in *Contributions to West Indian Herpetology: A Tribute to Albert Schwartz*, edited

by R. Powell, and R. W. Henderson. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.

Henderson, R. W., R. A. Sajdak, and R. M. Henderson. 1988b. The rediscovery of the West Indian colubrid snake *Chironius vincenti*. Amphib.-Rept. 9: 415-416.

Henderson, R. W., and A. Schwartz. 1984a. A guide to the identification of the amphibians and reptiles of Hispaniola. Milwaukee Public Museum Spec. Publ. Biol. Geol. 4: 1-70.

Henderson, R. W., and A. Schwartz. 1984b. *Uromacer frenatus*. Cat. Am. Amphib. Rept. 357.1-357.2.

Henderson, R. W., and A. Schwartz. 1986. The diet of the Hispaniolan colubrid snake, *Darlingtonia haetiana*. Copeia 1986: 529-531.

Henderson, R. W., A. Schwartz, and S. J. Incháustegui. 1984. Guía para la identificación de los anfibios y reptiles de la Hispaniola. Museum Nac. Hist. Nat. Ser. Monogr. 1: 1-128.

Henderson, R. W., A. Schwartz, and T. A. Noeske-Hallin. 1987c. Food habits of three colubrid tree snakes (genus *Uromacer*) on Hispaniola. Herpetologica 43: 235-242.

Henderson, R. W., and R. A. Winstel. 1992. Activity patterns, temperature relationships, and habitat utilization in *Corallus enydris* (Serpentes: Boidae) on Grenada. Carib. J. Sci. 28: 229-232.

Henderson, R. W., and R. A. Winstel. 1995. Aspects of habitat selection by an arboreal boa (*Corallus enydris*) in an area of mixed agriculture on Grenada. J. Herpetol. 29: 272-275.

Henderson, R. W., R. A. Winstel, and J. Friesch. 1996. *Corallus hortulanus* (Serpentes: Boidae) in the post-Columbian West Indies: New habitats, new prey species, and new predators, pp. 417-423 in Contributions to West Indian Herpetology: A Tribute to Albert Schwartz, edited by R. Powell, and R. W. Henderson. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.

Hennig, W. 1966. Phylogenetic Systematics. Univ. of Illinois Press, Urbana, IL.

- Hertz, P. E. 1976. *Anolis alumina*, new species of grass anole from the Barahona Peninsula of Hispaniola. *Breviora* 437: 1-19.
- Hertz, P. E. 1981. Adaptation to altitude in two West Indian anoles (Reptilia: Iguanidae): field thermal biology and physiological ecology. *J. Zool. London* 195: 25-37.
- Hertz, P. E. 1983. Eurythermy and niche breadth in West Indian *Anolis* lizards: a reappraisal, pp. 472-483 in *Advances in Herpetology and Evolutionary Biology: Essays in Honor of Ernest E. Williams*, edited by A. G. J. Rhodin, and K. I. Miyata. Museum Comp. Zool., Cambridge, MA.
- Hertz, P. E., R. B. Huey, and R. D. Stevenson. 1993. Evaluating temperature regulation by field-active ectotherms: the fallacy of the inappropriate question. *Am. Nat.* 142: 796-818.
- Heyer, W. R. 1969. The adaptive ecology of the species groups of the genus *Leptodactylus* (Amphibia, Leptodactylidae). *Evolution* 23: 421-428.
- Heyer, W. R. 1970. Studies on frogs of the genus *Leptodactylus* (Amphibia: Leptodactylidae). VI. Biosystematics of the *melanotus* group. *Contrib. Sci. Nat. Hist. Museum Los Angeles County*. 191: 1-48.
- Heyer, W. R. 1974. Systematics of the *marmoratus* group of the frog genus *Leptodactylus* (Amphibia, Leptodactylidae). *Los Angeles County Museum Nat. Hist. Contrib. Sci.* 253: 1-50.
- Heyer, W. R. 1978. Systematics of the *fuscus* group of the frog genus *Leptodactylus* (Amphibia, Leptodactylidae). *Smithsonian Contrib. Zool.* 301: 1-43.
- Heyer, W. R. 1979. Systematics of the *pentadactylus* species group of the frog genus *Leptodactylus* (Amphibia: Leptodactylidae). *Smithsonian Contrib. Zool.* 301: 1-43.
- Heyer, W. R. 1994. Variation within the *Leptodactylus podicipinus-wagneri* complex of frogs (Amphibia: Leptodactylidae). *Smithsonian Contrib. Zool.* 546: 1-124.
- Heyer, W. R., M. A. Donnelly, R. W. McDiarmid, L.-A. C. Hayek, and M. S. Foster. 1994. *Measuring and Monitoring Biological Diversity: Standard Methods for Amphibians*, Smithsonian Institution Press, Washington, DC.

- Hicks, R. 1973. New studies on a montane lizard of Jamaica, *Anolis reconditus*. *Breviora* 404: 1-23.
- Hicks, R. A., and R. L. Trivers. 1983. The social behavior of *Anolis valencienni*, pp. 570-595 in *Advances in Herpetology and Evolutionary Biology: Essays in Honor of Ernest E. Williams*, edited by A. G. J. Rhodin, and K. I. Miyata. *Museum Comp. Zool. Harvard Univ.*, Cambridge, MA.
- Hildebrand, A. R., and W. V. Boynton. 1990. Proximal Cretaceous-Tertiary boundary impact deposits in the Caribbean. *Science* 248: 843-847.
- Hillis, D. M., B. K. Mable, and C. Moritz. 1996. Applications of molecular systematics: the state of the field and a look to the future, pp. 515-543 in *Molecular Systematics*, edited by D. M. Hillis, C. Moritz, and B. K. Mable. *Sinauer Associates*, Sunderland, MA.
- Hirth, H. F. 1963. The ecology of two lizards on a tropical beach. *Ecol. Monogr.* 33: 83-112.
- Hoge, A. R. 1952. Notas erpetológicas. Revalidación de *Bothrops lanceolata* (Lace). *Mem. Inst. Butantan* 24: 231-236.
- Holdridge, L. R. 1967. *Life zone ecology*. Tropical Sci. Center, San Jose, Costa Rica.
- Hollingsworth, B. D. 1998. The systematics of chuckwallas (*Sauraomalus*) with a phylogenetic analysis of other iguanid lizards. *Herp. Monog.* 12: 38-191.
- Hoogmoed, M. S. 1973. Notes on the herpetofauna of Surinam IV. *Junk*, The Hague.
- Hoppe, J. 1989. *Los Parques Nacionales de la República Dominicana*. Ed. Corripio, Santo Domingo.
- Horn, H. S. 1969. Polymorphism and evolution of the Hispaniolan snake genus *Uromacer* (Colubridae). *Breviora* 324: 1-23.
- Howard, A. K., J. D. Forester, J. M. Ruder, R. Powell, and J. J. S. Parmerlee. MS. The diets of two sympatric frogs: *Eleutherodactylus abbotti* and *E. armstrongi* (Leptodactylidae) from the Sierra de Baoruco, Hispaniola. Unpublished manuscript.

- Huebeck, C., and P. Mann. 1991. Structural geology and Cenozoic tectonic history of the southeastern termination of the Cordillera Central, Dominican Republic. *Geol. Soc. Am. Spec. Pap.* 262: 315-336.
- Huey, R. B., and T. P. Webster. 1975. Thermal biology of a solitary lizard: *Anolis marmoratus* of Guadeloupe, Lesser Antilles. *Ecology* 56: 445-452.
- Huey, R. B., and T. P. Webster. 1976. Thermal biology of *Anolis* lizards in a complex fauna: the *crisatellus* group on Puerto Rico. *Ecology* 57: 985-994.
- Hull, D. L. 1988. *Science as a Process*. Univ. of Chicago Press, Chicago, IL.
- Humbolt, A. 1856. *The island of Cuba*. Derby & Jackson, New York.
- Inchaústegui, S. J., A. Schwartz, and R. W. Henderson. 1985. Hispaniolan giant *Diploglossus* (Sauria: Anguidae): description of a new species and notes on the ecology of *D. warreni*. *Amphib.-Rept.* 6: 195-201.
- Institute, W. R. 1994. *World Resources 1994-95*. Oxford Univ. Press, New York.
- Isacks, B., J. Oliver, and L. R. Sykes. 1968. Seismology and the new global tectonics. *J. Geophys. Res.* 73: 5855-5899.
- Iverson, J. B. 1978. The impact of feral cats and dogs on populations of the West Indian rock iguana, *Cyclura carinata*. *Biol. Conserv.* 14: 63-73.
- Iverson, J. B. 1979. Behavior and ecology of the rock iguana *Cyclura carinata*. *Bull. Florida State Museum, Biol. Sci.* 24: 175-358.
- Jan, G., and F. Sordelli. 1867. *Iconographie Générale des Ophidiens*. Livr. 25. Published by authors, Milan.
- Jeffrey-Smith, M. 1946. The whistling frog increases. *Nat. Hist. Notes Nat. Hist. Soc. Jamaica* 3: 26.
- Jenner, J. V. 1981. A zoogeographic study and the taxonomy of the xenodontine colubrid snakes. Ph.D. Dissertation. New York Univ.
- Jenner, J. V., and H. G. Dowling. 1985. Taxonomy of American xenodontine snakes: the tribe Pseudoboini. *Herpetologica* 41: 161-172.

- Jenssen, T. A. 1970. The ethoecology of *Anolis nebulosus* (Sauria, Iguanidae). *J. Herpetol.* 4: 1-38.
- Jenssen, T. A. 1973. Shift in structural habitat of *Anolis opalinus* due to congeneric competition. *Ecology* 54: 863-869.
- Jenssen, T. A. 1977. Morphological, behavioral and electrophoretic evidence of hybridization between the lizards, *Anolis grahami* and *Anolis lineatopus neckeri*, on Jamaica. *Copeia* 1977: 270-276.
- Jenssen, T. A. 1979. Display modifiers of *Anolis opalinus* (Lacertilia: Iguanidae). *Herpetologica* 35: 21-30.
- Jenssen, T. A., and R. M. Andrews. 1984. Seasonal growth rates in the Jamaican lizard, *Anolis opalinus*. *J. Herpetol.* 18: 338-341.
- Jenssen, T. A., and S. C. Nunez. 1994. Male and female reproductive cycles of the Jamaican lizard, *Anolis opalinus*. *Copeia* 1994: 767-780.
- Jenssen, T. A., D. L. Marcellini, K. A. Buhlmann, and P. H. Goforth. 1989. Differential infanticide in adult curly-tailed lizards, *Leiocephalus schreibersi*. *Anim. Behav.* 38: 1054-1061.
- Joger, U. 1984. Taxonomische Revision der Gattung *Tarentola* (Reptilia: Gekkonidae). *Bonn Zool. Beitr.* 35: 129-174.
- Joglar, R. L. 1983. Estudio fenético del género *Eleutherodactylus* en Puerto Rico. *Carib. J. Sci.* 19: 33-40.
- Joglar, R. L. 1989. Phylogenetic relationships of the West Indian frogs of the genus *Eleutherodactylus*: a morphological analysis, pp. 371-404 in *Biogeography of the West Indies: Past, Present, and Future*, edited by C. A. Woods. Sandhill Crane Press, Gainesville, FL.
- Joglar, R. L., and P. A. Burrowes. 1996. Declining amphibian populations in Puerto Rico, pp. 371-380 in *Contributions to West Indian Herpetology: A Tribute to Albert Schwartz*, edited by R. Powell, and R. W. Henderson. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.

- Jones, K. L. 1982. Prey patterns and trophic niche overlap in four species of Caribbean frogs, pp. 49-55 in *Herpetological Communities*, edited by J. N. Jr. Scott. Research Report 13, United States Fish and Wildlife Service.
- Kaiser, H. 1992. The trade-mediated introduction of *Eleutherodactylus martinicensis* (Anura: Leptodactylidae) on St. Barthélemy, French Antilles, and its implications for Lesser Antillean Biogeography. *J. Herpetol.* 26: 264-273.
- Kaiser, H. 1996. Systematics and biogeography of Eastern Caribbean *Eleutherodactylus* (Anura: Leptodactylidae): consensus from a multidisciplinary approach, pp. 129-140 in *Contributions to West Indian Herpetology: A Tribute to Albert Schwartz*, edited by R. Powell, and R. W. Henderson. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.
- Kaiser, H. 1997. Origins and introductions of the Caribbean frog, *Eleutherodactylus johnstonei* (Leptodactylidae): management and conservation concerns. *Biodivers. Conserv.* in press.
- Kaiser, H., and R. Altig. 1994. The atypical tadpole of the dendrobatid frog, *Colostethus chalcopis*, from Martinique, French Antilles. *J. Herpetol.* 28: 374-378.
- Kaiser, H., L. A. Coloma, and H. M. Gray. 1994b. A new species of *Colostethus* (Anura: Dendrobatidae) from Martinique, French Antilles. *Herpetologica* 50: 23-32.
- Kaiser, H., D. M. Green, and M. Schmid. 1994a. Systematics and biogeography of Eastern Caribbean *Eleutherodactylus* (Anura: Leptodactylidae) with the description of a new species from Dominica. *Can. J. Zool.* 72: 2217-2237.
- Kaiser, H., and J. J. D. Hardy. 1994. *Eleutherodactylus martinicensis*. *Cat. Am. Amphib. Rept.* 582: 1-4.
- Kaiser, H., J. J. D. Hardy, and D. M. Green. 1994c. Taxonomic status of Caribbean and South American frogs currently ascribed to *Eleutherodactylus urichi* (Anura: Leptodactylidae). *Copeia* 1994: 780-796.
- Kaiser, H., and R. W. Henderson. 1994. The conservation status of Lesser Antillean frogs. *Herpetol. Nat. Hist.* 2: 41-56.
- Kaiser, H., and H. Kaiser. 1994. *Eleutherodactylus johnstonei*. *Cat. Am. Amphib. Rept.* 581: 1-5.

- Kaiser, H., T. F. Sharbel, and D. M. Green. 1994d. Systematics and biogeography of eastern Caribbean *Eleutherodactylus* (Anura: Leptodactylidae): evidence from allozymes. *Amphib.-Rept.* 15: 375-394.
- Keegan, W. F., and J. M. Diamond. 1987. Colonization by humans: a biogeographical perspective. *Adv. Archeol. Method Theory.* 10: 49-92.
- Keller, G., J. G. Lopez-Oliva, W. Stinnesbeck, and T. Adatte. 1997. Age, stratigraphy, and deposition of near K/T siliciclastic deposits in Mexico: Relation to bolide impact? *GSA Bull.* 109: 410-428.
- Kerr, R. A. 1998. Research News: Sea-floor dust shows drought felled Akkadian Empire. *Science* 279: 325-326.
- Kerster, H. W., and H. M. Smith. 1955. The identity of the Puerto Rican species of *Phyllodactylus* (Reptilia: Squamata). *Herpetologica* 11: 229-232.
- Khudoley, K. M., and A. A. Myerhoff. 1971. Paleogeography and geological history of the Greater Antilles. *Geol. Soc. of Am. Memoirs* 129: 1-199.
- King, F. W. 1962a. Systematics of Lesser Antillean lizards of the genus *Sphaerodactylus*. *Bull. Florida State Museum* 7: 1-52.
- King, F. W. 1962b. The occurrence of rafts for dispersal of land animals into the West Indies. *Q. J. Florida Acad. Sci.* 25: 46-52.
- Kirch, P. V., and T. L. Hunt (Editors). 1997. *Historical Ecology in the Pacific Islands. Prehistoric Environmental and Landscape Change.* Yale Univ. Press, New Haven, CT.
- Klauber, L. M. 1956. *Rattlesnakes: Their Habits, Life Histories, and Influence on Mankind.* Univ. California Press, Berkeley.
- Kluge, A. G. 1964. A revision of the South American gekkonid lizard genus *Homonota* Gray. *Novitates* 2193: 1-41.
- Kluge, A. G. 1969. The evolution and geographical origin of the New World *Hemidactylus mabouia-brookii* complex (Gekkonidae, Sauria). *Misc. Publ. Museum Zool. Univ. Michigan* 138: 1-78.

- Kluge, A. G. 1988. A concern for the evidence and a phylogenetic hypothesis of cladistic relationships among *Epicrates* (Boidae, Serpentes). *Syst. Zool.* 38: 7-25.
- Kluge, A. G. 1991. Boine snake phylogeny and research cycles. *Misc. Publ. Museum Zool. Univ. Michigan* 178: i-iv + 1-58.
- Kluge, A. G. 1995. Cladistic relationships of sphaerodactyl lizards. *Novitates* 3139: 1-23.
- Koza, J. R., J. Roughgarden, and J. P. Rice. 1992. Evolution of food-foraging strategies for the Caribbean *Anolis* lizard using genetic programming. *Adapt. Behav.* 1: 171-199.
- Lacépède, B.-G. - É. 1789. *Histoire Naturelle des Quadrupèdes Ovipares et des Serpens*. Hôtel de Thou, Paris.
- Lack, D. 1976. *Island Biology Illustrated by the Land Birds of Jamaica*. *Studies in Ecology* Vol. 3. Univ. of California Press, Berkeley.
- Laessle, A. M. 1961. A micro-limnological study of Jamaican bromeliads. *Ecology* 42: 499-517.
- Lahey, G. J., R. A. Sosa, S. M. Micco, E. J. Censky, and R. Powell. MS. On the natural history of *Leiocephalus barahonensis* (Lacertilia: Tropiduridae) from the Barahona Peninsula, Hispaniola.
- Lambert, M. R. K. 1986. Herpetology in Jamaica with general reference to conservation. *British Herpetol. Soc. Bull.* 18.
- Landwer, A. J., G. W. Ferguson, R. Herber, and M. Brewer. 1995. Habitat use of introduced and native anoles (Iguanidae: *Anolis*) along the northern coast of Jamaica. *Texas J. Sci.* 47: 45-52.
- Lannoo, M. J., D. S. Townsend, and R. J. Wassersug. 1987. Larval life in the leaves: arboreal tadpole types, with special attention to the morphology, ecology and behavior of the oophagous *Osteopilus brunneus* (Hylidae) larvae. *Fieldiana Zool.* 38: iv + 31.
- Laurenti, J. N. 1768. *Specimen Medicum, Exhibens Synopsin [sic] Reptilium Emendatam cum Experimentis circa Venena*, Univ. of Vienna, Vienna.

- Lawson, R. 1987. Molecular studies of *Thamnophiine* snakes: 1. The phylogeny of the genus *Nerodia*. *J. Herpetol.* 21: 140-157.
- Lazell, J. D., Jr. 1961. A new species of *Sphaerodactylus* from northern Haiti. *Breviora* 139: 1-5.
- Lazell, J. D., Jr. 1962. The anoles (Sauria: Iguanidae) of the eastern Caribbean, Part V. Geographic differentiation in *Anolis oculatus* on Dominica. *Bull. Museum Comp. Zool.* 127: 466-475.
- Lazell, J. D., Jr. 1964a. The anoles (Sauria: Iguanidae) of the Guadeloupéen archipelago. *Bull. Museum Comp. Zool.* 131: 359-401.
- Lazell, J. D., Jr. 1964b. The Lesser Antillean representatives of *Bothrops* and *Constrictor*. *Bull. Museum Comp. Zool.* 132: 245-273.
- Lazell, J. D., Jr. 1966. Contributions to the herpetology of Jamaica – Studies on *Anolis reconditus* Underwood and Williams. *Bull. Inst. Jamaica Sci. Ser.* 18: 5-15.
- Lazell, J. D., Jr. 1967. Wiederentdeckung von zwei angeblich ausgestorbenen Schlangenarten der westindischen Inseln. *Salamandra* 3: 91-97.
- Lazell, J. D., Jr. 1972. The anoles (Sauria, Iguanidae) of the Lesser Antilles. *Bull. Museum Comp. Zool.* 143: 1-115.
- Lazell, J. D., Jr. 1973. The lizard genus *Iguana* in the Lesser Antilles. *Bull. Museum Comp. Zool.* 145: 1-28.
- Lazell, J. D., Jr. 1983. Biogeography of the herpetofauna of the British Virgin Islands, with description of a new anole (Sauria: Iguanidae), pp. 99-117 in *Advances in Herpetology and Evolutionary Biology. Essays in Honor of Ernest E. Williams*, edited by A. G. J. Rhodin, and K. Miyata. *Museum Comp. Zool.*, Cambridge. MA.
- Lazell, J. D., Jr. 1991. The herpetofauna of Guana island: diversity, abundance, rarity, and conservation, pp. 28-33 in *Status y Distribución de los Reptiles y Anfibios de la Region de Puerto Rico*, edited by J. A. Moreno. *Depto. Rec. Natur.*, Puerto Rico.
- Lazell, J. D., Jr. 1992. The family Iguanidae: disagreement with Frost and Etheridge (1989). *Herpetol. Rev.* 23: 109-112.

- Lazell, J. D., Jr. 1993. Tortoise, cf. *Geochelone carbonaria*, from the Pleistocene of Anguilla, northern Lesser Antilles. *J. Herpetol.* 27: 485-486.
- Lazell, J. D., Jr. 1994. A new *Sphaerodactylus* (Sauria: Gekkonidae) from Bequia, Grenada Bank, Lesser Antilles. *Breviora* 496: 1-20.
- Lazell, J. D., Jr. 1996. Careening Island and the Goat Islands: evidence for the arid-insular invasion wave theory of dichopatric speciation in Jamaica, pp. 195-205 in *Contributions to West Indian Herpetology: a Tribute to Albert Schwartz*, edited by R. Powell, and R. W. Henderson. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.
- Lazell, J. D., Jr., and T. Sinclair. 1990a. *Anolis trinitatus*. *Herpetol. Rev.* 21: 96.
- Lazell, J. D., Jr., and T. Sinclair. 1990b. Geographic Distribution: *Gymnophthalmus underwoodi*. *Herpetol. Rev.* 21: 96.
- Lazell, J. D., Jr., and T. Sinclair. 1990c. Geographic Distribution: *Sphaerodactylus* cf. *vincenti*. *Herpetol. Rev.* 21: 97.
- Leclair, R., Jr., and G. Provencher. 1988. Demographic traits of a small sphaerodactyline lizard from Martinique, West Indies, pp. 127-128 in *Herpetologists' League-ASIH Annu. Meeting. (abstract)*
- Lee, J. C. 1996. Pre-columbian human influences on the biogeography of amphibians and reptiles in the Yucatan Peninsula, pp. 435-436 in *Contributions to West Indian Herpetology: A Tribute to Albert Schwartz*, edited by R. Powell, and R. W. Henderson. SSAR Contrib. Herpetol., Ithaca.
- Leidy, J. 1868. Notes on some vertebrate remains from the West Indian islands. *Proc. Acad. Natl. Sci. Philadelphia* 20: 178-180.
- Lenart, L. A., R. Powell, J. J. S. Parmerlee, D. D. Smith, and A. Lathrop. 1994. The diet and a gastric parasite of *Anolis armouri*, a cybotoid anole from montane pine forest in southern Hispaniola. *Herpetol. Nat. Hist.* 2: 97-100.
- Lenart, L. A., R. Powell, J. J. S. Parmerlee, A. Lathrop, and D. D. Smith. 1997. Anoline diversity in three differentially altered habitats in the Sierra de Baoruco, República Dominicana, Hispaniola. *Biotropica* 29: 117-123.

- Lescure, J. 1979. Étude taxonomique et eco-ethologique d'un amphibien des Petites Antilles: *Leptodactylus fallax* Müller, 1926 (Leptodactylidae). Bull. Museum Natur. Hist. Nat. Paris 4e sér., 1, sect A: 757-774.
- Lescure, J. 1987. Le peuplement en reptiles et amphibiens des Petites Antilles. Bull. Soc. Zool. France 112: 327-342.
- Levins, R., and H. Heatwole. 1963. On the distribution of organisms on islands. Carib. J. Sci. 3:173-177.
- Levins, R., and H. Heatwole. 1973. Biogeography of the Puerto Rican Bank: Introduction of species onto Palominos Island. Ecology 54: 1056-1064.
- Lewis, A. R. 1986. Body size and growth in two populations of the Puerto Rican ground lizard (Teiidae). J. Herpetol. 20: 190-195.
- Lewis, A. R. 1989. Diet selection and depression of prey abundance by an intensively foraging lizard. J. Herpetol. 23: 164-170.
- Lewis, A. R., and J. F. Saliva. 1987. Effects of sex and size on home range, dominance, and activity budgets in *Ameiva exsul* (Lacertilia: Teiidae). Herpetologica 43: 374-383.
- Lewis, A. R., G. Tirado, A. Aponte, J. Blanco, and J. Sepulveda. 1996. Social control of male reproductive success in *Ameiva exsul*, pp. 445-449 in Contributions to West Indian Herpetology: A Tribute to Albert Schwartz, edited by R. Powell, and R. W. Henderson. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.
- Lewis, C. B. 1941. The frogs of the Clydesdale region. Nat. Hist. Notes Nat. Hist. Soc. Jamaica 1: 10-12.
- Lewis, C. B. 1943. Frog eggs. Nat. Hist. Notes Nat. Hist. Soc. Jamaica 2: 35.
- Lewis, C. B. 1950. Gosse's illustration of Jamaican reptiles and amphibia. Nat. Hist. Notes Nat. Hist. Soc. Jamaica 4: 103.
- Lewis, J. F. 1980. Cenozoic tectonic evolution and sedimentation in Hispaniola, pp. 65-73 in Transactions of the Ninth Caribbean Geologic Conference. Amigo Hogar, Santo Domingo.

- Lewis, J. F., and G. Draper. 1990. Geology and tectonic evolution of the northern Caribbean margin, pp. 77-140 in *The Geology of North America*. Vol. H. The Caribbean Region, edited by G. Dengo, and J. E. Case. Geological Society America, Boulder, CO.
- Licht, P., and G. C. Gorman. 1970. Reproductive and fat cycles in Caribbean *Anolis* lizards. *Univ. California Pub. Zool.* 95: 1-52.
- Lichtenstein, H. 1856. *Nomenclatur Reptilium et Amphibiorum Musei Zoologici Berolinensis*. Königl. Akad. Wiss., Berlin.
- Lieberman, S. S. 1986. Ecology of the leaf litter herpetofauna of a Neotropical rain forest: La Selva Costa Rica. *Acta Zool. Mexicana* 15: 1-72.
- Lillywhite, H. B., and R. W. Henderson. 1993. Behavioral and functional ecology of arboreal snakes, pp. 1-48 in *Snakes: Ecology and Behavior*, edited by R. A. Seigel, and J. T. Collins. McGraw-Hill, NY.
- Linnaeus, C. 1758. *Systema Naturae per Regna Tria Naturae, Secundum Classes, Ordines, Genera, Species, cum Characteribus, Differentiis, Synonymis, Locis*. L. Salvii, Stockholm.
- Linnaeus, C. 1939. *Systema Naturae. Regnum Animale*. Trustees of the British Museum (Natural History), London. [Reprint].
- Losos, J. B. 1990a. Ecomorphology, performance capability, and scaling of West Indian *Anolis* lizards: an evolutionary analysis. *Ecol. Mongr.* 60: 369-388.
- Losos, J. B. 1990b. Concordant evolution of locomotor behaviour, display rate and morphology in *Anolis* lizards. *Anim. Behav.* 39: 879-890.
- Losos, J. B. 1990c. The evolution of form and function: morphology and locomotor performance in West Indian *Anolis* lizards. *Evolution* 44: 1189-1203.
- Losos, J. B. 1992a. The evolution of convergent structure in Caribbean *Anolis* communities. *Syst. Biol.* 41: 403-420.
- Losos, J. B. 1992b. A critical comparison of the taxon-cycle and character-displacement models for size evolution of *Anolis* lizards in the Lesser Antilles. *Copeia* 1992: 279-288.

- Losos, J. B. 1994. Integrative approaches to evolutionary ecology: *Anolis* lizards as model systems. *Annu. Rev. Ecol. Syst.* 25: 467-493.
- Losos, J. B., M. R. Gannon, W. J. Pfeiffer, and R. B. Waide. 1990. Notes on the ecology and behavior of *Anolis cuvieri* (Lacertilia: Iguanidae) in Puerto Rico. *Carib. J. Sci.* 26: 65-66.
- Losos, J. B., J. C. Marks, and T. W. Schoener. 1993. Habitat use and ecological interactions of an introduced and native species of *Anolis* lizard on Grand Cayman, with a review of the outcomes of anole introductions. *Oecologia* 95: 525-532.
- Losos, J. B., and B. Sinervo. 1989. The effects of morphology and perch diameter on sprint performance of *Anolis* lizards. *J. Exp. Biol.* 145: 23-30.
- Luckow, M. and A. Bruneau. 1997. Circularity and independence in phylogenetic tests of ecological hypotheses. *Cladistics* 13: 145-151.
- Lynch, J. D. 1965. A new species of *Eleutherodactylus* from Guadeloupe, West Indies. *Breviora* 220: 1-7.
- Lynch, J. D. 1966. The status of the tree frog, *Hyla barbudensis* Auffenberg, from Barbuda, British West Indies. *Copeia* 1966: 524-530.
- Lynch, J. D. 1971. Evolutionary relationships, osteology, and zoo-geography of leptodactyloid frogs. *Misc. Publ. Museum Nat. Hist. Univ. Kansas* 53: 1-238.
- Lynch, J. D. 1986. The definition of the Middle American clade of *Eleutherodactylus* based on jaw musculature (Amphibia: Leptodactylidae). *Herpetologica* 42: 248-258.
- Lynch, J. 1991. Three replacement names for preoccupied names in the genus *Eleutherodactylus* (Amphibia: Leptodactylidae). *Copeia* 1991(4): 1138-1139.
- Lynch, J. D. 1996. The relationships of the Hispaniolan frogs of the subgenus *Pelorius* (*Eleutherodactylus*: Leptodactylidae), pp. 141-155 in *Contributions to West Indian Herpetology: A Tribute to Albert Schwartz*, edited by R. Powell, and R. W. Henderson. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.

- Lynch, J., and W. E. Duellman. 1997. Frogs of the genus *Eleutherodactylus* (Leptodactylidae) in Western Ecuador: systematics, ecology, biogeography. Univ. Kansas Nat. Hist. Museum Spec. Publ. 23: 1-236.
- Lynn, W. G. 1936. A study of the thyroid in embryos of *Eleutherodactylus nubicola*. Anat. Rec. 64: 525-535.
- Lynn, W. G. 1937. Two new frogs from Jamaica. Herpetologica 1: 88-90.
- Lynn, W. G. 1940. The herpetology of Jamaica I. Amphibians. Bull. Inst. Jamaica Sci. Ser. 1: 1-60.
- Lynn, W. G. 1942. The embryology of *Eleutherodactylus nubicola*, an anuran which has no tadpole stage. Carnegie Inst. Washington Publ. 541: 27-62.
- Lynn, W. G. 1944. Duplicitas anterior in the toad, *Eleutherodactylus alticola*. Anat. Rec. 89: 345-351.
- Lynn, W. G. 1954. Description of a new frog of the genus *Eleutherodactylus* from Jamaica. J. Washington Acad. Sci. 44: 400-402.
- Lynn, W. G. 1958. Some amphibians from Haiti and a new subspecies of *Eleutherodactylus schmidtii*. Herpetologica 14: 153-157.
- Lynn, W. G., and J. N. Dent. 1942. A new frog of the genus *Eleutherodactylus* from Jamaica. Herpetologica 2: 71-74.
- Lynn, W. G., and J. N. Dent. 1943. Notes on Jamaican amphibians. Copeia 1943: 234-242.
- Lynn, W. G., and C. Grant. 1940. The herpetology of Jamaica. Bull. Inst. Jamaica Sci. Ser. 1: 1-148.
- Lynn, W. G., and A. M. Peadon. 1955. The role of the thyroid gland in direct development in the anuran, *Eleutherodactylus martinicensis*. Growth 19: 263-286.
- Lynn, W. G., and G. A. Walsh. 1957. The morphology of the thyroid gland in the Lacertilia. Herpetologica 13: 157-172.

- Lynxwiler, J. R., R. Powell, J. J. S. Parmerlee, A. Lathrop, and D. D. Smith. 1991. Notes on the natural history of *Hemidactylus brookii haitianus* (Sauria: Gekkonidae) from the Dominican Republic. *Dactylus* 1: 1-9.
- MacArthur, R. H., J. M. Diamond, and J. R. Karr. 1972. Density compensation in island faunas. *Ecology* 53: 330-342.
- MacArthur, R. H., and E. O. Wilson. 1963. An equilibrium theory of insular zoogeography. *Evolution* 17: 373-387.
- MacArthur, R. H., and E. O. Wilson. 1967. *The Theory of Island Biogeography*. Princeton Univ. Press, Princeton, NJ.
- MacLean, W. P., and R. D. Holt. 1979. Distributional patterns in St. Croix *Sphaerodactylus* lizards: the taxon cycle in action. *Biotropica* 11: 189-195.
- MacLean, W. P., R. Kellner, and H. Dennis. 1977. Island lists of West Indian amphibians and reptiles. *Smithsonian Herpetol. Information Service* 40: 1-47.
- MacLeay, W. S. 1834. A few remarks tending to illustrate the natural history of two annulose genera, namely *Urania* of Babricus and *Mygale* of Walckenäer. *Proc. Zool. Soc. London* 1834: 9-12.
- MacPhee, R. D. E., D. C. Ford, and D. A. MacFarlane. 1989. Pre-Wisconsinan mammals from Jamaica and models of late Quarternary extinctions in the Caribbean. *Anat. Res.* 31: 94-106.
- MacPhee, R. D. E., and R. A. Marx. 1997. The 40,000-year plague. Humans, hyperdisease and first-contact extinctions, pp. 169-217 in *Natural Change and Human Impact in Madagascar*, edited by S. M. Goodman, and B. D. Patterson. Smithsonian Instit. Press, Washington D. C.
- Maglio, V. J. 1970. West Indian xenodontine colubrid snakes: their probable origin, phylogeny, and zoogeography. *Bull. Museum Comp. Zool.* 141: 1-54.
- Mahon, R., and K. Aiken. 1977. The establishment of the North American bullfrog, *Rana catesbeiana* (Amphibia, Anura, Ranidae) in Jamaica. *J. Herpetol.* 11: 197-199.
- Major, R. H. 1870. *Selected Letters of Columbus*. Hakluyt Soc., London.

- Malfait, B. T., and M. G. Dinkelman. 1972. Circum-Caribbean tectonic and igneous activity and the evolution of the Caribbean plate. *Bull. Geol. Soc. Am.* 83: 251-272.
- Malhotra, A., and R. S. Thorpe. 1991. Experimental detection of rapid evolutionary response in natural island populations. *Nature* 353: 347-348.
- Malnate, E. V. 1971. A catalog of primary types in the herpetological collections of the Academy of Natural Sciences, Philadelphia (ANSP). *Proc. Acad. Natl. Sci. Philadelphia* 123: 345-375.
- Margulis, L., and T. H. Kunz. 1984. Glimpses of biological research and education in Cuba. *Bioscience* 84: 634-639.
- Martin. 1838. Report on a collection of snakes procured by the Euphrates Expedition. *Proc. Zool. Soc. London* VI: 81-84.
- Martin, P. S., and R. Klein (Editors). 1984. *Quaternary Extinctions: A Prehistoric Revolution*. Univ. Arizona Press, Tucson.
- Martin, P. S., and D. W. Steadman. 1998. Prehistoric extinctions on islands and continents, in *Quaternary Extinctions* edited by R. D. E. MacPhee et al., Plenum Press, New York.
- Martin, R. F. 1972. Evidence from osteology pp. 37-70 in *Evolution in the genus Bufo*, edited by W. F. Blair. Univ. Texas Press, Austin, Texas.
- Mateo, J., and M. Acevedo. 1989. 5. Regionalización físico-geográfica, 1:3000000. XII Paisajes. *Nuevo Atlas Nacional de Cuba*. Academia de Ciencias de Cuba, Cuba.
- Matthew, W. D. 1915. *Climate and Evolution*. *Ann. New York Acad. Sci.* 24: 171-318.
- Maurrasse, F. J.-M. R. 1982. *Survey of the Geology of Haiti*. Miami Geol. Soc., Miami.
- McDiarmid, R. W., T. Touré, and J. M. Savage. 1996. The proper name of the neotropical tree boa often referred to as *Corallus enydris* (Serpentes: Boidae). *J. Herpetol.* 30: 320-326.

- McDowell, S. B. 1964. Partition of the genus *Clemmys* and related problems in the taxonomy of aquatic Testudinidae. Proc. Zool. Soc. London 143: 239-279.
- McDowell, S. B. 1979. A catalogue of the snakes of New Guinea and the Solomons, with special reference to those in the Bernice P. Bishop Museum. Part III. Boinae and Acrochordoidea (Reptilia, Serpentes). J. Herpetol. 13: 1-92.
- McKenna, M. C. 1973. Sweepstakes, filters, corridors, Noah's arks and beached Viking funeral ships in paleobiogeography, pp. 295-308 in Implications of Continental Drift to the Earth Sciences, edited by D. H. Tarling, and S. K. Runcorn. Academic Press, New York.
- McLaughlin, J. F., and J. Roughgarden. 1989. Avian predation on *Anolis* lizards in the northeastern Caribbean: an inter-island contrast. Ecology 70: 617-628.
- McLaughlin, P. P., W. A. van den Bold, and P. Mann. 1991. Geology of the Azua and Enriquillo basins, Dominican Republic; 1, Neogene lithofacies, biostratigraphy, biofacies, and paleogeography. Geol. Soc. Am., Spec. Pap. 262: 337-366.
- Meerwarth, H. 1901. Die westindischen Reptilien und Batrachier des naturhistorischen Museums in Hamburg. Mitt. Naturhist. Museum Hamburg 18: 1-41.
- Merrem, B. 1809. Amphibiologische Beiträge. Annu. Wetterausischen Gesell. Ges. Naturk. I: 1-191.
- Merrem, B. 1820. Tentamen Systematis Amphibiorum. Marburg.
- Mertens, R. 1938. Amphibien und Reptilien aus Santo Domingo, gesammelt von Prof. Dr. H. Böker. Senckenberg. Biol. 20: 332-342.
- Mertens, R. 1939. Herpetologische Ergebnisse einer Reise nach der Insel Hispaniola, Westindien. Abh. Senckenberg. Nat. Ges. 449: 1-84.
- Mertens, R. 1940. Zoolog Reisen auf der Insel Hispaniola (from Teirleben der Tropen by R. Mertens), pp. 155-241 Verlag Dr. Waldemar Kramer, Frankfurt.
- Mertens, R. 1950. Zur Taxonomie und Verbreitung der hispaniolischen Eidechsen der Gattung *Ameiva*. Senckenbergiana 31: 3-7.

- Mertens, R., and H. Wermuth. 1955. Die rezenten Schildkröten, Krokodile und Brückenechsen. Zool. Jahrb. Syst. Abt. 83: 323-440.
- Meshaka, W. E., Jr. 1996. Diet and the colonization of buildings by the Cuban treefrog, *Osteopilus septentrionalis* (Anura: Hylidae). Carib. J. Sci. 32.
- Meshaka, W. E., Jr., B. P. Butterfield, and B. Hauge. 1994a. *Hemidactylus mabouia* as an established member of the Florida herpetofauna. Herpetol. Rev. 25: 80-81.
- Meshaka, W. E., Jr., B. P. Butterfield, and B. Hauge. 1994b. Reproductive notes on the introduced gecko *Hemidactylus mabouia* in southern Florida. Herpetol. Nat. Hist. 2: 109-110.
- Meshaka, W. E., Jr., B. P. Butterfield, and B. Hauge. 1994c. Geographic distribution: *Hemidactylus mabouia*. Herpetol. Rev. 25: 165.
- Miles, D. B., and A. E. Dunham. 1996. The paradox of the phylogeny: character displacement of analyses of body size in island *Anolis*. Evolution 50: 594-603.
- Mittermeier, R. A. 1972. Jamaica's endangered species. Oryx 11: 258-262.
- Moermond, T. C. 1979. Habitat constraints on the behavior, morphology, and community structure of *Anolis* lizards. Ecology 60: 152-164.
- Monroe, W. H. 1976. The karst landforms of Puerto Rico. Geol. Surv. Prof. Paper 899: iv + 69.
- Moore, C. 1991. Cabaret: lithic workshop sites in Haiti. pp. 92-104 in Proceedings of the 13 International congress of Caribbean Archeology, edited by E. N. Ayubi, and J. B. Haviser. Reports Archeological-Anthropological Institute, Netherlands Antilles 9.
- Moreau de Jonnés, A. 1818. Monographie du mabouia des murailles, ou Gecko Mabouia de Antilles. Bull. Sci. Soc. Philomath. Paris 3: 138-139.
- Morgan, G. S., and C. A. Woods. 1986. Extinction and the zoogeography of West Indian land mammals. Biol. J. Linnean Soc. London 28: 167-203.
- Morison, S. E. 1963. Journals and Other Documents on the Life and Voyages of Christopher Columbus. Heritage Press, NY.

- Morrison, L. W. 1997. The insular biogeography of small Bahamian cays. *J. Ecol.* 85:441-454.
- Moster, J. A., R. Powell, J. J. S. Parmerlee, D. D. Smith, and A. Lathrop. 1992. Natural history notes on a small population of *Anolis brevirostris* (Sauria: Polychridae) from altered habitat in the Dominican Republic. *Bull. Maryland Herpetol. Soc.* 28: 150-161.
- Müller, L. 1923. Neue oder seltene Reptilien und Batrachier der Zoologischen Sammlung des bayr. Staates. *Zool. Anz.* 57: 49-61.
- Müller, L. 1926. *Leptodactylus fallax* nom. nov. fr *Leptodactylus dominicensis* L. Mill. (non Cochran). *Zool. Anz.* 65: 200.
- Murphy, J. C. 1996. Crossing Bond's Line: the herpetofaunal exchange between the eastern Caribbean and mainland South America, pp. 207-216 in *Contributions to West Indian Herpetology: A Tribute to Albert Schwartz*, edited by R. Powell, and R. W. Henderson. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.
- Murphy, J. C., H. K. Voris, and D. R. Karns. 1994. A field guide to the snakes of the Danum Valley, a Bornean tropical forest ecosystem. *Bull. Chicago Herpetol. Soc.* 29: 133-151.
- Murphy, R. W. The phylogenetic analysis of allozyme data: invalidity of coding alleles by presence/absence and recommended procedures. *Biochem. Syst. Ecol.* 21: 25-38.
- Murphy, R. W., F. C. McCollum, G. C. Gorman, and R. Thomas. 1984. Genetics of hybridizing populations of Puerto Rican *Sphaerodactylus*. *J. Herpetol.* 18: 93-105.
- Myers, C. W. 1973a. A new genus for Andean snakes related to *Lygophis boursieri* and a new species (Colubridae). *Novitates* 2522: 1-37.
- Myers, C. W. 1973b. Anguid lizards of the genus *Diploglossus* in Panama, with the description of a new species. *Novitates* 2523: 1-20.
- Myers, C. W., and J. A. Campbell. 1981. A new genus and species of colubrid snake from the Sierra Madre del Sur of Guerrero, Mexico. *Novitates* 2708: 1-20.

- Myers, G. S. 1962. The American leptodactylid frog genera *Eleutherodactylus*, *Hylodes* (= *Elosia*), and *Caudiverbera* (= *Calyptocephalus*). *Copeia* 1962: 195-202.
- Nelson, G. 1969. The problem of historical biogeography. *Syst. Zool.* 18: 243-246.
- Nelson, G. 1973. Comments on Leon Croizat's biogeography. *Syst. Zool.* 22: 312-320.
- Nelson, G. 1974. Historical biogeography: an alternative formalization. *Syst. Zool.* 23: 555-558.
- Nganuma, K. H., and J. D. Roughgarden. 1990. Optimal body size in Lesser Antillean *Anolis* lizards – a mechanistic approach. *Ecol. Monogr.* 60: 239-256.
- Noble, G. K. 1921. The bony structure and phyletic relations of *Sphaerodactylus* and allied lacertilian genera, with the description of a new genus. *Novitates* 4: 1-16.
- Noble, G. K. 1923a. Six new batrachians from the Dominican Republic. *Novitates* 61: 1-6.
- Noble, G. K. 1923b. Four new lizards from Beata Island, Dominican Republic. *Novitates* 64: 1-5.
- Noble, G. K. 1927. The value of life history data in the study of the evolution of the Amphibia. *Annu. New York Acad. Sci.* 30:31-128.
- Noble, G. K. 1933. The mating behavior of lizards; its sexual bearing on the theory of sexual selection. *Annu. New York Acad. Sci.* 35: 25-100.
- Noble, G. K., and W. G. Hassler. 1933. Two new species of frogs, five new species and a new race of lizards from the Dominican Republic. *Novitates* 652: 1-17.
- Noble, G. K., and G. C. Klingel. 1932. The reptiles of Great Inagua Island, British West Indies. *Novitates* 549: 1-25.
- Norell, M. A. and K. de Queiroz. 1991. The earliest iguanine lizard (Reptilia: Squamata) and its bearing on iguanine phylogeny. *Novitates* 2997: 1-16.

- Ober, F. A. 1880. Camps in the Caribees: the adventures of a naturalist in the Lesser Antilles. Lee & Shepard, Boston.
- Oliver, J. A. 1948. The anoline lizards of Bimini, Bahamas. *Novitates* 1383: 1-36.
- Oliver, W. L. R. 1982. The coney and the yellow snake: the distribution and status of the Jamaican hutia *Geocapromys brownii* and the Jamaican boa *Epicrates subflavus*. *Dodo J. Jersey Wildl. Pres. Trust* 19: 6-33.
- Olson, R. E. 1995. Microhabitat in ecologically devastated areas. *Bull. Maryland Herpetol. Soc.* 31: 93-95.
- Olson, S. L., and W. B. Hilgartner. 1982. Fossil and subfossil birds from the Bahamas, pp. 22-60 in *Fossil Vertebrates from the Bahamas*, edited by S. L. Olson. *Smithsonian Contributions Paleobiol.* 48. Smithsonian Institution, Washington, DC.
- Olson, S. L., G. K. Pregill, and W. B. Hilgartner. 1990. Studies on fossil and extant vertebrates from San Salvador (Watling's) Island, Bahamas. *Smithsonian Contrib. Zool.* 508:1-15.
- Osborn, H. F. 1931. *Cope: Master Naturalist*. Princeton Univ. Press, Princeton, NJ.
- Ottenwalder, J. A. 1989. A summary of conservation trends in the Dominican Republic, pp. 845-850 in *Biogeography of the West Indies: Past, Present, and Future*, edited by C. A. Woods. Sandhill Crane Press, Gainesville.
- Ottenwalder, J. A. 1996. The current status of sea turtles in Haiti, pp. 381-393 in *Contributions to West Indian Herpetology: A Tribute to Albert Schwartz*, edited by R. Powell, and R. W. Henderson. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.
- Oviedo, G. F. de. in Stoudmere, S. A. (Trans. & Ed.). 1959. *Natural History of the West Indies*. Univ. North Carolina Press, Chapel Hill.
- Pacala, S. W., and J. Roughgarden. 1982. Resource partitioning and interspecific competition in two-species insular *Anolis* lizard communities. *Science* 217: 444-446.

- Pacala, S. W., and J. Roughgarden. 1985. Population experiments with the *Anolis* lizards of St. Maarten and St. Eustatius. *Ecology* 66: 129-141.
- Page, R. D. M., and C. Lydeard. 1994. Towards a cladistic biogeography of the Caribbean. *Cladistics* 10: 21-41.
- Panton, E. S. 1952. Our ground and tree frogs – glimpses into their life and habits. *Nat. Hist. Notes Nat. Hist. Soc. Jamaica* 5: 87-92 + 94.
- Parker, H. W. 1926a [August]. A new brachycephalid frog from Brazil. *Annu. Mag. Nat. Hist.* (9)18: 201-203.
- Parker, H. W. 1926b. The neotropical lizards of the genera *Lepidoblepharis*, *Pseudogonatodes*, *Lathrogecko*, and *Sphaerodactylus*, with the description of a new genus. *Annu. Mag. Nat. Hist.* 17: 291-301.
- Parker, H. W. 1933. Some amphibians and reptiles from the Lesser Antilles. *Annu. Mag. Nat. Hist.* 10: 155-158.
- Parker, H. W. 1935. Some lizards from Venezuela and the Dutch Leeward Islands. *Annu. Mag. Nat. Hist.* 15: 480-484.
- Parker, H. W. 1936. Some extinct snakes of the West Indies. *Annu. Mag. Nat. Hist.* 10: 227-233.
- Parsons, K. J. 1995. *Peltophryne guentheri* (NCN). *Diet. Herpetol. Rev.* 26: 202.
- Paull, D., E. E. Williams, and W. P. Hall. 1976. Lizard karyotypes from the Galapagos islands: chromosomes in phylogeny and evolution. *Breviora Museum Comp. Zool.* 441: 1-31.
- Perfit, M. R., and E. E. Williams. 1989. Geological constraints and biological retrodictions in the evolution of the Caribbean sea and its islands, pp. 47-102 in *Biogeography of the West Indies, Past, Present and Future*, edited by C. A. Woods. Sandhill Crane Press, Gainesville.
- Perkins, L. 1942. Whistling frogs in St. Annu. *Nat. Hist. Notes Nat. Hist. Soc. Jamaica* 1: 4-5.

- Peters, G., and A. Schwartz. 1972. Ein neuer, bromelien bewohnder Kugelfingergecko (Gekkonidae, *Sphaerodactylus*) aus Oriente Cuba. Mitteilungen aus dem Zool. Museum Berlin 48: 393-399.
- Peters, J. A., and R. Donoso-Barros. 1970. Catalogue of the Neotropical Squamata. Part II. Lizards and amphisbaenians. Bull. U.S. Natl. Museum 297: 1-293.
- Peters, J. A., and R. Donoso-Barros. 1986. Catalogue of the Neotropical Squamata. Part II. Lizards and Amphisbaenians. Smithsonian Institute Press, Washington, DC.
- Peters, J. A., and B. Orejas-Miranda. 1970. Catalogue of the Neotropical Squamata. Part I. Snakes. Bull. U.S. Natl. Museum 297: viii+347.
- Peters, W. 1863. Über einige neue oder weniger bekannte Schlangenarten des zoologischen Museums zu Berlin. Monatsb. Königl. Akad. Wiss. Berlin 1863: 272-289.
- Peters, W. 1869. Mitteilung über neue Gattungen und Arten von Eidechsen. Monatsb. Königl. Akad. Wiss. Berlin: 57-66.
- Peters, W. 1879. Neue Amphibien des Kgl. zoologischen Museums (*Euprepes*, *Acontias*, *Typhlops*, *Zamenis*, *Spilotes*, *Oedipus*). Monatsb. Königl. Akad. Wiss. Berlin.
- Peterson, J. A. 1983a. The evolution of the subdigital pad in *Anolis*. I. Comparisons among the anoline genera, pp. 245-283 in *Advances in Herpetology and Evolutionary Biology*, edited by A. G. J. Rhodin, and K. Miyata. Museum of Comparative Zoology, Cambridge, MA.
- Peterson, J. A. 1983b. The evolution of the subdigital pad of *Anolis* 2. Comparisons among the iguanid genera related to the anolines and a view from outside the radiation. *J. Herpetol.* 17: 371-397.
- Peterson, J. A., and R. L. Bezy. 1985. The microstructure and evolution of scale surfaces in xantusiid lizards. *Herpetologica* 41: 298-324.
- Pfeiffer, W. J. 1996. Litter invertebrates, pp. 137-181 in *The Food Web of a Tropical Rain Forest*, edited by D. P. Reagan, and R. B. Waide. Univ. Chicago Press, Chicago.

- Pindell, J., and S. F. Barrett. 1990. Geological evolution of the Caribbean region: a plate tectonic perspective, pp. 405-432 in *The Geology of North America*. Vol. H. The Caribbean Region, edited by G. Dengo, and J. E. Case. Geol. Soc. Am., Inc., New York.
- Pinou, T., and H. G. Dowling. 1994. The phylogenetic relationships of the Central American snake *Tretanorhinus*: data from morphology and karyology. *Amphib.-Rept.* 15: 297-305.
- Pitman, W. C., III, S. Cande, J. Labreque, and J. Pindell. 1993. Separation of Africa from South America, pp. 15-36 in *Biological Relations between Africa and South America*, edited by P. Goldblatt. Yale Univ. Press, New Haven, CT.
- Poe, S. 1996. Data set incongruence and the phylogeny of crocodylians. *Syst. Biol.* 45: 393-414.
- Poe, S. 1998. Skull characters and the cladistic relationships of the Hispaniolan dwarf twig *Anolis*. *Herp. Mong.* 12: 192-236.
- Poey, F. 1851-55. Circulacion del cocodrilo. *Mem. Sobre Hist. Nat. Cuba* 1: 258-267.
- Poinar, G. O., Jr. 1992. *Life in Amber*. Stanford Univ. Press, Stanford, CA.
- Poinar, G. O., Jr., and D. C. Cannatella. 1987. An Upper Eocene frog from the Dominican Republic and its implications for Caribbean biogeography. *Science* 237: 1215-1216.
- Pough, F. H., M. M. Stewart, and R. G. Thomas. 1977. Physiological basis of habitat partitioning in Jamaican *Eleutherodactylus*. *Oecologia* 27: 285-293.
- Pounds, J. A. 1988. Ecomorphology, locomotion, and microhabitat structure: patterns in a tropical mainland *Anolis* community. *Ecol. Monogr.* 58: 299-320.
- Powell, G. L., and A. P. Russell. 1992. Locomotor correlates of ecomorph designation in *Anolis*: an examination of three sympatric species from Jamaica. *Canadian J. Zool.* 70: 725-739.
- Powell, R. 1992a. *Peltophryne guentheri*. *Cat. Am. Amphib. Rept.* 535: 1-4.
- Powell, R. 1992b. *Amphisbaena gonavensis*. *Cat. Am. Amphib. Rept.* 539: 1-3.

- Powell, R. 1993. Comments on the taxonomic arrangement of some Hispaniolan amphibians and reptiles. *Herpetol. Rev.* 24: 135-137.
- Powell, R., S. S. Duer, J. J. S. Parmerlee, and D. D. Smith. 1989b. Life history notes: *Ameiva taeniura* (food habits). *Herpetol. Rev.* 20: 49.
- Powell, R., and R. W. Henderson. 1992. Life history notes: *Anolis gingivinus* (nocturnal activity). *Herpetol. Rev.* 23: 117.
- Powell, R., and R. W. Henderson. 1994. *Ialtris*. *Cat. Am. Amphib. Rept.* 590.1-590.2.
- Powell, R., and R. W. Henderson. 1996a. Introduction, pp. 11-14 in *Contributions to West Indian Herpetology: A Tribute to Albert Schwartz*, edited by R. Powell, and R. W. Henderson. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.
- Powell, R., and R. W. Henderson. 1996b. A brief history of West Indian herpetology, pp. 29-50 in *Contributions to West Indian Herpetology: A Tribute to Albert Schwartz*, edited by R. Powell, and R. W. Henderson. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.
- Powell, R., and R. W. Henderson. 1998. *Alsophis melanichnus*. *Cat. Amer. Amphib. Rept.* 660:1-2.
- Powell, R., R. W. Henderson, K. Adler, and H. A. Dundee. 1996a. An annotated checklist of West Indian amphibians and reptiles, pp. 51-93 in *Contributions to West Indian Herpetology: A Tribute to Albert Schwartz*, edited by R. Powell, and R. W. Henderson. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.
- Powell, R., and J. J. S. Parmerlee. 1992. In the spotlight: *Aristelliger lar*. *Dactylus* 1: 37-38.
- Powell, R., and J. J. S. Parmerlee. 1993. In the spotlight: *Hemidactylus haitianus*. *Dactylus* 2: 54-55.
- Powell, R., J. J. S. Parmerlee, M. A. Rice, and D. D. Smith. 1990a. Ecological observations of *Hemidactylus brookii haetianus* Meerwarth (Sauria: Gekkonidae) from Hispaniola. *Carib. J. Sci.* 26: 67-70.

- Powell, R., J. J. S. Parmerlee, and D. D. Smith. 1996b. Evidence of spatial niche partitioning by a Hispaniolan lizard community in a xeric habitat, pp. 317-326 in Contributions to West Indian Herpetology: A Tribute to Albert Schwartz, edited by R. Powell, and R. W. Henderson. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.
- Powell, R., R. J. Passaro, and R. W. Henderson. 1992. Noteworthy herpetological records from Saint Maarten, Netherlands Antilles. *Carib. J. Sci.* 28: 234-235.
- Powell, R., D. D. Smith, J. S. Parmerlee, C. V. Taylor, and M. L. Jolley. 1990b. Range expansion by an introduced anole: *Anolis porcatus* in the Dominican Republic. *Amphib.-Rept.* 11: 421-425.
- Powell, R., S. G. Thornhill, and D. G. [sic] Smith. 1989a. Geographic distribution: *Sphaerodactylus a. altavelensis*. *Herpetol. Rev.* 20: 13.
- Pregill, G. K. 1981a. Cranial morphology and the evolution of West Indian toads (Salientia: Bufonidae): resurrection of the genus *Peltophryne* Fitzinger. *Copeia* 1981: 273-285.
- Pregill, G. K. 1981b. Late Pleistocene herpetofaunas from Puerto Rico. *Misc. Publ. Univ. Kansas Museum Nat. Hist.* 71: 1-72.
- Pregill, G. K. 1981c. An appraisal of the vicariance hypothesis of Caribbean biogeography and its application to West Indian terrestrial vertebrates. *Syst. Zool.* 30: 147-155.
- Pregill, G. K. 1984. An extinct species of *Leiocephalus* from Haiti (Sauria: Iguanidae). *Proc. Biol. Soc. Washington* 97: 827-833.
- Pregill, G. K. 1986. Body size of insular lizards: a pattern of Holocene dwarfism. *Evolution* 40: 997-1008.
- Pregill, G. K. 1992. Systematics of the West Indian lizard genus *Leiocephalus* (Squamata: Iguania: Tropicuridae). *Univ. Kansas Museum Nat. Hist. Misc. Publ.* 84: 1-69.
- Pregill, G. K., R. I. Crombie, D. W. Steadman, L. K. Gordon, F. W. Davis, and W. B. Hilgartner. 1992. Living and late Holocene fossil vertebrates, and the vegetation of the Cockpit Country, Jamaica. *Atoll. Res. Bull.* 353: 1-19.

- Pregill, G. K., and T. Dye. 1989. Prehistoric extinction of giant iguanas in Tonga. *Copeia* 1989: 505-508.
- Pregill, G. K., and S. L. Olson. 1981. Zoogeography of the West Indies in relation to Pleistocene climatic cycles. *Annu. Rev. Ecol. Syst.* 12: 75-98.
- Pregill, G. K., D. W. Steadman, S. L. Olson, and F. V. Grady. 1988. Late Holocene fossil vertebrates from Burma Quarry, Antigua, Lesser Antilles. *Smithson. Contrib. Zool.* 463: 1-27.
- Pregill, G. K., D. W. Steadman, and D. R. Watters. 1994. Late Quaternary vertebrate faunas of the Lesser Antilles: historical components of Caribbean biogeography. *Bull. Carnegie Museum Nat. Hist.* 30: 1-51.
- Presch, W. 1971. Tongue structure of the teiid lizard genera *Ameiva* and *Cnemidophorus* with a reallocation of *Ameiva vanzoi*. *J. Herpetol.* 5: 183-185.
- Presch, W. 1988. Phylogenetic relationships of the Scincomorpha, pp. 471-487 in *Phylogenetic Relationships of the Lizard Families: Essays Commemorating Charles L. Camp*, edited by R. Estes, and G. Pregill. Stanford Univ. Press, Stanford, CA.
- Pritchard, P. C. H. 1979. *Encyclopedia of Turtles*. T. F. H., Neptune, NJ.
- Pritchard, P. C. H. and P. Trebbau. 1984. *The Turtles of Venezuela*. Soc. Stud. Amphib. Rept. Contrib. Herpetol. 3. Athens, Ohio.
- Proctor, G. R. 1959. Observations on Navassa Island. *Geonotes Q. J. Jamaica Group Geologists' Assoc.* 2: 49-51.
- Pruna Goodgall, P. M. 1994. *Cronologia: Hechos historicos relacionados con la ciencia y la tecnologia acaecidos en la Habana, 1521-1988*. Editorial Academia, La Habana.
- Rage, J. C. 1982. La phylogénie des lépidosauriens (Reptilia): une approche cladistique. *C. R. Acad. Sci. Paris* 294: 563-566.
- Rand, A. S. 1961. Notes on Hispaniolan herpetology 4. *Anolis koopmani*, new species, from the southwestern peninsula of Haiti. *Breviora* 137: 1-4.
- Rand, A. S. 1962. Notes on Hispaniolan herpetology. 5. The natural history of three sympatric species of *Anolis*. *Breviora* 154: 1-15.

- Rand, A. S. 1964a. Inverse relationship between temperature and shyness in the lizard *Anolis lineatopus*. *Ecology* 45: 863-864.
- Rand, A. S. 1964b. Ecological distribution in anoline lizards of Puerto Rico. *Ecology* 45: 745-752.
- Rand, A. S. 1967a. The ecological distribution of the anoline lizards around Kingston, Jamaica. *Breviora* 272: 1-18.
- Rand, A. S. 1967b. Ecology and social organization in the iguanid lizard *Anolis lineatopus*. *Proc. U.S. Natl. Museum* 122: 1-79.
- Rand, A. S. 1969. Competitive exclusion among anoles (Sauria: Iguanidae) on small islands in the West Indies. *Breviora Museum Comp. Zool.* 319: 1-16.
- Rand, A. S., and S. S. Humphrey. 1968. Interspecific competition in the tropical rain forest: ecological distribution among lizards at Belém, Pará. *Proc. U.S. Natl. Museum* 125: 1-17.
- Rand, A. S., and C. W. Myers. 1990. The herpetofauna of Barro Colorado Island, Panama: an ecological summary, pp. 386-409 in *Four Neotropical Rainforests*, edited by A. H. Gentry. Yale Univ. Press, New Haven, CT.
- Rand, A. S., and E. E. Williams. 1969. The anoles of La Palma: aspects of their ecological relationships. *Breviora* 327: 1-17.
- Ray, C. E. 1964. A small assemblage of vertebrate fossils from Spring Bay, Barbados. *J. Barbados Museum Hist. Soc.* 31: 11-22.
- Reagan, D. P. 1992. Congeneric species distribution and abundance in a three-dimensional habitat: the rain forest anoles of Puerto Rico. *Copeia* 1992: 392-403.
- Reagan, D. P. 1996a. The role of amphibians and reptiles in a West Indian rain forest food web, pp. 217-227 in *Contributions to West Indian Herpetology: A Tribute to Albert Schwartz*, edited by R. Powell, and R. W. Henderson. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.
- Reagan, D. P. 1996b. Anoline lizards, pp. 321-345 in *The Food Web of a Tropical Rain Forest*, edited by D. P. Reagan, and R. B. Waide. Univ. Chicago Press, Chicago, IL.

- Reagan, D. P. and R. B. Waide (eds.) 1996. The food web of a tropical rain forest. Univ. Chicago Press, Chicago, IL.
- Reinhardt, J., and C. F. Lütken. 1862. Bidrag til det vestindiske Öeriges og navnlig til de dansk-vestindiske Öers Herpetologie. Vidensk. Meddel. Naturhist. Foren. Kjøbenhavn 10-18: 153-291.
- Reitz, E. J., L. A. Newsome, and S. J. Scudder (Editors). 1996. Case Studies in Environmental Archaeology. Plenum, NY.
- Richmond, N. D. 1964. The blind snakes (*Typhlops*) of Haiti with descriptions of three new species. *Breviora* 202: 1-12.
- Richmond, N. D. 1966. The blind snakes, *Typhlops*, of Guadeloupe and Dominica with the description of a new species. *Herpetologica* 22: 129-132.
- Ricklefs, R. 1970. Stage of taxon cycle and distribution of birds on Jamaica. *Evolution* 24: 475-477.
- Ricklefs, R., and G. W. Cox. 1972. Taxon cycles in the West Indian avifauna. *Am. Nat.* 106: 195-219.
- Ricklefs, R., and G. W. Cox. 1978. Stage of taxon cycle, habitat distribution, and population density in the avifauna of the West Indies. *Am. Nat.* 112: 875-895.
- Rieppel, O. 1980. Green anole in Dominican amber. *Nature* 286: 486-487.
- Rivero, J. A. 1978. Los anfibios y reptiles de Puerto Rico/The amphibians and reptiles of Puerto Rico. Univ. of Puerto Rico Press, San Juan.
- Rodda, G., E. W. Campbell III, and T. H. Fritts. 1995. Extraordinary lizard densities on Guam, as revealed by complete removal sampling, Annu. Meetg. ASIH, AES, HL., Univ. of Alberta, Canada. (abstracts)
- Rodriguez, G. A., and D. P. Reagan. 1984. Bat predation by the Puerto Rican boa, *Epicrates inornatus*. *Copeia* 1984: 219-220.
- Rodríguez-Robles, J. A., and H. W. Greene. 1996. Ecological patterns in Greater Antillean macrostomatan snake assemblages, with comments on body size evolution in *Epicrates* (Boidae), pp. 339-357 in Contributions to West In-

- dian Herpetology: A Tribute to Albert Schwartz, edited by R. Powell, and R. W. Henderson. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.
- Rodríguez-Robles, J. A., and M. Leal. 1993. Effects of prey type on the feeding behavior of *Alsophis portoricensis* (Serpentes: Colubridae). *J. Herpetol.* 27: 163-168.
- Rodríguez-Schettino, L. 1986. Algunos patrones distribucionales y ecológicos de los reptiles cubanos. *Poeyana* 305: 1-15.
- Rodríguez-Schettino, L. 1993. Areas faunísticas de Cuba según la distribución ecogeográfica actual y el endemismo de los reptiles. *Poeyana* 436: 1-17.
- Rodríguez-Schettino, L. 1995. Checklist and bibliography (1837-1991) of Cuban iguanid lizards. *Smithsonian Herpetological Information Service* 103: 1-29.
- Rosen, D. E. 1976. A vicariance model of Caribbean biogeography. *Syst. Zool.* 24: 431-464.
- Rosen, D. E. 1978. Vicariant patterns and historical explanation in biogeography. *Syst. Zool.* 27: 159-188.
- Rosen, D. E. 1985. Geological hierarchies and biogeographic congruence in the Caribbean. *Ann. Missouri Bot. Gard.* 72: 636-659.
- Roughgarden, J. 1990. Origin of the eastern Caribbean: data from reptiles and amphibians, pp. 10-26 in *Transactions of the 12th Caribbean Geological Conference, St. Croix, USVI*, edited by D. K. Larue, and G. Draper. Miami Geol. Soc., Miami.
- Roughgarden, J. 1995. *Anolis* Lizards of the Caribbean: Ecology, Evolution, and Plate Tectonics. Oxford Univ. Press, New York.
- Roughgarden, J., D. Heckel, and E. R. Fuentes. 1983. Coevolutionary theory and the biogeography and community structure of *Anolis*, pp. 371-410 in *Lizard Ecology*, edited by R. B. Huey, E. R. Pianka, and T. W. Schoener. Harvard Univ. Press, Cambridge, MA.
- Roughgarden, J., and S. Pacala. 1989. Taxon cycle among *Anolis* lizard populations: review of evidence, pp. 403-432 in *Speciation and Its Consequences*, edited by D. Otte, and J. Endler. Sinauer, Sunderland, MA.

- Roughgarden, J., W. Porter, and D. Heckel. 1981. Resource partitioning of space and its relationship to body temperature in *Anolis* lizard populations. *Oecologia* 50: 256-264.
- Rouse, I. 1989. Peopling and re-peopling of the West Indies, pp. 119-136 in *Biogeography of the West Indies: Past, Present, and Future*, edited by C. A. Woods. Sandhill Crane Press, Inc., Gainesville. FL.
- Rouse, I. 1992. *The Tainos: Rise and Decline of the People Who Greeted Columbus*. Yale Univ. Press, New Haven, CT.
- Ruibal, R. 1946. A new *Sphaerodactylus* from the Dominican Republic. *Novitates* 1308: 1-4.
- Ruibal, R. 1959. *Bufo gundlachi*, a new species of Cuban toad. *Breviora, Mus. Comp. Zool.* 105: 1-14.
- Ruibal, R. 1961. Thermal relations of five species of tropical lizards. *Evolution* 15: 98-111.
- Ruibal, R., and R. Philibosian. 1974. The population ecology of the lizard *Anolis acutus*. *Ecology* 55: 525-537.
- Ruibal, R. and E. E. Williams. 1961. Two sympatric anoles of the *carolinensis* group. *Bull. Museum Comp. Zool.* 125(7): 183-208.
- Russel, A. P. 1972. The foot of gekkonid lizards: a study in comparative and functional anatomy. Ph.D. Dissertation, Univ. London, England.
- Russell, A. P. 1979. Parallelism and integrated design in the foot structure of Gekkonine and Diplodactyline geckos. *Copeia* 1979: 1-21.
- Russell, A. P., and A. M. Bauer. 1991a. *Anolis garmani*. *Cat. Am. Amphib. and Rept.* 513: 1-3.
- Russell, A. P., and A. M. Bauer. 1991b. *Anolis grahami*. *Cat. Am. Amphib. and Rept.* 514: 1-4.
- Russell, A. P., and A. M. Bauer. 1991c. *Anolis opalinus*. *Cat. Am. Amphib. and Rept.* 515: 1-2.

- Sajdak, R. A., and R. W. Henderson. 1991. Status of West Indian racers in the Lesser Antilles. *Oryx* 25: 33-38.
- Salomao, M. da G., W. Wüster, R. S. Thorpe, J.-M. Touzet, and BBBSP. 1997. DNA evolution of South American pitvipers of the genus *Bothrops* (Reptilia: Serpentes: Viperidae), pp. 89-98 in *Venomous Snakes: Ecology, Evolution and Snakebite*, edited by R. S. Thorpe, W. Wüster, and A. Malhorta. Clarendon Press, Oxford, UK.
- Sampedro Marin, A., and O. T. Fundora. 1982. Hábitos alimentarios y actividad de *Bufo peltocephalus* Tschudi (Amphibia: Bufonidae) en el Jardín Botánico de Cianguegos. *Poeyana* 233: 1-14.
- Sampson, L. V. 1904. A contribution to the embryology of *Hylodes martiniensis*. *Am. J. Anat.* 3: 473-504.
- Sauer, J. D. 1969. Oceanic islands and biogeographic theory: a review. *Geographr. Rev.* 59: 582-593.
- Savage, J. M. 1955. The lizard family Xantusiidae: an evolutionary study. Ph.D. dissertation, Stanford Univ., Stanford, California.
- Savage, J. M. 1963. Studies on the lizard family Xantusiidae IV. The genera. *Contrib. Sci. Los Angeles County Museum* 71: 1-38.
- Savage, J. M. 1964. Studies on the lizard family Xantusiidae. V. The Cuban night lizard, *Cricosaura typica* Gundlach and Peters. *Copeia* 1964: 536-542.
- Savage, J. M. 1966. The origins and history of the Central American herpetofauna. *Copeia* 1966: 719-766.
- Savage, J. M. 1982. The enigma of the Central American herpetofauna: dispersals or vicariance? *Ann. Missouri Bot. Gard.* 69: 464-547.
- Savage, J. M. 1987. Systematics and distribution of the Mexican and Central American rainfrogs of the *Eleutherodactylus gollmeri* group (Amphibia: Leptodactylidae). *Fieldiana Zool.* 33: 1-57.
- Savage, J. M., and K. R. Lips. 1993. A review of the status and biogeography of the lizard genera *Celestus* and *Diploglossus* (Squamata: Anguidae), with description of two new species from Costa Rica. *Rev. Biol. Tropics* 41: 817-842.

- Savage, J. M., and J. Villa R. 1986. Introduction to the herpetofauna of Costa Rica. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.
- Schatzinger, R. A. 1980. New species of *Palaeoxantusia* (Reptilia: Sauria) from the Uintan (Eocene) of San Diego Co., California. *J. Paleontol.* 54: 460-471.
- Schell, P. T., J. S. Parmerlee, Jr., and R. Powell. 1993a. *Amieva chrysolema*. *Cat. Am. Amphib. Rept.* 563: 1-6.
- Schell, P. T., R. Powell, J. J. S. Parmerlee, A. Lathrop, and D. D. Smith. 1993b. Notes on the natural history of *Ameiva chrysolema* (Sauria: Teiidae), from Barahona, Dominican Republic. *Copeia* 1993: 859-862.
- Schlegel, H. 1837. *Essai sur la Physionomie des Serpens*. Arnz, Leiden.
- Schlegel, H. 1837-1844. *Abbildungen neuer oder unvollständig bekannter Amphibien, nach der Natur oder dem Leben entworfen, herausgegeben und mit einem erläuternden Texte begleitet*. Arnz & Comp., Düsseldorf.
- Schmidt, K. P. 1919. Descriptions of new amphibians and reptiles from Santo Domingo and Navassa. *Bull. Am. Museum Nat. Hist.* 41: 519-525.
- Schmidt, K. P. 1920. A new *Ameiva* from Nevis Island, British West Indies. *Proc. Linn. Soc. New York* 33: 1-8.
- Schmidt, K. P. 1921a. Notes on the herpetology of Santo Domingo. *Bull. Am. Museum Nat. Hist.* 44: 7-20.
- Schmidt, K. P. 1921b. The herpetology of Navassa Island. *Bull. Am. Museum Nat. Hist.* 44: 555-559.
- Schmidt, K. P. 1926. The amphibians and reptiles of Mona Island, West Indies. *Field Museum Nat. Hist. Ser.* 12: 149-162.
- Schmidt, K. P. 1928. Amphibians and land reptiles of Porto Rico, with a list of those reported from the Virgin Islands. *New York Acad. Sci.* 10: 1-160.
- Schmidt, K. P. 1955. Herpetology, pp. 591-627 in *A Century of Progress in the Natural Sciences 1853-1953*. California Academy of Sciences, San Francisco.

- Schoener, A., and T. W. Schoener. 1984. Experiments on dispersal: short-term flotation of insular anoles, with a review of similar abilities in other terrestrial animals. *Oecologia* 63: 289-294.
- Schoener, T. W. 1968. The *Anolis* of Bimini: Resource partitioning in a complex fauna. *Ecology* 49: 704-726.
- Schoener, T. W. 1975. Presence and absence of habitat shift in some widespread lizard species. *Ecol. Monogr.* 45: 233-258.
- Schoener, T. W. 1983. Rate of species turnover decreases from lower to higher organisms: a review of the data. *Oikos* 41: 372-377.
- Schoener, T. W., and G. C. Gorman. 1968. Some niche differences in three Lesser Antillean lizards of the genus *Anolis*. *Ecology* 49: 819-830.
- Schoener, T. W., and A. Schoener. 1971a. Structural habitats of West Indian *Anolis* lizards I. Lowland Jamaica. *Breviora* 368: 1-53.
- Schoener, T. W., and A. Schoener. 1971b. Structural habitats of West Indian *Anolis* lizards II. Puerto Rico uplands. *Breviora* 375: 1-39.
- Schoener, T. W., and A. Schoener. 1980. Densities, sex ratios, and population structure in four species of Bahamian *Anolis* lizards. *J. Anim. Ecol.* 49: 19-53.
- Schoener, T. W., and A. Schoener. 1983a. Distribution of vertebrates on some very small islands. II. Patterns in species number. *J. Anim. Ecol.* 52: 237-262.
- Schoener, T. W., and A. Schoener. 1983b. On the voluntary departure of lizards from very small islands, pp. 491-498 in *Advances in Herpetology and Evolutionary Biology: Essays in Honor of Ernest E. Williams*, edited by A. G. J. Rhodin and K. Miyata. Museum Comp. Zool., Harvard Univ. Press., Cambridge, MA.
- Schoener, T. W., and A. Schoener. 1983c. Time of extinction of a colonizing propagule of lizards increases with island area. *Nature* 302: 332-334.
- Schoener, T. W., J. B. Slade, and C. H. Stinson. 1982. Diet and sexual dimorphism in the very catholic lizard genus, *Leiocephalus* of the Bahamas. *Oecologia* 61: 160-169.

- Schoener, T. W., and D. A. Spiller. 1987. Effect of lizards on spider populations: manipulative reconstruction of a natural experiment. *Science* 236: 949-952.
- Schoener, T. W., and C. A. Toft. 1983. Spider populations: extraordinary densities on islands without top predators. *Science* 219: 1353-1355.
- Schreckenberg, S. M. G. 1956. The embryonic development of the thyroid gland in the frog, *Hyla brunnea*. *Growth* 20: 295-313.
- Schreiber, M. C., R. Powell, J. S. Parmerlee, Jr., A. Lathrop, and D. D. Smith. 1993. Natural history of a small population of *Leiocephalus schreibersi* (Sauria: Tropiduridae) from altered habitat in the Dominican Republic. *Florida Sci.* 56: 82-90.
- Schubert, A. 1993. Conservation of biological diversity in the Dominican Republic. *Oryx* 27: 115-121.
- Schubert, A., and G. Santana. 1996. Conservation of the American crocodile (*Crocodylus acutus*) in the Dominican Republic, pp. 425-433 in *Contributions to West Indian Herpetology: A Tribute to Albert Schwartz*, edited by R. Powell, and R. W. Henderson. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.
- Schuchert, C. 1935. *Historical Geology of the Antillean-Caribbean Region*. Wiley, New York.
- Schwanner, T. D., and H. C. Dessauer. 1981. Immunodiffusion evidence for the relationships of Papuan boids. *J. Herpetol.* 15: 250-253.
- Schwartz, A. 1957. A new species of *Eleutherodactylus* (Amphibia: Leptodactylidae) from Cuba. *Proc. Biol. Soc. Washington* 70(23): 209-212.
- Schwartz, A. 1959. A new species of toad, *Bufo cataulaciceps*, from the Isla de Pinos and Western Cuba. *Proc. Biol. Soc. Washington* 72(4): 109-120.
- Schwartz, A. 1960. Nine new Cuban frogs of the genus *Eleutherodactylus*. *Sci. Publ. Reading Public Museum and Art Gallery* 11: 1-60.
- Schwartz, A. 1961. A review of geckoes of the *Sphaerodactylus scaber* group of Cuba. *Herpetologica* 17: 19-26.

- Schwartz, A. 1964 (1965). Two new species of *Eleutherodactylus* from the eastern Cordillera Central of the República Dominicana. *Carib. J. Sci.* 4: 473-484.
- Schwartz, A. 1964a. Three new species of frogs (Leptodactylidae, *Eleutherodactylus*) from Hispaniola. *Breviora* 208: 1-15.
- Schwartz, A. 1964b. *Diploglossus costatus* Cope (Sauria, Anguidae) and its relatives in Hispaniola. *Sci. Publ. Reading Public Museum Art Gallery* 13: 1-57.
- Schwartz, A. 1964c. A review of *Sphaerodactylus vincenti* on the southern windward islands. *Carib. J. Sci.* 4: 391-409.
- Schwartz, A. 1965a. Variation and natural history of *Eleutherodactylus ruthae* on Hispaniola. *Bull. Museum Comp. Zool.* 132: 481-508.
- Schwartz, A. 1965b. Two new subspecies of the anguid lizard *Wetmorena* from Hispaniola. *Proc. Biol. Soc. Washington* 78: 39-48.
- Schwartz, A. 1965c. Geographic variation in two species of Hispaniolan *Eleutherodactylus*, with notes on Cuban members of the *ricordi* group. *Stud. Fauna Curaçao Carib. Isl.* 22: 99-114.
- Schwartz, A. 1965d. A new species of *Eleutherodactylus* (Amphibia: Leptodactylidae) from the Sierra de Baoruco, República Dominicana. *Proc. Biol. Soc. Washington* 78: 165-168.
- Schwartz, A. 1965e. The *Ameiva* (Reptilia: Teiidae) of Hispaniola. I. *Ameiva lineolata* Duméril and Bibron. *Carib. J. Sci.* 5: 45-57.
- Schwartz, A. 1965f. A new subspecies of the gecko *Sphaerodactylus microlepis*. *Herpetologica* 21: 261-269.
- Schwartz, A. 1966a. The relationships of four small Hispaniolan *Eleutherodactylus* (Leptodactylidae). *Bull. Museum Comp. Zool.* 133: 371-399.
- Schwartz, A. 1966b. The *Leiocephalus* (Lacertilia, Iguanidae) of Hispaniola. I. *Leiocephalus melanochlorus* Cope. *J. Ohio Herpetol. Soc.* 5: 39-48.
- Schwartz, A. 1966c. Snakes of the genus *Alsophis* in Puerto Rico and the Virgin Islands. *Studies Fauna Curaçao Carib. Isl.* 23: 175-227.

- Schwartz, A. 1967a. The *Leiocephalus* (Lacertilia, Iguanidae) of Hispaniola. II. The *Leiocephalus personatus* complex. Tulane Stud. Zool. 14: 1-53.
- Schwartz, A. 1967b. The *Ameiva* (Lacertilia, Teiidae) of Hispaniola. III. *Ameiva taeniura* Cope. Bull. Museum Comp. Zool. 135: 345-375.
- Schwartz, A. 1967c. Frogs of the genus *Eleutherodactylus* in the Lesser Antilles. Stud. Fauna Curaçao Carib. Isl. 24: 1-62.
- Schwartz, A. 1968a. Two new subspecies of *Ameiva* (Lacertilia, Teiidae) from Hispaniola. Herpetologica 24: 21-28.
- Schwartz, A. 1968b. The *Leiocephalus* (Lacertilia, Iguanidae) of Hispaniola. III. *Leiocephalus schreibersi*, *L. semilineatus*, and *L. pratensis*. J. Herpetol. 1: 39-63.
- Schwartz, A. 1968c. Geographic variation in the Hispaniolan frog *Eleutherodactylus wetmorei* Cochran. Breviora 290: 1-13.
- Schwartz, A. 1968d. Geographic variation in *Anolis distichus* Cope (Lacertilia, Iguanidae) in the Bahama Islands and Hispaniola. Bull. Museum Comp. Zool. 137: 255-309.
- Schwartz, A. 1969a. A review of the Hispaniolan lizard *Anolis coelestinus* Cope. Carib. J. Sci. 9: 33-38.
- Schwartz, A. 1969b. Two new subspecies of *Leiocephalus* from Hispaniola. J. Herpetol. 3: 79-85.
- Schwartz, A. 1969c. The Antillean *Eleutherodactylus* of the *auriculatus* group. Stud. Fauna Curaçao Carib. Isl. 30: 99-115.
- Schwartz, A. 1970a. A new species of large *Diploglossus* (Sauria: Anguidae) from Hispaniola. Proc. Biol. Soc. Washington 82: 777-788.
- Schwartz, A. 1970b. A new species of gecko (Gekkonidae, *Sphaerodactylus*) from Hispaniola. J. Herpetol. 4: 63-67.
- Schwartz, A. 1970c. A systematic review of *Uromacer catesbyi* Schlegel (Serpentes, Colubridae). Tulane Stud. Zool. Bot. 16: 131-149.

- Schwartz, A. 1970d. *Darlingtonia haetiana* (Serpentes, Colubridae): a new subspecies. *Herpetologica* 26: 324-331.
- Schwartz, A. 1971a. A new species of bromeliad-inhabiting galliwasp (Sauria: Anguidae) from Jamaica. *Breviora* 371: 1-10.
- Schwartz, A. 1971b. A new species of *Eleutherodactylus* (Amphibia, Leptodactylidae) from Hispaniola. *Annu. Carnegie Museum* 43: 25-31.
- Schwartz, A. 1971c. A systematic account of the Hispaniolan snake genus *Hypsirhynchus*. *Stud. Fauna Curaçao Carib. Isl.* 35: 63-94.
- Schwartz, A. 1971d. A new subspecies of *Leiocephalus personatus* (Sauria, Iguanidae). *Herpetologica* 27: 176-182.
- Schwartz, A. 1971e. Two new subspecies of *Diploglossus costatus* from Hispaniola (Sauria, Anguidae). *J. Herpetol.* 5: 161-165.
- Schwartz, A. 1972. The native toads (Anura, Bufonidae) of Hispaniola. *J. Herpetol.* 6: 217-231.
- Schwartz, A. 1973a. A new subspecies of *Ameiva chrysoleama* (Sauria, Teiidae) from Haiti. *Herpetologica* 29: 101-105.
- Schwartz, A. 1973b. A third species in the Hispaniolan *shrevei* group of *Sphaerodactylus* (Sauria, Gekkonidae). *Proc. Biol. Soc. Washington* 86: 35-40.
- Schwartz, A. 1973c. Six new species of *Eleutherodactylus* (Anura, Leptodactylidae) from Hispaniola. *J. Herpetol.* 7: 249-273.
- Schwartz, A. 1973d. A new species of montane *Anolis* (Sauria, Iguanidae) from Hispaniola. *Ann. Carnegie Museum* 44: 183-195.
- Schwartz, A. 1974a. A new species of primitive *Anolis* (Sauria, Iguanidae) from the Sierra de Bahoruco, Hispaniola. *Breviora* 423: 1-19.
- Schwartz, A. 1974b. An analysis of variation in the Hispaniolan giant anole, *Anolis ricordi* Duméril and Bibron. *Bull. Museum Comp. Zool.* 146: 89-146.
- Schwartz, A. 1975a. New subspecies of *Sphaerodactylus copei* Steindachner (Sauria, Gekkonidae) from Hispaniola. *Herpetologica* 31: 1-18.

- Schwartz, A. 1975b. Variation in the Antillean boid snake *Tropidophis haetianus* Cope. *J. Herpetol.* 9: 303-311.
- Schwartz, A. 1975c. A new subspecies of *Anolis baleatus* Cope (Sauria: Iguanidae) from the República Dominicana. *Florida Sci.* 38: 30-35.
- Schwartz, A. 1976a. Two new species of Hispaniolan *Eleutherodactylus* (Leptodactylidae). *Herpetologica* 32: 163-171.
- Schwartz, A. 1976b. Variation and relationships of some Hispaniolan frogs (Leptodactylidae, *Eleutherodactylus*) of the *ricordi* group. *Bull. Florida St. Museum* 21: 1-46.
- Schwartz, A. 1976c. A new species of *Sphaerodactylus* (Sauria, Gekkonidae) from the República Dominicana. *Florida Sci.* 39: 65-70.
- Schwartz, A. 1976d. Variations in the Hispaniolan colubrid snake *Uromacer frenatus* Günther (Reptilia, Serpentes, Colubridae). *J. Herpetol.* 10: 319-327.
- Schwartz, A. 1977a. A new subspecies of *Eleutherodactylus wetmorei* Cochran (Anura: Leptodactylidae) from northern Haiti. *Herpetologica* 33: 66-72.
- Schwartz, A. 1977b. The geckoes (Sauria, Gekkonidae) of the genus *Sphaerodactylus* of the Dominican Peninsula de Barahona, Hispaniola. *Proc. Biol. Soc. Washington* 90: 243-254.
- Schwartz, A. 1977c. A new subspecies of *Anolis baleatus* (Sauria: Iguanidae) from Isla Saona, República Dominicana. *Florida Sci.* 40: 401-405.
- Schwartz, A. 1978a. Some aspects of the herpetogeography of the West Indies, pp. 31-51 in *Zoogeography in the Caribbean, The 1975 Leidy Medal Symposium*, edited by F. B. Gill. *Acad. Nat. Sci. Philadelphia Spec. Publ.*, Philadelphia.
- Schwartz, A. 1978b. A new species of aquatic *Anolis* (Sauria, Iguanidae) from Hispaniola. *Ann. Carnegie Museum* 47: 261-279.
- Schwartz, A. 1978c. The Hispaniolan *Anolis* (Reptilia, Lacertilia, Iguanidae) of the *hendersoni* complex. *J. Herpetol.* 12: 355-370.
- Schwartz, A. 1979a. A new species of cybotoid anole (Sauria, Iguanidae) from Hispaniola. *Breviora* 451: 1-27.

- Schwartz, A. 1979b. A new species of *Eleutherodactylus* (Amphibia, Anura, Leptodactylidae) from northwestern Haiti, Hispaniola. *J. Herpetol.* 13: 199-202.
- Schwartz, A. 1979c. A new species of *Leiocephalus* (Reptilia: Iguanidae) from Hispaniola. *Proc. Biol. Soc. Washington* 92: 272-279.
- Schwartz, A. 1979d. The status of *Uromacer frenatus* and *U. dorsalis* (Reptilia: Serpentes: Colubridae). *Herpetologica* 35: 207-216.
- Schwartz, A. 1979e. The herpetofauna of Ile à Cabrit, Haiti, with the descriptions of two new subspecies. *Herpetologica* 35: 248-255.
- Schwartz, A. 1979f. The status of Greater Antillean *Phyllodactylus* (Reptilia, Gekkonidae). *J. Herpetol.* 13: 419-426.
- Schwartz, A. 1980a. Two new species of *Eleutherodactylus* (Amphibia, Anura, Leptodactylidae) from Hispaniola. *Ann. Carnegie Museum* 49: 103-112.
- Schwartz, A. 1980b. The herpetogeography of Hispaniola, West Indies. *Stud. Fauna Curaçao Carib. Isl.* 61: 86-127.
- Schwartz, A. 1980c. Variation in Hispaniolan *Anolis whitemani* Williams. *J. Herpetol.* 14: 399-406.
- Schwartz, A. 1981. Variation in Hispaniola *Anolis olssoni* Schmidt (Reptilia, Sauria, Iguanidae). *Milwaukee Public Museum Contrib. Biol. Geol.* 47: 1-21.
- Schwartz, A. 1983. The *difficilis* complex of *Sphaerodactylus* (Sauria, Gekkonidae) of Hispaniola. Pt. 1. *Sphaerodactylus difficilis*, *S. clenchi*, and *S. lazelli*. *Bull. Carnegie Museum Nat. Hist.* 22: 5-30.
- Schwartz, A., and M. Carey. 1977. Systematics and evolution in the West Indian iguanid genus *Cyclura*. *Stud. Fauna Curaçao Carib. Is.* 53: 15-97.
- Schwartz, A., and R. I. Crombie. 1975. A new species of the genus *Aristelliger* (Sauria: Gekkonidae) from the Caicos Islands. *Proc. Biol. Soc. Washington* 88: 305-314.
- Schwartz, A., and D. C. Fowler. 1973. The Anura of Jamaica: a progress report. *Stud. Fauna Curaçao Carib. Is.* 43: 50-142.

- Schwartz, A., and R. Franz. 1976. A new species of *Sphaerodactylus* (Sauria: Gekkonidae) from Hispaniola. Proc. Biol. Soc. Washington 88: 367-372.
- Schwartz, A. and O. H. Garrido. 1972. The lizards of the *Anolis equestris* complex in Cuba. Stud. Fauna Curaçao and Carib. Is. 39(134): 1-86.
- Schwartz, A., and O. H. Garrido. 1975. A reconsideration of some Cuban *Tropidophis* (Serpentes, Boidae). Poc. Biol. Soc. Washington 88: 77-90.
- Schwartz, A., and O. H. Garrido. 1981a. Las salamanquitas cubanos del género *Sphaerodactylus* (Sauria: Gekkonidae). 1. El grupo *copei*. Poeyana 230: 1-27.
- Schwartz, A., and O. H. Garrido. 1981b. A review of the Cuban members of the genus *Arrhyton* (Reptilia, Serpentes, Colubridae). Ann. Carnegie Museum Nat Hist. 50: 207-230.
- Schwartz, A., and O. H. Garrido. 1985. The Cuban lizards of the genus *Sphaerodactylus* (Sauria, Gekkonidae). Milwaukee Public Museum Contrib. Biol. Geol. 62: 1-67.
- Schwartz, A., and J. E. D. Graham. 1980. The *shrevei* group of Hispaniolan *Sphaerodactylus* (Reptilia, Gekkonidae). Tulane Stud. Zool. Bot. 22: 1-15.
- Schwartz, A., J. E. D. Graham, and J. J. Duval. 1979. A new species of *Diploglossus* (Sauria: Anguillidae) from Hispaniola. Proc. Biol. Soc. Washington 92: 1-9.
- Schwartz, A., and R. W. Henderson. 1982. *Anolis cybotes* (Reptilia, Iguanidae): the eastern Hispaniolan populations. Milwaukee Public Museum Contrib. Biol. Geol. 49: 1-8.
- Schwartz, A., and R. W. Henderson. 1985. A Guide to the Identification of the Reptiles and Amphibians of the West Indies Exclusive of Hispaniola. Milwaukee Public Museum, Milwaukee, WI.
- Schwartz, A., and R. W. Henderson. 1988. West Indian amphibians and reptiles: a check-list. Milwaukee Public Museum Contrib. Biol. Geol. 74: 1-264.
- Schwartz, A., and R. W. Henderson. 1991. Amphibians and Reptiles of the West Indies: Descriptions, Distributions, and Natural History. Univ. Florida Press, Gainesville, FL.

- Schwartz, A., and S. J. Incháustegui. 1976. A new species of *Diploglossus* (Reptilia, Lacertilia, Anguinae) from Hispaniola. *J. Herpetol.* 10: 241-246.
- Schwartz, A., and J. F. Jacobs. 1989. A new subspecies of *Celestus costatus* (Sauria: Anguinae) from the República Dominicana. *J. Herpetol.* 23: 193-195.
- Schwartz, A., and R. F. Klinikowski. 1966. The *Ameiva* (Lacertilia, Teiidae) of Hispaniola. II. Geographic variation in *Ameiva chrysolema* Cope. *Bull. Museum Comp. Zool.* 133: 427-487.
- Schwartz, A., and R. J. Marsh. 1960. A review of the *pardalis-maculatus* complex of the booid genus *Tropidophis* of the West Indies. *Bull. Museum Comp. Zool.* 123: 49-84.
- Schwartz, A., and C. J. McCoy. 1970. A systematic review of *Ameiva auberi* Cocteau (Reptilia, Teiidae) in Cuba and the Bahamas. *Ann. Carnegie Museum* 41: 43-168.
- Schwartz, A., and D. A. Rossman. 1976. A review of the Hispaniolan colubrid snake genus *Ialtris*. *Stud. Fauna Curaçao Carib. Isl.* 50: 76-102.
- Schwartz, A., and R. Thomas. 1964 (1965). Subspeciation in *Sphaerodactylus copei*. *Q. J. Florida Acad. Sci.* 27: 316-332.
- Schwartz, A., and R. Thomas. 1965. The genus *Darlingtonia* (Serpentes) in Hispaniola, including a new subspecies from the Dominican Republic. *Breviora* 229: 1-10.
- Schwartz, A., and R. Thomas. 1975. A check-list of West Indian amphibians and reptiles. *Carnegie Museum Nat. Hist. Spec. Publ.* 1: 1-216.
- Schwartz, A., and R. Thomas. 1977. Two new species of *Sphaerodactylus* (Reptilia, Lacertilia, Gekkonidae) from Hispaniola. *J. Herpetol.* 11: 61-66.
- Schwartz, A., and R. Thomas. 1983. The *difficilis* complex of *Sphaerodactylus* (Sauria, Gekkonidae) of Hispaniola. *Bull. Carnegie Museum Nat. Hist.* 22: 1-60.
- Schwartz, A., R. Thomas, and L. D. Ober. 1978. First supplement to a check-list of West Indian amphibians and reptiles. *Carnegie Museum Nat. Hist. Spec. Publ.* 5: 1-35.
- Schwenk, K. 1988. Comparative morphology of the lepidosaur tongue and its relevance to squamate phylogeny, pp. 569-598 in *Phylogenetic Relationships of*

the Lizard Families: Essays Commemorating Charles L. Camp, edited by R. Estes and G. Pregill. Stanford Univ. Press, Stanford, CA.

Scott, N. J. 1976. The abundance and diversity of the herpetofauna of tropical forest litter. *Biotropica* 8: 41-58.

Secretaría de Estado de Agricultura/Departamento de Vida Silvestre (SEA/DVS). 1990. La Diversidad Biológica en la República Dominicana: Reporte Preparado por el Departamento de Vida Silvestre para el Servicio Alemán de Cooperación Social-Técnica y Fondo Mundial para la Naturaleza (WWF-US). Secretaría de Estado de Agricultura, SURENA/DVS, Santo Domingo, República Dominicana.

Seidel, M. E. 1988a. Revision of the West Indian emydid turtles. *Novitates* 2918: 1-41.

Seidel, M. E. 1988b. *Trachemys decussata*. *Cat. Am. Amphib. Rept.*, 440: 1-3.

Seidel, M. E. 1990. Growth and population characteristics of the slider turtle, *Trachemys decussata* on Grand Cayman Island. *J. Herpetol.*, 24(2): 191-196.

Seidel, M. E. 1996. Current status of biogeography of the West Indian turtles in the genus *Trachemys* (Emydidae), pp. 169-174 in *Contributions to West Indian Herpetology: A Tribute to Albert Schwartz*, edited by R. Powell, and R. W. Henderson. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.

Seidel, M. E., and M. D. Adkins. 1987. Biochemical comparisons among West Indian *Trachemys* (Emydidae: Testudines). *Copeia* 1987: 485-489.

Seidel, M. E., and S. I. Miranda. 1984. Status of the trachemyd turtles (Testudines: Emydidae) on Hispaniola. *J. Herpetol.* 18: 468-479.

Seifriz, W. 1943. The plant life of Cuba. *Ecol. Monographs* 13: 375-426.

Sharpton, V. L., K. Burke, A. Camargo-Zanoguera, S. A. Hall, S. Lee, L. Marin, G. Suarez-Reynoso, J. M. Quezada-Muneton, P. D. Spudis, and J. Urrutia-Fucugauchi. 1993. Chicxulub multiring impact basin: size and other characteristics derived from gravity analysis. *Science* 261: 1564-1567.

Shaw, G. 1802. *General Zoology or Systematic Natural History*, 3: Amphibia. Part I & Part II. G. Kearsley, London.

- Sheplan, B. R., and A. Schwartz. 1974. Hispaniolan boas of the genus *Epicrates* (Serpentes, Boidae) and their Antillean relationships. *Ann. Carnegie Museum* 45: 57-143.
- Shochat, D., and H. C. Dessauer. 1981. Comparative immunological study of albumins of *Anolis* lizards of the Caribbean islands. *Comp. Biochem. Physiol.* 68A: 67-73.
- Shreve, B. J. 1936. A new *Anolis* and new Amphibia from Haiti. *Proc. New England Zool. Club* 15: 93-99.
- Shreve, B. J. 1968. The *notatus* group of *Sphaerodactylus* (Sauria: Gekkonidae) in Hispaniola. *Breviora* 280: 1-28.
- Shreve, B. J., and E. E. Williams. 1963. The herpetology of the Port-au-Prince region and Gonave Island, Haiti. Part II. The frogs. *Bull. Museum Comp. Zool.* 129: 302-342.
- Silva Lee, A. 1996. Cuba Natural. Natural Cuba. Pangaea, St. Paul.
- Simberloff, D., and W. Boeklin. 1991. Patterns of extinction in the introduced Hawaiian avifauna: a reexamination of the role of competition. *Am. Nat.* 138: 300-327.
- Simpson, G. G. 1956. Zoogeography of West Indian land mammals. *Am. Museum Novit.* 1759:1-28.
- Singh, R. S., R. C. Lewontin, and A. A. Felton. 1976. Genic heterogeneity within electrophoretic "alleles" of xanthine dehydrogenase in *Drosophila pseudoobscura*. *Genetics* 84: 609-629.
- Sites, J. W., S. K. Davis, T. Guerra, J. B. Iverson, and H. L. Snell. 1996. Character congruence and phylogenetic signal in molecular and morphological data sets: A case study in the living iguanas (Squamata, Iguanidae). *Mol. Biol. Evol.* 13: 1087-1105.
- Sloane, H. 1725. A voyage to the islands Madera, Barbadoes, Nieves, St. Christophers, and Jamaica; with the natural history....of the last of these islands. 2: 335-336, London.

- Slowinski, J. B. 1995. A phylogenetic analysis of the New World coral snakes (Elapidae: *Leptomicrurus*, *Micruroides*, and *Micrurus*) based on allozymic and morphological characters. *J. Herpetol.* 29: 325-338.
- Smith, D. D., and R. Powell. 1991. Geographic distribution: *Uromacer frenatus*. *Herpetol. Rev.* 22: 135-136.
- Smith, E. E. 1954. The forests of Cuba. Maria Moors Cabot Foundation Publ. 2: 1-98.
- Smith, H. M., and C. Grant. 1958. The proper name of some Cuban snakes: an analysis of dates of publication of Ramon de la Sagra's *Historia Natural de Cuba*, and of Fitzinger's *Systema Reptilium*. *Herpetologica* 4: 215-222.
- Smith, H. M., and E. H. Taylor. 1950. An annotated checklist and key to the reptiles of Mexico exclusive of the snakes. *Bull. U.S. Natl. Museum* 199: i-v + 1-253.
- Smith, H. M., and V. Wallach. 1993. The date of publication of *Ialtris* Cope (Reptilia: Serpentes). *Bull. Maryland Herpetol. Soc.* 29: 80-84.
- Smith, J. W., R. Powell, J. J. S. Parmerlee, D. D. Smith, and A. Lathrop. 1992. Natural history notes on a population of grass anoles, *Anolis olssoni* (Sauria: Polychrotidae), from the Dominican Republic. *Bull. Maryland Herpetol. Soc.* 30: 67-75.
- Smith, M. H., D. O. Straney, and G. E. Drewery. 1981. Biochemical similarities among Puerto Rican *Eleutherodactylus*. *Copeia* 1981: 463-466.
- Sonnini, C. S., and P.-A. Latreille. 1801. *Histoire Naturelle des Reptiles*. Lerrault, Paris.
- Sparman, A. 1784. *Lacerta sputator* och *Lacerta bimaculata*, tva nya Odlor fran America. *Kongliga Svenska Vetenskaps-Akad Handl.* 5: 164-167.
- Speed, R. C. 1985. Cenozoic collision of the Lesser Antilles arc and continental South America and the origin of the El Pilar Fault. *Tectonics* 4: 41-69.
- Speed, R. C., L. C. Gerhard, and E. H. McKee. 1979. Ages of deposition, deformation, and intrusion of Cretaceous rocks, eastern St. Croix, Virgin Islands. *Bull. Geol. Soc. Am.* 90: 629-632.

- Spix, J. B. 1825. *Animalia Nova sive Species Novae Lacertarum, quas in Itinere per Brasiliam Annis. F. S. Hübschmanni, Monachii.*
- Sproston, A. L., R. E. Glor, L. M. Hartley, E. J. Censky, R. Powell, and J. S. Parmerlee. 1998. Niche differences among three sympatric species of *Ameiva* (Reptilia: Teiidae) on Hispaniola. *J. Herpetol.*, in press.
- Stamps, J. A. 1976. Rainfall, activity, and social behavior in the lizard *Anolis aeneus*. *Anim. Behav.* 24: 603-608.
- Stamps, J. A. 1983. The relationship between ontogenetic habitat shifts, competition and predator avoidance in a juvenile lizard (*Anolis aeneus*). *Behav. Ecol. Sociobiol.* 12: 19-33.
- Stamps, J. A. 1985. Arboreal habitat use and parachuting by a subtropical forest frog. *J. Herpetol.* 19: 391-401.
- Stamps, J. A. 1987. The effect of familiarity with a neighborhood on territory acquisition. *Behav. Ecol. Sociobiol.* 21: 273-277.
- Stamps, J. A. 1988. The effect of body size on habitat and territory choice in juvenile lizards. *Herpetologica* 44: 369-376.
- Stamps, J. A. 1995. Climate driven population fluctuations in rain forest frogs. *J. Herpetol.* 29: 437-446.
- Stamps, J. A., and G. E. Martin. 1980. Coconut husk-piles: a unique habitat for Jamaican terrestrial frogs. *Biotropica* 12: 107-117.
- Stamps, J. A., and F. H. Pough. 1983. Population density of tropical forest frogs: relation to retreat sites. *Science* 221: 570-572.
- Stamps, J. A., and A. S. Rand. 1991. Vocalizations and the defense of retreat sites by male and female frogs, *Eleutherodactylus coqui*. *Copeia* 1991: 1013-1024.
- Stamps, J. A., and S. K. Tanaka. 1981. The relationship between food and social behavior in juvenile lizards (*Anolis aeneus*). *Copeia* 1981: 422-434.
- Stamps, J. A., and L. L. Woolbright. 1998. Amphibians, in *The Food Web of a Tropical Rain Forest*, edited by D. P. Reagan, and R. B. Waide. Univ. Chicago Press, Chicago.

- Steadman, D. W., G. K. Pregill, and S. L. Olsen. 1984a. Fossil vertebrates from Antigua, Lesser Antilles: Evidence for late Holocene human-caused extinctions in the West Indies. *Proc. Natl. Acad. Sci.* 81: 4448-4451.
- Steadman, D. W., D. R. Watters, E. J. Reitz, and G. K. Pregill. 1984b. Vertebrates from archaeological sites on Montserrat, West Indies. *Ann. Carnegie Museum* 53(1):1-29.
- Stearns, S. C. 1992. *The evolution of life histories*. Oxford Univ. Press, Oxford and New York.
- Steers, J. A. 1940a. The cays and the Palisadoes, Port Royal, Jamaica. *Geogr. Rev.* 30: 279-296.
- Steers, J. A. 1940b. The coral cays of Jamaica. *Geogr. J.* 95: 30-42.
- Steers, J. A., V. J. Chapman, J. Colman, and J. A. Lofthouse. 1940. Sand cays and mangroves in Jamaica. *Geogr. J.* 96: 305-328.
- Stehli, F. G., and S. D. Webb (Editors). 1985. *The Great American Biotic Interchange*. Plenum, New York.
- Steindachner, F. 1863. Über eine neue *Epicrates*-Art aus Columbien. *Denskschr. Kaiserl. Akad. Wiss. Wien, math.-nat. cl.* 22: 89-93.
- Steindachner, F. 1867. Reptilien, pp. 1-98 in *Reise der Österreichischen Fregatte Novara um die Erde in den Jahren 1857, 1858, 1859 unter den Befehlen des Commodore B. von Wüllerstorff-Urbair*. K. Gerold's Sohn, Wien.
- Steindachner, F. 1917. Cuban amphibians and reptiles collected for the United States National Museum from 1899 to 1902. *Proc. U.S. Natl. Museum* 53: 259-291.
- Stejneger, L. 1899. A new name for the Great Crested Lizard of Jamaica. *Am. Nat.* 33: 601-602.
- Stejneger, L. 1901. A new systematic name for the Yellow boa of Jamaica. *Am. Nat.* 23: 469-470.
- Stejneger, L. 1904. The herpetology of Porto Rico. *Rep. U.S. Nat. Museum* 1902: 551-724.

- Stejneger, L. 1916. Notes on amphisbaenid nomenclature. Proc. Biol. Soc. Washington 29: 86.
- Stewart, M. M. 1979. The role of introduced species in a Jamaican frog community. Act. Symp. Int. Ecol. Trop. 1: 111-146.
- Stewart, M. M., 1985. Arboreal habitat use and parachuting by a subtropical forest frog. J. Herpetol. 19(3): 391-401.
- Stewart, M. M., and G. E. Martin. 1980. Coconut-husk piles – a unique habitat for Jamaican terrestrial frogs. Biotropica 12: 107-116.
- Stewart, M. M. and F. H. Pough. 1983. Population density of tropical forest frogs: relation to retreat sites. Science 221: 570-572.
- Stewart, M. M., and A. S. Rand. 1991. Vocalization and the defense retreat sites by male and female frogs, *Eleutherodactylus coqui*. Copeia 1991: 1013-1024.
- Stewart, M. M., and L. L. Woolbright. 1996. Amphibians. in The Food Web of a Tropical Rain Forest, edited by D. P. Reagan, and R. B. Waide. Univ. Chicago Press, Chicago.
- Stinnesbeck, W., G. Keller, J. de la Cruz, C. de León, N. MacLeod, and J.E. Whittaker. 1997. The Cretaceous-Tertiary transition in Guatemala: limestone breccia deposits from the South Petén basin. Geol. Rundsch 86: 686-709.
- Stouder, S. A. 1959. Natural History of the West Indies, by Gonzalo F. de Oviedo. Univ. North Carolina Press, Chapel Hill.
- Strahm, M. H., and A. Schwartz. 1977. Osteoderms in the anguid lizard subfamily Diploglossinae and their taxonomic importance. Biotropica 9: 58-72.
- Stuart, L. C. 1941. Studies on the neotropical Colubrinae. VIII. A revision of the genus *Dryadophis* Stuart, 1939. Misc. Publ. Museum Zool. Univ. Michigan 49: 1-106.
- Stull, O. G. 1928. A revision of the genus *Tropidophis*. Occas. Pap. Museum Zool. Univ. Michigan 195: 1-49.

- Stull, O. G. 1935. A checklist of the family Boidae. Proc. Boston Soc. Nat. Hist. 40: 387-408.
- Swofford, D. L. 1993. PAUP: phylogenetic analysis using parsimony, version 3.1.1 Computer program distributed by Illinois Natural History Survey, Champaign, Illinois.
- Taigen, T. L., F. H. Pough, and M. M. Stewart. 1984. Water balance of terrestrial anuran (*Eleutherodactylus coqui*) eggs: importance of parental care. Ecology 65: 248-255.
- Taylor, C. E., and G. C. Gorman. 1975. Population genetics of a "colonizing" lizard: natural selection for allozyme morphs in *Anolis grahami*. Heredity 35: 241-247.
- Taylor, E. H. 1952. A new Panamanian tree frog. Breviora 1: 1-3.
- Thomas, R. 1964. The races of *Sphaerodactylus fantasticus* Duméril and Bibron in the Lesser Antilles. Carib. J. Sci. 4: 373-390.
- Thomas, R. 1965a. Two new subspecies of *Amphisbaena* (Amphisbaenia, Reptilia) from the Barahona Peninsula of Hispaniola. Breviora 215: 1-14.
- Thomas, R. 1965b. The genus *Leptotyphlops* in the West Indies with description of a new species from Hispaniola (Serpentes, Leptotyphlopidae). Breviora 222: 1-12.
- Thomas, R. 1965c. A new species of *Typhlops* from the Barahona Peninsula of Hispaniola. Copeia 1965: 436-439.
- Thomas, R. 1965d. The smaller teiid lizards (*Gymnophthalmus* and *Bachia*) of the southeastern Caribbean. Proc. Biol. Soc. Washington 78: 141-154.
- Thomas, R. 1966a. A reassessment of the herpetofauna of Navassa Island. J. Ohio Herpetol. Soc. 5: 73-89.
- Thomas, R. 1966b. A new Hispaniolan gecko. Breviora 253: 1-5.
- Thomas, R. 1966c. Leeward Island *Typhlops* (Reptilia, Serpentes). Proc. Biol. Soc. Washington 79: 255-266.

- Thomas, R. 1966d. Additional notes on the amphisbaenids of greater Puerto Rico. *Breviora* 249: 1-23.
- Thomas, R. 1968. Notes on Antillean geckos (*Sphaerodactylus*). *Herpetologica* 24: 46-60.
- Thomas, R. 1971. A new species of *Diploglossus* (Sauria: Anguinae) from Hispaniola. *Occas. Pap. Museum Zool. Louisiana State Univ.* 40: 1-9.
- Thomas, R. 1974a. A new species of *Typhlops* (Serpentes: Typhlopidae) from Hispaniola. *Proc. Biol. Soc. Washington* 87: 11-18.
- Thomas, R. 1974b. A new species of Lesser Antillean *Typhlops* (Serpentes: Typhlopidae). *Occ. Pap. Museum Zool. Louisiana State Univ.* 46: 1-9.
- Thomas, R. 1975. The *argus* group of West Indian *Sphaerodactylus* (Sauria: Gekkonidae). *Herpetologica* 31: 177-195.
- Thomas, R. 1976. Systematics of Antillean blind snakes of the genus *Typhlops* (Serpentes: Typhlopidae). Ph.D. dissertation, Louisiana State Univ., Baton Rouge.
- Thomas, R. 1982. A new dwarf *Sphaerodactylus* from Haiti (Lacertilia: Gekkonidae). *Proc. Biol. Soc. Washington* 95: 81-88.
- Thomas, R. 1989. The relationships of Antillean *Typhlops* (Serpentes: Typhlopidae) and the description of three new Hispaniolan species, pp. 409-432 in *Biogeography of the West Indies: Past, Present, and Future*, edited by C. A. Woods. Sandhill Crane Press, Gainesville, FL.
- Thomas, R. 1996. Recollections of Albert Schwartz, pp. 16-18 in *Contributions to West Indian Herpetology: A Tribute to Albert Schwartz*, edited by R. Powell, and R. W. Henderson. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.
- Thomas, R., and A. Gaa. 1998. The non-anoline reptiles of the El Verde forest, in *The Food Web of a Tropical Rain Forest*, edited by R. P. Reagan, and R. B. Waide. Univ. Chicago Press, Chicago.
- Thomas, R., and S. B. Hedges. 1988. Two new geckos (*Sphaerodactylus*) from the Sierra Martin Garcia of Hispaniola. *Herpetologica* 44: 96-104.

- Thomas, R., and S. B. Hedges. 1989. A new *Celestus* (Sauria: Anguinae) from the Chaîne de la Selle of Haiti. *Copeia* 1989: 886-891.
- Thomas, R., and S. B. Hedges. 1992. An unusual new *Sphaerodactylus* from Hispaniola (Squamata: Gekkonidae). *J. Herpetol.* 26: 289-292.
- Thomas, R., and S. B. Hedges. 1993. A new banded *Sphaerodactylus* from eastern Hispaniola (Squamata: Gekkonidae). *Herpetologica* 49: 350-354.
- Thomas, R., S. B. Hedges, and O. H. Garrido. 1992. Two new species of *Sphaerodactylus* from eastern Cuba (Squamata: Gekkonidae). *Herpetologica* 48: 358-367.
- Thomas, R., and R. Joglar. 1996. The herpetology of Puerto Rico: Past, present and future. *Annals New York Acad. Sci.* 776: 181-200.
- Thomas, R. and A. G. Kessler. 1996. Non-anoline reptiles. The Food Web of a Tropical Rain Forest, edited by D. P. Reagan, and R. B. Waide. Univ. Chicago Press, Chicago.
- Thomas, R., R. W. McDiarmid, and F. G. Thompson. 1985. Three new species of thread snakes (Serpentes: Leptotyphlopidae) from Hispaniola. *Proc. Biol. Soc. Washington* 98: 204-220.
- Thomas, R., and J. A. Prieto-Hernández. 1985. The use of venom by the Puerto Rican snake, *Alsophis portoricensis*. *Decim. Simp. Recur. Nat.* 1983: 13-22a.
- Thomas, R., and A. Schwartz. 1965. Hispaniolan snakes of the genus *Dromicus* (Colubridae). *Rev. Biol. Tropics*. 13: 59-83.
- Thomas, R., and A. Schwartz. 1966. *Sphaerodactylus* (Gekkonidae) in the Greater Puerto Rico region. *Bull. Florida State Museum* 10: 193-260.
- Thomas, R., and A. Schwartz. 1967. The *monticola* group of the lizard genus *Anolis* in Hispaniola. *Breviora* 261: 1-27.
- Thomas, R., and A. Schwartz. 1974. The status of *Sphaerodactylus gilvitorques* Cope and of *Sphaerodactylus nigropunctatus* Gray (Sauria: Gekkonidae). *J. Herpetol.* 8: 353-358.
- Thomas, R., and A. Schwartz. 1977. Three new species of *Sphaerodactylus* (Sauria: Gekkonidae) from Hispaniola. *Ann. Carnegie Museum* 46: 33-43.

- Thomas, R., and A. Schwartz. 1983a. The *difficilis* complex of *Sphaerodactylus* (Sauria, Gekkonidae) of Hispaniola. Pt. 2. *Sphaerodactylus savagei*, *S. cochranae*, *S. darlingtoni*, *S. armstrongi*, *S. streptophorus*, and conclusions. Bull. Carnegie Museum Nat. Hist. 22: 31-60.
- Thomas, R., and A. Schwartz. 1983b. Variation in Hispaniolan *Sphaerodactylus* (Sauria, Gekkonidae), pp. 86-98 in *Advances in Herpetology and Evolutionary Biology. Essays in Honor of Ernest E. Williams*, edited by A. G. J. Rhodin, and K. Miyata. Museum Comp. Zool., Cambridge, MA.
- Thompson, R. L. 1996. Larval habitat, ecology, and parental investment of *Osteopilus brunneus* (Hylidae), pp. 259-269 in *Contributions to West Indian Herpetology: a Tribute to Albert Schwartz*, edited by R. Powell, and R. W. Henderson. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.
- Thorbjarnarason, J. B. 1988. The status and ecology of the American crocodile in Haiti. Bull. Florida State Museum, Biol. Sci. 33: 1-86.
- Tihen, J. A. 1962. Osteological observations on New World *Bufo*. Am. Midl. Natur. 67:157-183.
- Toft, C. A. 1980. Seasonal variation in populations of Panamanian litter frogs and their prey: a comparison of wetter and drier sites. *Oecologia* 47: 34-36.
- Tolson, P. J. 1987. Phylogenetics of the boid snake genus *Epicrates* and Caribbean vicariance theory. Occ. Papers. Museum Zool., Univ. Michigan 715: 1-68.
- Tolson, P. J. 1988. Critical habitat, predator pressures, and the management of *Epicrates monensis* (Serpentes: Boidae) on the Puerto Rico Bank: a multivariate analysis, pp. 228-238 in *Management of Amphibians, Reptiles, and Small Mammals in North America*, Gen. Tech. Rept. U. S. Dept. of Agriculture, Washington, DC.
- Tolson, P. J. 1996. Conservation of *Epicrates monensis* on the satellite islands of Puerto Rico, pp. 407-416 in *Contributions to West Indian Herpetology: A Tribute to Albert Schwartz*, edited by R. Powell, and R. W. Henderson. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.

- Tolson, P. J., and R. W. Henderson. 1993. The Natural History of West Indian Boas. R&A, Somerset, UK.
- Townsend, D. S., and M. M. Stewart. 1986. The effect of temperature on development in a terrestrial neotropical *Eleutherodactylus*. *Copeia* 1986: 520-523.
- Townsend, D. S. 1996. Patterns of parental care in frogs of the genus *Eleutherodactylus*, pp. 229-239 in Contributions to West Indian Herpetology: a Tribute to Albert Schwartz, edited by R. Powell, and R. W. Henderson. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.
- Trivers, R. L. 1976. Sexual selection and resource-accruing abilities in *Anolis garmani*. *Evolution* 30: 253-269.
- Trueb, L. 1970. Evolutionary relationships of casque-headed tree frogs with co-ossified skulls (family Hyliidae). *Univ. Kansas Pub. Museum Nat. Hist.* 18: 547-716.
- Trueb, L. 1972. *Hyla crucialis* Harlan, 1826 (Amphibia): proposed suppression under the plenary powers Z.N.(S.) 1982. *Bull. Zool. Nomencl.* 29: 39-40.
- Trueb, L., and M. J. Tyler. 1974. Systematics and evolution of the Greater Antillean hylid frogs. *Occas. Pap. Museum Nat. Hist., Univ. Kansas* 24: 1-60.
- Tschudi, J. J. 1838. Classification der Batrachier, mit Berücksichtigung der fossilen Thiere dieser Abtheilung der Reptilien. *Mems. Soc. Sci. Nat. Neuchtel* 2: 1-100.
- Tuck, R. G., and J. D. Hardy. 1973. Status of the Ober Tobago collection, Smithsonian Institution, and the proper allocation of *Amieva suranamensis tobaganus* Cope (Sauria: Teiidae). *Proc. Biol. Soc. Washington* 86: 231-242.
- Turner, F. B. 1960. Estimation of a Louisiana population of the skink, *Lygosoma laterale*. *Ecology* 41: 574-577.
- Turner, F. B. 1977. The dynamics of populations of squamates, crocodylians, and rhynchocephalians, pp. 157-264 in *Ecology and Behaviour*, edited by C. Gans, and D. W. Tinkle. Academic Press, New York.
- Turner, R. D. 1960. Land shells of Navassa Island, West Indies. *Bull. Museum Comp. Zool.* 122: 233-244.

- Underwood, G. 1949. Notes on a hitherto unpublished Gosse illustration of Jamaican herpetology. *Nat. Hist. Notes Nat. Hist. Soc. Jamaica* 4: 46-48.
- Underwood, G. 1950. Reptiles at Clydesdale. *Nat. Hist. Notes Nat. Hist. Soc. Jamaica* 4: 84.
- Underwood, G. 1951. Reptilian retinas. *Nature* 167: 209-213.
- Underwood, G. 1951-52. Introduction to the study of Jamaican reptiles (parts 1-10). *Nat. Hist. Notes Nat. Hist. Soc. Jamaica* 4-5: 104-107, 127-141, 144-149, 161-163, 209-213, 253-257, 15-21, 54-65, 77-82, 97-105.
- Underwood, G. 1954a. Categories of adaptation. *Evolution* 8: 365-379.
- Underwood, G. 1954b. On the classification and evolution of geckos. *Proc. Zool. Soc. London* 124: 469-492.
- Underwood, G. 1954c. The biological resources of Jamaica (Presidential Address). *Nat. Hist. Notes Nat. Hist. Soc. Jamaica* 6: 163-165.
- Underwood, G. 1954d. A Cuban pond-turtle in Jamaica. *Nat. Hist. Notes Nat. Hist. Soc. Jamaica* 6: 120.
- Underwood, G. 1954e. The distribution of Antillean reptiles. *Nat. Hist. Notes Nat. Hist. Soc. Jamaica* 6: 121-129.
- Underwood, G. 1959a. A new Jamaican galliwasp (Sauria, Anguinae). *Breviora* 102: 1-13.
- Underwood, G. 1959b. The anoles of the eastern Caribbean, Part III. Revisionary notes. *Bull. Museum Comp. Zool.* 121: 191-226.
- Underwood, G. 1962. Reptiles of the eastern Caribbean. *Carib. Affairs, New Series* 1: 1-192.
- Underwood, G. 1964. An anguid lizard from the Leeward islands. *Breviora* 200: 1-10.
- Underwood, G. 1976. A systematic analysis of boid snakes, pp. 151-175 in *Morphology and Biology of Reptiles*, edited by Ad'A. Bellairs, and C. B. Cox. Linnaean Society Symposium Series 3, London.

- Underwood, G. 1977. Simplification and degeneration in the course of evolution of Squamate reptiles. *Colloq. Int. C.N.R.S.* 266: 341-352.
- Underwood, G. 1993. A new snake from St. Lucia, West Indies. *Bull. Nat. Hist. Museum (Zool.)* 59: 1-9.
- Underwood, G., and E. E. Williams. 1959. The anoline lizards of Jamaica. *Bull. Inst. of Jamaica. Sci. Ser.* 9: 1-48.
- Valdés, A., and F. Ruíz. 1980. Consideraciones sistemáticas sobre *Bufo longinasus* (Anura: Bufonidae) de Cuba y descripción de una nueva subespecie. *Poeyana* 206: 1-33.
- Valdés de la Osa, A., and F. N. R. García. 1980. Consideraciones sistemáticas sobre *Bufo longinasus* (Anura: Bufonidae) de Cuba y descripción de una nueva subespecie. *Poeyana* 206: 1-33.
- Valdez Sierra, G., and J. M. M. Félix. 1992. Sistema de Areas Protegidas de República Dominicana. *Dir. Nac. Parques, Santo Domingo.*
- Vane-Wright, R. I., C. J. Humphries, and P. H. Williams. 1991. What to protect? - Systematics and the agony of choice. *Biol. Conserv.* 55: 235-254.
- Vanzolini, P. E. 1951. A systematic arrangement of the family Amphisbaenidae (Sauria). *Herpetologica* 7: 113-123.
- Vanzolini, P. E. 1961. *Bachia*: espécies brasileiras e conceito genérico (Sauria, Teiidae). *Pap. Avulsos S. Paulo* 14: 193-209.
- Vanzolini, P. E. 1968. Geography of the South American Gekkonidae (Sauria). *Archos Zool. Est. S. Paulo* 17:85-112.
- Vanzolini, P. E. 1978. On South American *Hemidactylus* (Sauria, Gekkonidae). *Pap. Avul. Zool.* 31: 307-343.
- Vanzolini, P. E. 1990. Geographic Distribution: *Gymnophthalmus underwoodi*. *Herpetol. Review* 21: 96.
- Vanzolini, P. E., and E. E. Williams. 1962. Jamaican and Hispaniolan *Gonatotodes* and allied forms (Sauria, Gekkonidae). *Bull. Museum Comp. Zool.* 127: 479-498.

- Varona, L. S. 1966. Notas sobre los cocodrilos de Cuba y descripción de una nueva especie del pleistoceno. *Poeyana* (serie A) 16: 1-34.
- Varona, L. S. 1976. *Caiman crocodilus* (Reptilia: Alligatoridae) en Cuba. *Misc. Zool.* 5: 2.
- Villa, J. 1984. The venomous snakes of Nicaragua. *Milwaukee Public Museum Contr. Biol. Geol.* 59: 1-41.
- Villa, J., L. D. Wilson, and J. D. Johnson. 1988. Middle American Herpetology: A Bibliographic Checklist. Univ. Missouri Press, Columbia.
- Vitt, L. J., and C. M. de Carvalho. 1995. Niche partitioning in a tropical wet season: lizards in the Lavrado area of northern Brazil. *Copeia* 1995: 305-329.
- Vitt, L. J., and L. D. Vangilder. 1983. Ecology of a snake community in north-eastern Brazil. *Amphib.-Rept.* 4: 273-296.
- Vitt, L. J., and P. A. Zani. 1996a. Ecology of the lizard *Ameiva festiva* (Teiidae) in southeastern Nicaragua. *J. Herpetol.* 30: 110-117.
- Vitt, L. J., and P. A. Zani. 1996b. Ecology of the South American lizard *Norops chrysolepis* (Polychrotidae). *Copeia* 1996: 56-68.
- Vogel, P. 1983. On an ant-lizard interaction. *Oecologia* 58: 269-271.
- Vogel, P. 1984. Seasonal hatchling recruitment and juvenile growth of the lizard *Anolis lineatopus*. *Copeia* 1984: 747-757.
- Vogel, P., and F. v. Brockhusen-Holzer. 1984. Ants as prey of juvenile *Anolis lineatopus* (Rept., Iguanidae) in prey choice experiments. *Zeitschr. Tierpsychol.* 65: 66-76.
- Vogel, P., and E. Curio. 1983. Response of a predator to harmful and similar harmless prey. *Verh. Deutsch. Zool. Gesell.* 247: 247.
- Vogel, P., W. Hettrich, and K. Ricono. 1986. Weight growth of juvenile lizards, *Anolis lineatopus*, maintained on different diets. *J. Herpetol.* 20: 50-58.
- Vogel, P., R. Nelson, and R. Kerr. 1996. Conservation strategy for the Jamaican iguana, *Cyclura collei*, pp. 395-406 in *Contributions to West Indian Herpe-*

tology: A Tribute to Albert Schwartz, edited by R. Powell, and R. W. Henderson. Society for the Study of Amphibians and Reptiles Contributions in Herpetology, Ithaca, NY.

Wagler, J. G. 1830. Natürliches System der Amphibien, mit vorangehender Classification der Säugethiere und Vögel. Ein Beitrag zur vergleichenden Zoologie. J. G. Cotta, München.

Wake, D. B. 1987. Adaptive radiation of salamanders in Middle American cloud forests. *Ann. Missouri Bot. Gard.* 74: 242-264.

Wake, D. B., and P. Elias. 1983. New genera and a new species of Central American salamanders, with a review of the tropical genera (Amphibia, Caudata, Plethodontidae). *Contrib. Sci. Nat. Hist. Museum Los Angeles County* 345: 1-19.

Wake, D. B., and J. D. Johnson. 1989. A new genus and species of plethodontid salamander from Chiapas, Mexico. *Contrib. Sci. Nat. Hist. Museum Los Angeles County* 411: 1-10.

Wake, D. B., and J. F. Lynch. 1976. The distribution, ecology, and evolutionary history of plethodontid salamanders in tropical America. *Sci. Bull. Nat. Hist. Museum Los Angeles County* 25: 1-65.

Ward, W. C., A. E. Weidie, and W. Back. 1985. Geology and hydrogeology of the Yucatan and Quaternary geology of northeastern Yucatan Peninsula. *New Orleans Geol. Soc.* 1-19.

Weil, T. E., J. K. Black, H. I. Blutstein, K. T. Johnston, D. S. McMorris, and F. P. Munson. 1982. Dominican Republic: A Country Study. 2nd ed. Department of the Army, Washington, DC.

Weinland, D. F. 1862. Beschreibung und Abbildung von drei neuen Sauriern. (*Embryopus habichii* und *Amphisbaena innocens* von Haiti, und *Brachymeles leuckartii* von Neuholland.). *Abh. Senckenb. Naturf. Ges.* 4: 131-143.

Wermuth, H., and R. Mertens. 1961. Schildkröten, Krokodile, Brückenechsen. Fischer-Verlag, Jena, Germany.

Wermuth, H., and R. Mertens. 1977. Liste der rezenten Amphibien und Reptilien: Testudines, Crocodylia, Rhynchocephalia. *Das Tierreich*, Berlin.

- Werner, F. 1909. Ueber neue oder seltene Reptilien des naturhistorischen Museums in Hamburg. 1. Schlangen. Jahrb. Wiss. Anst. Hamburg (2. Beiheft) 26: 205-247.
- Westermann, J. H. 1953. Nature preservation in the Caribbean. A review of literature on the destruction and preservation of flora and fauna in the Caribbean area. Foundation Sci. Res. Surinam Netherland Antilles 9: 1-106.
- Wetmore, A., and B. H. Swales. 1931. The birds of Haiti and the Dominican Republic. Bull. U.S. Natl. Museum 155: 1-483.
- White, L. R., R. Powell, J. J. S. Parmerlee, A. Lathrop, and D. D. Smith. 1992. Food habits of three syntopic reptiles from the Barahona Peninsula, Hispaniola. J. Herpetol. 26: 518-520.
- Wiens, J. J. 1995. Polymorphic characters in phylogenetic systematics. Syst. Biol. 44: 482-500.
- Wijffels, L. C. M. 1997. *Anolis bimaculatus* als huisgenoot. Lacerta 55(5): 201-205.
- Wiley, E. O. 1978. The evolutionary species concept reconsidered. Syst. Zool. 27: 17-26.
- Willet, B. M. 1987. Philips' Certificate atlas for the Caribbean. Philip, London.
- Williams, E. E. 1956. *Pseudemys scripta callirostris* from Venezuela with a general survey of the *scripta* series. Bull. Museum Comp. Zool. 115: 145-160.
- Williams, E. E. 1959. Preface. The anoles of the eastern Caribbean. Bull. Museum comp. Zool. 121: 187-189.
- Williams, E. E. 1960. Notes on Hispaniolan herpetology. 1. *Anolis christophei*, new species, from the Citadel of King Christophe, Haiti. Breviora 117: 1-7.
- Williams, E. E. 1961. Notes on Hispaniolan herpetology 3. The evolution and relationships of the *Anolis semilineatus* group. Breviora 136: 1-8.
- Williams, E. E. 1962a. Notes on Hispaniolan herpetology 6. The giant anoles. Breviora 155: 1-15.

- Williams, E. E. 1962b. Notes on the herpetology of Hispaniola 7. New material of two poorly known anoles: *Anolis monticola* Shreve and *Anolis christophei* Williams. *Breviora* 164: 1-11.
- Williams, E. E. 1962c. The anoles of the northern Leewards, Anguilla to Montserrat: new data and a new species. *Bull. Museum Comp. Zool.* 127: 453-465.
- Williams, E. E. 1963a. Notes on Hispaniolan herpetology 8. The forms related to *Anolis hendersoni* Cochran. *Breviora* 186: 1-13.
- Williams, E. E. 1963b. *Anolis whitemani*, new species from Hispaniola (Sauria, Iguanidae). *Breviora* 197: 1-8.
- Williams, E. E. 1965a. The species of Hispaniolan green anoles (Sauria, Iguanidae). *Breviora* 227: 1-16.
- Williams, E. E. 1965b. Hispaniolan giant anoles (Sauria, Iguanidae): new data and a new subspecies. *Breviora* 232: 1-7.
- Williams, E. E. 1969. The ecology of colonization as seen in the zoogeography of anoline lizards on small islands. *Q. Rev. Biol.* 44: 345-389.
- Williams, E. E. 1972. The origin of faunas. Evolution of lizard congeners in a complex island fauna: a trial analysis, pp. 47-89 in *Evolutionary Biology*, edited by T. Dobzhansky, M. K. Hecht, and W. C. Steere. Appleton-Century-Crofts, New York.
- Williams, E. E. 1975. *Anolis marcanoii* new species: sibling to *Anolis cybotes*: description and field evidence. *Breviora* 430: 1-9.
- Williams, E. E. 1976a. West Indian anoles: a taxonomic and evolutionary summary 1. Introduction and a species list. *Breviora* 440: 1-21.
- Williams, E. E. 1976b. South American anoles: the species groups. *Papeis Avulsos Zool.*, S. Paulo 29(26): 259-268.
- Williams, E. E. 1977. The macrosystematics of the anoles, pp. 122-131 in *The Third Anolis Newsletter*, edited by E. E. Williams. Museum Comp. Zool., Cambridge, MA.

- Williams, E. E. 1983. Ecomorphs, faunas, island size, and diverse end points in island radiations of *Anolis*, pp. 326-370 in *Lizard Ecology*, edited by R. B. Huey, E. R. Pianka, and T. W. Schoener. Harvard Univ. Press, Cambridge, MA.
- Williams, E. E. 1989a. A critique of Guyer and Savage (1986): cladistic relationships among anoles (Sauria: Iguanidae): are the data available to reclassify the anoles?, pp. 433-478 in *Biogeography of the West Indies: Past, Present, and Future*, edited by C. A. Woods. Sandhill Crane Press, Gainesville, FL.
- Williams, E. E. 1989b. Old problems and new opportunities in West Indian biogeography, pp. 1-46 in *Biogeography of the West Indies: Past, Present, and Future*, edited by C. A. Woods. Sandhill Crane Press, Inc., Gainesville, FL.
- Williams, E. E., and S. M. Case. 1986. Interactions among members of the *Anolis distichus* complex in and near the Sierra de Baoruco, Dominican Republic. *J. Herpetol.* 20: 535-546.
- Williams, E. E., and A. S. Rand. 1961. Notes on Hispaniolan herpetology 2. A review of the *Anolis semilineatus* group with the description of *Anolis cochranæ*, new species. *Breviora* 135: 1-11.
- Williams, E. E., and A. S. Rand. 1969. *Anolis insolitus*, a new dwarf anole of zoogeographic importance from the mountains of the Dominican Republic. *Breviora* 326: 1-21.
- Williams, E. E., and T. P. Webster. 1974. *Anolis rupinae* new species a syntopic sibling of *A. monticola* Shreve. *Breviora* 429: 1-22.
- Williamson, M. 1981. *Island populations*. Oxford Univ. Press.
- Wilson, E. O. 1961. The nature of the taxon cycle in the Melanesian ant fauna. *Am. Nat.* 95: 169-193.
- Wilson, E. O. 1991. *Ants. Wings* 1991: 4-13.
- Wilson, L. D., J. R. McCranie, and G. A. Cruz. 1994. A new species of *Plectrohyla* (Anura: Hylidae) from a premontane rainforest in northern Honduras. *Proc. Biol. Soc. Washington* 107: 67-78.
- Wilson, L. D., and J. R. Meyer. 1985. *The Snakes of Honduras*. Milwaukee Public Museum, Milwaukee, WI.

- Wilson, L. D., L. Porras, and J. R. McCranie. 1986. Distributional and taxonomic comments on some members of the Honduran herpetofauna. *Contributions Biol. Geol. Milwaukee Publ. Museum* 66: 1-18.
- Wolcott, G. N. 1923. The food of Porto Rican lizards. *J. Dept. Agric. Porto Rico* 7: 5-38.
- Wood, G. C. 1939. The genus *Tretanorhinus* in Cuba and the Isle of Pines. *Proc. New England Zool. Club* 18: 5-11.
- Woodley, J. D. 1971. Hellshire Hills Scientific Survey, 1970. *Sci. Rep. Univ. West Indies*.
- Woodley, J. D. 1980. Survival of the Jamaican iguana, *Cyclura collei*. *J. Herpetol.* 14: 45-49.
- Woods, C. A. 1989. *Biogeography of the West Indies: Past, Present, and Future*. Sandhill Crane Press, Gainesville, FL.
- Woolbright, L. L. 1985. Patterns of nocturnal movement and calling by the tropical frog, *Eleutherodactylus coqui*. *Herpetologica* 41: 1-9.
- Woolbright, L. L., and M. M. Stewart. 1987. Foraging success of a tropical frog *Eleutherodactylus coqui*: the cost of calling. *Copeia* 1987: 69-75.
- Wunderle, J. M., Jr. 1981. Avian predation upon *Anolis* lizards on Grenada. *Herpetologica* 37: 104-108.
- Wüster, W., M. G. Salomao, R. S. Thorpe, G. Puerto, M. F. D. Furtado, S. A. Hoge, R. D. G. Theakston, and D. A. Warrel. 1997. Systematics of the *Bothrops atrox* complex: new insights from multivariate analysis and mitochondrial DNA sequence information, pp. 99-113 in *Venomous Snakes: Ecology, Evolution and Snakebite*, edited by R. S. Thorpe, W. Wüster, and A. Malhorta. Clarendon Press, Oxford, UK.
- Wyles, J. S., and G. C. Gorman. 1980. The classification of *Anolis*: Conflict between genetic and osteological interpretation as exemplified by *Anolis cybotes*. *J. Herpetol.* 14: 149-153.

- Yang, S. Y., M. Soulé, and G. C. Gorman. 1974. *Anolis* lizards of the eastern Caribbean: A case study in evolution. I. Genetic relationships, phylogeny, and colonization sequence of the *roquet* group. *Syst. Zool.* 23: 387-399.
- Zug, G. R., and A. Schwartz. 1958. Variation in the species of *Cadea* (Amphisbaenidae), and a record of *C. blanooides* from the Isla de Pinos. *Herpetologica* 14: 176-179.
- Zweifel, R. G. 1967. Identity of the frog *Cornufer unicolor* and application of the generic name *Cornufe*. *Copeia* 1967: 117-121.

Systematic Index

A

- Abronia*, 362
Acrantophis dumerili, 308
Adelphicos, 362
Agalychnis, 361, 363
Agave, xxv
Ahaetulla, 146, 158
 frenata, 146, 158
Aloponotus, 152
 ricordi, 152
 ricordii, 152
Alsophis, 4, 11, 20, 51, 60, 67, 76, 114, 130, 136, 146, 157, 165, 171, 173, 174, 176, 186, 215, 216, 217, 219, 221, 227, 230, 232, 243, 251, 252, 254, 256, 260, 267, 268, 312, 315, 316, 317, 318, 319, 333, 354, 362
 anomalus, 114, 130, 136, 146, 157, 316
 antiquae, 186, 216, 219
 a. sajdaki, xxix, 216, 219
 antillensis, 186, 215, 216, 317
 a. danforthi, 216
 a. manselli, xxvii, 216
 a. sanctonum, 216
 a. sibonius, 216
 ater, 11, 20, 67, 76, 227, 316
 cantherigerus, xxiii, 51, 60, 251, 252, 315, 316, 317
 c. adpersus, 51, 61
 c. cantherigerus, 51, 60
 c. pepeii, 51, 61
 c. schwartzii, 51, 60
 cinereus, 216
 elegans, 317
 melanichnus, 114, 130, 136, 157, 165, 316
 portoricensis, 173, 174, 230, 316, 317
 punctifer, 216
 rijersmai, 186, 216, 219, 317
 rufiventris, 186, 216, 317
 sancticrucis, 176
 vudii, 316, 317
Amazona, 353
 vittata, 351
Amblyrhynchus, 286
Ameiva, 1, 3, 5, 8, 12, 15, 18, 28, 29, 49, 60, 67, 71, 75, 111, 112, 118, 129, 135, 146, 155, 161, 162, 163, 173, 174, 175, 176, 185, 190, 191, 199, 201, 209, 219, 223, 227, 229, 232, 238, 239, 240, 243, 244, 248, 251, 255, 256, 257, 262, 267, 299, 300, 301, 332, 354, 364
 ameiva, 185, 239
 a. aquilina, 201
 a. tobagana, 201
 analifera, 200
 aquilina, 200
 atrata, 185, 219
 auberi, 28, 29, 49, 60, 301
 a. abducta, 49
 a. atrothorax, 49
 a. auberi, 49
 a. cacuminus, 49
 a. citra, 49
 a. denticola, 49
 a. extorris, 49
 a. extraria, 49
 a. galbiceps, 49
 a. garridoi, 49
 a. gemmea, 49
 a. granti, 49
 a. hardyi, 49
 a. llanensis, 49
 a. marcida, 49
 a. nigriventris, 49
 a. orlandoi, 49
 a. paulsoni, 49
 a. peradusta, 49
 a. procer, 49
 a. pullata, 49
 a. sabulicolor, 49
 a. sanfelipensis, 49
 a. schwartzii, 49
 a. secta, 49
 a. sublesta, 49
 a. ustulata, 49
 a. zugii, 49
 barbouri, 155
 beatensis, 155
 chrysolaeama, 28, 111, 118, 129, 135, 146, 155, 162, 163, 238, 257, 299, 300, 301
 c. abbotti, 111

- c. alacris*, 111
c. boekeri, 111, 155
c. chrysolaeama, 111
c. defensor, 111
c. evulsa, 111, 155
c. ficta, 111, 155
c. jacta, 111
c. parvoris, 111
c. procax, 111
c. quadrijugis, 111, 155
c. regularis, 112
c. richardthomasi, 112
c. secessa, 112
c. umbratilis, 112
c. woodi, 112, 155
cinereacea, 185, 190, 200, 201, 300, 354
corax, 185, 201, 219
corvina, 185, 201
dorsalis, 8, 18, 67, 71, 75, 227, 301
erythrocephala, 15, 185, 219, 300, 301
exsul, 174, 175, 229, 257, 301
fuscata, 185, 238, 240, 255, 300
griswoldi, xxvii, 185, 200, 262, 300, 301
leberi, 112, 129, 135, 155
lineolata, 28, 112, 129, 155, 300, 301
 l. beatensis, 112
 l. lineolata, 112
 l. meracula, 112
 l. perplicata, 112
 l. privigna, 112
 l. semota, 112
major, 185, 190, 201, 300, 354, 360
maynardi, 301
navassae, 155, 161
nevisana, 200
plei, 181, 200, 201, 229, 257, 300
pluvianotata, 185, 201, 300
 p. atrata, 201
polops, 176, 300, 301
quadri-lineata, 239
regularis, 146, 155
rosamondae, 155
scutata, 200
surinamensis tobaganus, 201
taeniura, 28, 112, 129, 135, 155, 161, 162, 163, 257, 300, 301
 t. aequorea, 112
 t. azuae, 112
 t. barbouri, 112
 t. ignobilis, 112
 t. meyerabichi, 112, 155
 t. pentamerinthus, 112, 156
 t. regnatrix, 112
 t. rosamondae, 112
 t. taeniura, 112
 t. tofacea, 112
 t. vafra, 112
 t. varica, 112
 t. vulcanalis, 112
undulata, 299
wetmorei, 173, 300, 301
Amiva [sic] vittipunctata, 155, 299, 300
Amphisbaena, 2, 4, 5, 12, 16, 25, 45, 53, 104, 118, 124, 125, 132, 146, 148, 149, 172, 173, 174, 232, 256, 278, 337
 bakeri, 173, 279
 blanoides, 45, 53
 caeca, 16, 25, 279
 caudalis, 104, 124, 132, 149
 cubana, 45, 53, 279
 c. barbouri, 45, 53, 279
 c. cubana, 45, 53, 279
 fenestrata, 174, 279
 gonavensis, 104, 124, 132, 148, 149
 hyporissor, 104, 118, 124, 132, 149
 h. hyporissor, 104
 h. leberi, 104, 125
 innocens, 104, 124, 132, 146, 149, 279
 manni, 104, 124, 132, 279
 palirostrata, 45, 53
 punctata, 2, 5, 309
 schmidti, 173
 weinlandi, 149
 xera, 173
Anguis, 12, 15, 159
 jamaicensis, 15
Anolis, xxvi, xxvii, 2, 3, 8-10, 12-16, 18, 19-23, 26, 28, 29, 32, 33, 46-49, 55-57, 66, 71, 74-79, 82, 85, 86, 88, 102, 108-111, 127, 131, 134, 135, 137, 138, 140, 144, 146-148, 152-154, 161-163, 171, 172, 174,

- 175-177, 185, 190, 191, 193, 194, 199, 209, 219, 220, 224, 225, 227-232, 234-236, 239, 240, 242, 244, 246, 247-257, 262-265, 268, 282, 289, 291, 292, 294, 345, 347, 354, 360, 364, 366, 368, 369
- acutus*, 26, 176, 236, 294
- aeneus*, 185, 194, 213, 229, 236, 240
- ahli*, 46, 57
- alayoni*, 46, 57
- alfaroi*, 46, 57
- alligator*, 22, 190
- aliniger*, 108, 127, 134, 147, 154
- allisoni*, 46, 55
- allogus*, 46, 56
- altavelensis*, 108, 128, 134, 153, 154
- alumina*, 108, 134, 148
- alutaceus*, 46, 55
- anfiloquioi*, 46, 57
- angusticeps*, xxviii, 46, 55, 236
- a. angusticeps*, 46, 56
- a. ologapsi*, 56
- antigua*, 191
- argenteolus*, 46, 57
- argillaceus*, 46, 57
- armouri*, 108, 134, 153
- asper*, 22, 190, 191
- baharucuensis*, 26
- bahorucoensis*, xxv, 26, 108, 134, 154, 254
- b. southerlandi*, 108
- b. bahorucoensis*, 108
- baleatus*, 108, 128, 134, 152
- b. altager*, 108
- b. caeruleolatus*, 108
- b. fraudator*, 108
- b. lineatacervix*, 108, 128
- b. litorisilva*, 108
- b. multistruppus*, 108
- b. samanae*, 108
- b. scelestus*, 108
- b. sublimis*, 108
- baracoae*, 46, 57
- barahonae*, 108, 134, 153, 154, 267
- b. albocellatus*, 108
- b. barahonae*, 108
- b. ininquinatus*, 108
- b. mulitus*, 108
- barbatus*, 46, 56
- barbouri*, xxiv, 108, 128, 134, 153, 263
- bartischi*, 46, 56
- biauritus*, 154
- bimaculatus*, 177, 185, 191, 192, 194, 236, 262, 294, 295, 354, 360
- b. leachi*, 194
- birama*, 46, 57
- blanquillanus*, 294
- bonairensis*, 239, 294
- bremeri*, 46, 56
- b. insulaepinarum*, 47, 56
- brevirostris*, 30, 108, 127, 134, 146, 148, 153, 154, 163, 236, 262
- b. bremeri*, 46, 56
- b. brevirrostris*, 108
- b. deserticola*, 108
- b. wetmorei*, 108
- carolinensis*, 16, 161, 270, 297
- caudalis*, xxviii, 108, 127, 128, 134, 153, 154
- centralis*, 47, 57
- c. centralis*, 47, 57
- c. litoralis*, 47, 57
- chamaeleonides*, xxiii, 47, 55
- chlorocyanus*, 109, 127, 134, 147, 154, 162, 163, 255, 262
- c. aliniger*, 154
- c. chlorocyanus*, 109
- c. cyanostictus*, 109
- christophei*, 109, 134
- cinereus*, 22, 190
- clivicola*, 47, 57
- cochranae*, 154
- coelestinus*, 109, 128, 134, 147, 152, 154, 161, 163, 257, 262
- c. coelestinus*, 109, 128
- c. demissus*, 109, 128
- c. pecuarius*, 109, 128
- confusus*, 47, 57
- conspersus*, 23
- cooki*, 173
- cristatellus*, xxvi, 22, 102, 109, 127, 131, 153, 172, 174, 176, 230, 237, 257, 262, 296, 360
- c. cristatellus*, 109
- cupeyalensis*, 47, 57

- cuvieri*, xxviii, 15, 16, 173, 205, 237, 360
cyanopleurus, 47, 57
 c. cyanopleurus, 57
 c. orientalis, 47, 57
cybotes, xxvi, xxviii, 27, 109, 127, 134, 137, 147, 152, 154, 162, 163, 237, 255, 257, 262
 c. cybotes, 109, 128
 c. doris, 109, 128, 154
 c. ravifaux, 109, 128
darlingtoni, 109, 134, 153
delafuentei, 47, 57
desechensis, 176
distichus, 29, 109, 127, 134, 146, 147, 153, 154, 161, 163, 237, 262
 d. aurifer, 109
 d. dominicensis, 29, 30, 109, 146, 154, 163
 d. favillarum, 30, 109
 d. ignigularis, 29, 109
 d. juliae, 109, 128
 d. patruelis, 109, 128
 d. properus, 109
 d. ravitergum, 30, 109
 d. sejunctus, 109, 128
 d. suppar, 109
 d. tostus, 109, 128
 d. vinosus, 109
dolichocephalus, 110, 134, 154
 d. dolichocephalus, 110
 d. portusalus, 110
 d. sarmenticola, 110
dominicanus, 162
dominicensis, 154
 d. altavelensis, 154
 d. caudalis, 154
doris, 147
edwardsii, 12, 20, 23
equestris, 15, 47, 55, 257
 e. buidei, 47
 e. cinoleguas, 47
 e. equestris, 47
 e. juraguensis, 47
 e. persparsus, 47
 e. potior, 47
 e. thomasi, 47
 e. verreonensis, 47
ernestwilliamsi, 174
etheridgei, 110, 134, 153
eugenegrahami, 27, 110, 134, 153
evermanni, xxviii, 237
extremus, 22, 185, 190, 191, 194
ferreus, 185, 192
flabellatus, 19, 77
forresti, 26, 191, 192
fowleri, 27, 110, 134
fuscoauratus, 239, 240
fugitivus, 47, 57
garmani, 8, 9, 10, 23, 66, 227, 292, 297
garridoi, 47, 57
gentilis, 22, 190
gingivinus, 22, 26, 185, 190, 192, 194, 237
grahami, xxiv, 18, 19, 23, 75, 79, 85, 227, 257
 g. aquarum, 66
 g. grahami, 66
griseus, 22, 185, 190, 191, 194, 237
guafe, 47, 57
guamuhaya, 47, 57
guazuma, 47, 57
gundlachi, 173, 237, 257
haetianus, 110, 134, 147, 153
hendersoni, 110, 134, 154
 h. bahorucoensis, 154
 h. dolichocephalus, 154
 h. hendersoni, 110
 h. ravidormitans, 110
homolechis, 28, 47, 55
 h. homolechis, 47
 h. turquinensis, 47, 56
humilis, 239, 249
imias, 47, 57
inexpectatus, 47, 57
insolitus, xxviii, 110, 134, 148
iodurus, 19
isolepis, 47, 57
 i. altitudinalis, 47, 58
 i. isolepis, 47, 57
juangundlachi, 47, 56
jubar, 47, 57
 j. albertschwartzi, 47, 58
 j. balaenarum, 47, 58
 j. cocoensis, 47, 58

- j. cuneus*, 48, 58
j. gibarensis, 48, 58
j. jubar, 47, 58
j. maisensis, 48, 58
j. oriens, 48, 58
j. santamariae, 48, 58
j. yaguajayensis, 48, 58
koopmani, 110, 134, 148
krugi, 26, 173
laeviceps, 154
latirostris, 154, 161
leachi, 22, 190, 191, 295
leucomelas, 216
 l. antiquae, 216
 l. manselli, 216
limifrons, 239
lineatopus, 18, 19, 66, 75, 79, 82, 85, 86, 229, 257
 l. ahenobarbus, 66
 l. coxi, 82
 l. lineatopus, 66
 l. merope, 66
 l. neckeri, 66, 82
lividus, 22, 185, 190, 191, 194
longiceps, 138, 140, 161
longitibialis, 110, 134, 153
 l. longitibialis, 110, 128
 l. specuum, 110
loysianus, 48, 55
luciae, 22, 185, 190, 191, 194, 213, 294
lucius, 48, 55
luteogularis, xxviii, 48, 56
 l. calceus, 48
 l. coctilis, 48
 l. delacruzii, 48
 l. hassleri, 48
 l. jaumeai, 48
 l. luteogularis, 48
 l. nivevultus, 48
 l. sanfelipensis, 48
 l. sectilis, 48
macilentus, 48, 57
maculatus, 19
marcanoi, 110, 135, 153
marmoratus, xxvii, 192, 193
marron, 110, 135, 153
mayeri, 191
maynardi, 138, 140
mestrei, xxviii, 48, 56
mimus, 48, 57
monensis, 175, 296
monticola, 110, 135
 m. monticola, 110
 m. quadrisartus, 110
noblei, 48, 57, 106
 n. galeifer, 48, 57
 n. noblei, 48, 57
nubilus, 22, 185, 190, 191, 194, 219
occultus, 173
oculatus, 185, 192, 193, 194, 237, 240, 255, 360
olssoni, xxviii, 110, 127, 135, 161, 237
 o. alienus, 110
 o. dominigensis, 110
 o. extentus, 110
 o. ferrugicauda, 110
 o. insulanus, 110, 128
 o. montivagus, 110
 o. olssoni, 110
 o. palloris, 110
opalinus, 9, 19, 20, 66, 77, 227, 250
ophiolepis, 48, 55
paternus, 48, 56
 p. paternus, 48
 p. pinarensis, 48, 56
pigmaequestris, 48, 58
placidus, 110, 135
polylepis, 369
poncensis, 26, 173
porcatus, xxviii, 48, 55, 110, 127, 131, 138, 140, 144, 262
porcus, 48, 57
pulchellus, xxviii, 26, 173, 174, 175, 237
pumilus, 48, 56
quadriocellifer, 48, 56
reconditus, 66, 71, 80, 85, 292
richardi, 22, 185, 190, 192, 194, 237, 257, 322
ricordii, 110, 127, 153, 154, 162
 r. barahonae, 154
 r. leberi, 110
 r. ricordii, 110
 r. subsolanus, 110

- r. viculus*, 110
rimarum, 110, 135
roosevelti, 174
roquet, 14, 185, 191, 194, 294
rubribarbus, 48, 57
rupinae, 110, 135, 148
sabanus, xxvii, 22, 185, 190, 191, 194, 219
sagrei, 19, 48, 55, 66, 76, 229, 237, 262, 296, 297
 s. greyi, 48, 56
 s. sagrei, 48, 66
scriptus, 22, 176, 296, 360
semilineatus, 111, 127, 135, 147, 154, 161
sheplani, 27, 111, 135
shrevei, 111, 135, 153
sibonius, 216
singularis, 111, 128, 135
smallwoodi, xxiii, 48, 57, 267
 s. palardis, 49
 s. saxuliceps, 49
 s. smallwoodi, 48
spectrum, 49, 56
speciosus, 22, 190, 191, 192
strahmi, 111, 127, 135
 s. abditus, 111
 s. strahmi, 111
stratulus, xxvi, 174, 175, 237, 257
terraealtae, 191
trinitatis, 185, 194, 237
tropidolepis, 369
trossulus, 22, 190, 191
valencienni, 1, 19, 66, 74, 76, 250, 297
vanidicus, 49
 v. rejectus, 49
 v. vanidicus, 49
vermiculatus, 49, 56, 255, 257
vescus, 49
vicentii, 22
vincenti, 190, 191
virgatus, 22, 190
wattsi, 22, 26, 185, 191, 193, 194, 237, 294, 354
websteri, 111, 135
whitemani, xxiv, 111, 128, 135, 153
 w. breslini, 111
 w. lapidosus, 111
 w. whitemani, 111
Anothea, 361
Antillophis, 4, 5, 51, 60, 61, 114, 130, 136, 137, 146, 147, 157, 164, 243, 253, 256, 260, 267, 312, 315, 317, 318, 319, 333, 362
 andreae, 51, 61, 315, 317
 a. andreae, 51, 61
 a. melopyrrha, 51, 61
 a. morenoi, 51, 61
 a. nebulatus, 51, 61
 a. orientalis, 51, 61
 a. peninsulae, 51, 61
 parvifrons, 114, 130, 136, 137, 146, 147, 157, 164, 253, 315, 317, 318
 p. alleni, 114, 130
 p. lincolni, 114, 130
 p. niger, 114
 p. paraniger, 114, 130
 p. parvifrons, 114, 130
 p. protenus, 114, 146, 157
 p. rosamondae, 114, 130
 p. stygius, 114, 130
 p. tortuganus, 114, 130
Aristelligella, 150, 280
Aristelliger, 1, 3, 66, 76, 80, 91, 105, 118, 126, 132, 138, 140, 150, 162, 163, 227, 242, 246, 267, 280, 282, 286, 331
 barbouri, 150, 280
 cochranae, 138, 140, 150, 162, 280
 expectatus, 105, 118, 126, 132, 138, 140, 150, 162, 163
 georgeensis, 280
 hechti, 280
 lar, 80, 91, 105, 126, 132, 150, 162, 280
 praesignis, 66, 76, 80, 91, 227, 280
 p. nelsoni, 91
 titan, 91
Arrhyton, 4, 11, 51, 60, 61, 67, 71, 76, 87, 158, 173, 174, 232, 243, 256, 260, 312, 315, 316, 318, 333, 362
 ainictum, 51, 61
 callilaemum, 67, 315, 318
 dolichura, 51, 61
 exiguum, xxvi, 173, 174, 315, 318, 319
 funereum, 11, 67, 315, 318

funereus, 76, 315
landoi, 51, 61, 315, 318
polylepis, 67, 71, 87, 239, 315, 318
procerum, 51, 61
supernum, 51, 61
taeniatum, 51, 61, 315
tanyplectum, 51, 61
vittatum, 51, 61
Atelophryniscus, 363
Atractaspis, 322
Atractus, 251, 365
Audantia shrevei, 153
Avicularia laeta, 257

B

Bachia, 185, 190, 201, 205, 242, 299, 301, 332, 366
alleni
a. parviceps, 205
a. alleni, 205
heteropa, 205
h. alleni, 206
heteropus, 185, 301
pallidiceps, 301
Basiliscus, 363
Boa, 7, 12, 20, 23, 186, 213, 214, 243, 256, 306, 308, 333, 354, 363
constrictor, 213, 214, 306, 308, 354
c. orophias, 213, 214
c. nebulosus, 213, 214, 239
enydris, 214
grenadensis, 214
madagascariensis, 308
manditra, 308
Bolitoglossa, 363
Bothriechis, 362
Bothriopsis, 324
Bothrops, 4, 14, 186, 214, 243, 324, 325, 334, 363, 366
asper, 325
atrox, 214, 325, 326
caribbaea, 186, 214, 325
lanceolata, 14, 186, 214, 325
Bubulcus ibis, 257
Bufo, 1, 3, 38, 39, 41, 42, 65, 99, 118, 119, 120, 121, 139, 142, 166, 174, 178, 185,

187, 232, 242, 245, 249, 256, 265, 266, 270, 361
cataulaciceps, 39, 42,
empusus, 39, 41
fluviaticus, 99, 119, 121, 142
fractus, 99, 118, 119, 121, 142
fustiger, 39, 42
granulosa, 271
granulosus, 361
guentheri, 99, 119, 121, 142, 249
gundlachi, 39, 41
gutturatus, 142
haematiticus, 249
lemur, 174, 249
longinasus, 39, 41, 249
l. cajalbanensis, 39, 41
l. dunni, 39, 41
l. longinasus, 39
l. ramsdeni, 39, 42
marinus, 38, 41, 65, 99, 119, 120, 139, 142, 166, 178, 185, 187, 265, 270
peltocephalus, xxii, 39, 42, 119, 249
strumosus, 142
taladai, 39, 42
Burhinus, 351
Buteo, 257
jamaicensis, 257
platypterus, 257
ridgewayi, 257

C

Cadea, 4, 5, 53, 125, 172, 278, 279, 337
blanoides, 5
palirostrata,
Caecilia, 12
Caiman, 52, 60, 62, 178, 186, 218, 328
crocodilus, 52, 62, 178, 218, 328
Calyptahyla, xxiv, 3, 17, 65, 70, 76, 79, 86, 89, 271, 361
crucialis, 17, 65, 70, 75, 76, 79, 86, 89, 271
Canis familiaris, 257
Caretta caretta, 33
Celestus, 1, 3, 5, 10, 65, 66, 70, 71, 75, 78, 81, 82, 83, 85, 90, 104, 105, 119, 125, 132, 138, 140, 147, 148, 149, 150, 159, 162,

- 163, 165, 171, 172, 232, 242, 247, 256,
279, 362, 364
- agasepsoides*, 104, 119, 125, 132
- anelpistus*, xxix, 104, 132, 165
- badius*, 159
- barbouri*, 65, 81, 83, 90
- carraui*, xxix, 104, 132,
- costatus*, 87, 104, 125, 132, 138, 140,
149, 160, 162, 163
- c. aenetergum*, 104
- c. badius*, 138, 140
- c. chalcorhabdus*, 104
- c. costatus*, 104
- c. emys*, 104
- c. leionotus*, 104
- c. melanchrous*, 104
- c. neiba*, 104
- c. nesobous*, 104
- c. oreistes*, 104
- c. psychothoes*, 104
- c. saonae*, 104
- crusculus*, xxiv, 66, 78, 81, 82, 90
- c. crusculus*, 65
- c. cundalli*, 66
- c. molesworthi*, 66, 82, 90
- curtissi*, 104, 125, 132, 148
- c. aporus*, 104
- c. curtissi*, 104
- c. diastatus*, 105
- c. hylonomus*, 105
- darlingtoni*, 105, 130
- duquesneyi*, 66, 81, 90
- fowleri*, 66, 70, 71, 90, 91
- haetianus*, 105, 125, 132
- h. haetianus*, 105
- h. mylicus*, 105
- h. surdus*, 105
- hewardii*, 66, 75, 81, 82, 91
- impressus*, 81
- macrotus*, 105, 132
- marcanoi*, 105, 132, 148
- microblepharis*, 66, 85, 90
- occiduus*, 10, 15, 66, 81, 82, 91
- sepsoides*, 105, 132
- stenurus*, 105, 125, 132, 147, 162, 163
- s. allooides*, 105
- s. stenurus*, 105
- s. weinlandi*, 105
- warreni*, 105, 132
- Centropyx*, 201
- copii*, 201
- intermedius*, 20
- Chamaeleolis*, xxiv, 1, 3, 55, 128, 153,
172, 289
- chamaeleonides*, 1
- Chamaelinorops*, xxiv, 2, 3, 128, 153, 161,
172, 289
- barbouri*, 161
- wetmorei*, 153
- Chelonia*, 33
- mydas*, xxix, 33
- virgata*, 33
- Chilabothrus*, 144, 157
- inornatus*, 20
- Chironius*, 4, 22, 186, 217, 243, 251, 261,
310, 333
- vincenti*, 22, 186, 217, 251, 261, 310
- Chondrodactylus*, 280
- Chrysemys*, 6, 149
- palustris*, 6
- Cladium jamaicense*, 38
- Claudius*, 363
- Clelia*, 85, 186, 217, 243, 312, 315, 317,
333, 354
- clelia*, 186, 217, 312, 354
- c. groomei*, 217
- errabunda*, 186, 217, 312
- equatoriana*, 312
- rustica*, 312
- scytalina*, 312
- Cnemidophorus*, 3, 155, 185, 199, 243,
299, 301
- affinis*, 155
- lemniscatus*, 301
- vanzoi*, 185, 200, 301
- Cocos*, xxvi
- Coccyzus minor*, 257
- Coleodactylus*, 283
- Colostethus*, 185, 189, 242
- chalcopsis*, 185, 189
- Coluber*, 12, 14, 214, 363
- Coniophanes*, 243, 312, 313, 314, 333, 363
- andresensis*, 312
- Conolophus*, 286
- Conophis*, 312, 316, 317, 363

- Corallus*, 186, 213, 214, 230, 238, 240, 243, 251, 254, 256, 263, 267, 306, 308, 309, 333, 366
 annulatus, 308
 caninus, 308
 cooki, 214
 enydris, 186, 214, 308
 e. cooki, 214
 grenadensis, xxii, xxxvii, 238, 240, 241, 263, 267
 hortulanus, 213, 214, 306, 308
 h. cooki, 213
Cornufer unicolor, 143
Corvus, 351
Corytophanes, 363
Craugastor, 361
Crematogaster brevispinosa, 257
Crepidophryne, 271
Cricocaura, 337
Cricosaura, 2, 32, 51, 60, 232, 243, 303, 304, 305, 306, 330, 332, 361, 363
 typica, 32, 51, 60, 232, 303, 361, 363
Crisantophis, 312
Crocodilus, 31, 144, 328, 329, 334
Crocodylus, 16, 52, 62, 67, 116, 124, 159, 186, 218, 229, 257, 328
 acutus, xxix, 32, 33, 52, 62, 67, 116, 124, 144, 159, 164, 229, 257, 328, 329
 cataphractus, 329
 intermedius, 186, 218, 328, 329
 johnsoni, 328, 329
 mindorensis, 329
 moreletii, 328, 329
 niloticus, 328, 329
 novaeguineae, 328, 329
 palustris, 328, 329
 porosus, 329
 rhombifer, 16, 31, 32, 33, 52, 62, 328, 329
 siamensis, 328, 329
Crotalus, 12, 364
Crotophagous ani, 257
Ctenocercus, 152
Ctenonotus, 153, 291
Ctenosaura, 242, 286
Cyclura, xxviii, 1, 3, 8, 12, 15, 17, 18, 32, 46, 55, 66, 75, 86, 87, 102, 108, 127, 134, 138, 140, 144, 152, 160, 164, 167, 175, 176, 179, 184, 190, 196, 227, 229, 237, 242, 256, 257, 265, 266, 286, 288, 289, 331, 351, 354
 carinata, 17, 152, 229, 237, 257, 286, 288, 289
 collei, xxix, 8, 12, 18, 66, 75, 227, 286, 289
 cornuta, xxix, 108, 127, 134, 138, 140, 144, 152, 160, 164, 167, 175, 176, 286, 289
 c. cornuta, 102, 108
 c. nigerrima, 138, 140
 c. onchiopsis, 138, 167
 c. stejnegeri, xxix
 cyclura, 286, 288, 289
 mattea, 286
 nubila, xxix, 46, 55, 179, 286, 289
 n. caymanensis, 55
 n. lewisi, 55
 n. nubila, 46, 55
 nigerrima, 152
 onchiopsis, 152, 160
 pinguis, 238, 286, 288, 351
 portoricensis, 286
 ricordii, 108, 127, 134, 135, 164, 274, 286, 288, 289
 rileyi, 286, 288, 289
 r. cristata, 288
 stejnegeri, 152
Cyrtodactylus, 282
- ## D
- Dactyloa*, 153, 291
Darlingtonia, xxvi, 4, 5, 114, 130, 136, 158, 243, 251, 254, 312, 315, 316, 318, 319, 333, 362
 haetiana, 114, 130, 136, 158, 251, 255
 h. haetiana, 114
 h. perfector, 114
 h. vaticinata, 114
Dasypeltis, 310
Dasyprocta, 353
Dendrobates, 249
Dendrobates pumilio, 249
Dendrophis, 144, 158
Dermochelys coriacea, 33

- Diadophis*, 309
 punctatus, 306, 309
Didelphis marsupialis, 257
Diploglossus, xxix, 3, 5, 45, 53, 73, 85, 87, 91, 125, 148, 149, 150, 160, 162, 171, 173, 185, 190, 212, 219, 242, 247, 256, 261, 279, 362
 anelpistus, 91
 carrui, 91, 148
 delasagra, 45, 53
 fowleri, 87
 garridoi, 45, 53
 impressus, 87
 monotropis, 73
 montisserrati, 185, 190, 212, 219, 261
 pleii, xxvi, 173
 warreni, 162
Dipsas, 251
Draco, 12
Draconura catenata, 19, 76
Dromicus, 28, 87, 146, 147, 157, 216, 314
 mentalis, 158
 parvifrons, 157
 p. paraniger, 158
 p. rosamondae, 158
 p. stygius, 158
 w-nigrum, 158
Drymobius, 363
Duellmanohyla, 361
- E**
- Egretta thula*, 257
Elaphe guttata, 306, 309
Eleutherodactylus, xxvi, 1-3, 20, 27, 33, 40-43, 65, 70, 71, 74, 77-81, 83-88, 90, 100-103, 118, 120-123, 139-143, 162, 163, 165, 171-175, 177, 178, 185, 187-190, 219, 220, 225, 226, 231, 232, 234-236, 238, 239, 242, 244, 245, 248, 249, 251, 252, 256, 257, 261, 262, 264-268, 273-275, 277, 278, 282, 331, 360, 361, 363, 364, 368, 369
 abbotti, 100, 111, 121, 123, 155
 acmonis, 39, 43
 albipes, 39, 43
 alcoae, 100, 121, 123
 alticola, 65, 71, 80, 87
 amadeus, 100
 amplinympa, 185, 189
 andrewsi, 65, 71, 80
 antillensis, 172, 173, 174, 236
 apostates, 100, 121
 armstrongi, 100, 121,
 atkinsi, xxiii, 39, 43
 a. atkinsi, 39, 43
 a. estradai, 39, 43, 44
 audanti, 100, 121, 141
 a. audanti, 100
 a. melatrigonum, 100
 a. notidodes, 100
 auriculatoides, 100, 121
 auriculatus, 39, 42, 43, 274
 bakeri, 100, 121
 barlagnei, xxvii, 185, 188, 189
 bartonsmithi, 39, 43
 bransfordi, 238, 239
 bresslerae, 39, 43
 brevirostris, 100, 121
 brittoni, 173
 caribe, 100, 121
 casparii, 39, 43
 cavernicola, 65, 80
 chlorophenax, 100, 118, 121
 cochranae, 173, 174
 cooki, 173
 coqui, xxv, 27, 172, 178, 229, 236, 238, 249, 257, 266
 corona, 100, 121
 counouspeus, 100, 121
 cubanus, 39, 43
 cundalli, 65, 66, 84, 87, 90, 236
 c. cundalli, 87
 cuneatus, 39, 43
 darlingtoni, 100, 121, 143
 dimidiatus, 39, 42, 43
 d. amelasma, 39
 d. dimidiatus, 39
 dolomedes, 100, 121
 eileenae, 39, 43
 emiliae, 39, 43
 eneidae, xxiv, 173, 277
 etheridgei, 39, 43
 eunaster, 100, 121
 euphronides, 185, 189, 219

- fitzingeri*, 239
flavescens, 100, 121
fowleri, 100, 121
furcyensis, 100, 121
fuscus, 65, 71, 80
glamyrus, 39, 43
glandulifer, 100, 121
glanduliferoides, 100, 121
glaphycompus, 100, 121
glaucoreius, 65, 71, 88
goini, 39, 43
gossei, 84, 87, 236, 274
 g. gossei, 65
 g. oligaulax, 65
grabhami, 65, 79
grahami, 100, 121
greyi, 39, 43
griphus, 65, 71
gryllus, 277
guanahacabibes, 39, 42
guantanamera, 39, 43
gundlachi, 39, 41
haitianus, 100, 122, 143
hedricki, 236
heminota, 100, 122
hypostenor, 100, 122
iberia, 39, 43
inoptatus, 100, 122, 123, 143, 274
intermedius, 39, 43, 143,
ionthus, 40, 43
jamaicensis, 65, 70, 77, 78, 90
jasperi, 27, 172, 173
jaumei, 40, 43
johnstonei, xxvii, 65, 81, 85, 185, 187,
188, 190, 219, 236, 262, 268
jugans, 100, 122, 143
junori, 65
klinikowskii, 40, 42
lamprotes, 100, 122
leberi, 40, 43
lentus, 77, 174
leoncei, 100, 122
lewisi, 80, 84
limbatus, 40, 43
locustus, xxvi, 277
lucioi, 100, 106, 122
luteolus, 20, 65, 80, 84
lynni, 82
mariposa, 40, 43
martinicensis, xxvii, 77, 81, 85, 185,
187, 189, 190, 219
melacara, 40, 43
minutus, 100, 122
monensis, 175
montanus, 100, 122, 274
neodreptus, 141
nortoni, 100, 122
nubicola, 65, 71, 79, 274
orcutti, 65, 71, 79
orientalis, 40, 43
oxyrhynchus, 100, 122, 139
pantoni, 87
 p. amiantus, 65
 p. pantoni, 65
parabates, 101, 122
parapelates, 101, 122
patriciae, 101, 122
paulsoni, 101, 122
pentasyringos, 65, 71, 79, 88
pezopetrus, 40, 43
pictissimus, 101, 122, 123
 p. apanteatus, 101
 p. eremus, 101
 p. pictissimus, 101
pinarensis, 40, 43
pinchoni, 185, 189
pituinus, 101, 122
planirostris, 40, 43, 65, 80, 236, 274
 p. planirostris, 40, 65
poolei, 101, 122
portoricensis, 178, 236, 277
principalis, 40, 43
probolaeus, 101, 122, 175
rhodesi, 101, 122
richmondi, xxix, 236
ricordii, 2, 40, 42, 43, 80
ronaldi, 40, 43
rostralis, 361
rufifemorialis, 101, 122
ruthae, 101, 122
 r. aporostegus, 101
 r. bothroboans, 101
 r. ruthae, 101
 r. tyathrous, 101
schmidti, 101, 122
 s. limbensis, 101

- s. rucillensis*, 101
s. schmidti, 101
schwartzii, 174
sciagraphus, 101, 122
semipalmatus, 101, 122
shrevei, 185, 189
sisyphodemus, 65, 71
stejnegarianus, 238, 239
symingtoni, 40, 42
tetajulia, 40, 43
thomasi, 40, 43
 t. thomasi, 40, 43
 t. trinidadensis, 40, 43
 t. zayasi, 40, 43
thorectes, 101, 122, 165
toa, 40, 43
tonyi, 40, 43
turquinensis, 40, 43
unicolor, 173
unistrigatus, 274, 364
urichi, 189
varians, 40, 43
 v. olibrus, 40, 44
 v. staurometopon, 40, 44
 v. varians, 40, 44
varleyi, 40, 42, 43
ventrilineatus, 101, 122, 143, 165
warreni, 101, 123
weinlandi, 101, 123, 163
 w. chersonesodes, 101
 w. paralius, 101
 w. weinlandi, 101
wetmorei, 101, 123
 w. ceraemerus, 101
 w. diplasius, 101
 w. sommeri, 101
 w. wetmorei, 101
wightmanae, 173, 236, 257
zeus, 40, 42
zugii, 40, 43
 z. erythroproctus, 40, 43
 z. zugii, 40, 43
Embryopus habichii, 150
Enulius, 312
Enyalioides, 365
Epicrates, 2, 4, 5, 11, 12, 23, 24, 25, 33, 51, 60, 67, 113, 114, 129, 136, 144, 146, 157, 173, 177, 230, 238, 240, 243, 251, 253, 254, 260, 267, 306, 308, 309, 330, 333, 362, 363
 angulifer, xxiii, 33, 51, 60, 309
 cenchria, 254
 chrysogaster, 309
 exsul, 309
 fordii, 113, 129, 136, 146, 157, 309
 f. agametus, 114
 f. fordii, 114
 f. manototus, 114
 gracilis, 114, 129, 136, 144, 157, 273
 g. gracilis, 114
 g. hapalus, 114
 inornatus, xxvi, 20, 24, 158, 173, 177, 309
 monensis, xxix, 177, 230, 238, 240, 309
 striatus, xxv, 25, 114, 129, 136, 144, 157, 251, 267
 s. exagistus, 114
 s. striatus, 114
 s. warreni, 114
 subflavus, 11, 12, 23, 24, 67, 80, 177, 309
Epilobocera situatifrons, 257
Eretmochelys fimbriata, 33
Eridiphas, 313
Euhyas, 42, 143, 360, 363
Eumeces, 9, 289, 364
 sloanei, 9
Eunectes, 309
Euparkerella, 2
Eupristus, 152

F

- Falco sparverius*, 257
Farancia, 319
Felis, 166, 257
 catus, 257
 domesticus, 166
Ferminia cerverai, 257
Florida caerulea, 257
Frostius, 271

G

- Gastrophryne*, 249, 270
pictiventris, 249
Gecko mabouia, 151, 206
Gekko, 185, 190, 206, 212,
gekko, 185, 190, 212
Geochelone, 8, 162, 179, 185, 218, 326,
 354
carbonaria, 8, 179, 185, 218, 326
denticulata, 8, 186, 218
sombrerensis, 185, 218, 354
Geophis, 251, 312, 313
Gonatodes, 3, 45, 53, 66, 92, 102, 105,
 126, 132, 150, 185, 190, 206, 212, 238,
 242, 280, 281, 282, 331, 364
albogularis, 3, 45, 53, 66, 92, 102,
 105, 126, 132, 151, 185, 190, 212, 280
a. fuscus, 45
a. notatus, 66, 92, 102, 105
antillensis, 238
Gymnodactylus, 1, 3, 4, 14, 150
fasciatus, 3, 4, 14
Gymnophthalmus, 2, 3, 186, 190, 201, 204,
 219, 242, 299, 301
pleii, 186, 201, 202, 204, 219, 301
lineatus, 301
luetkenii, 201, 202
nesydron, 202
speciosus, 186, 204
underwoodi, 186, 204, 302

H

- Helicops*, 315, 317
Hemidactylus, 45, 53, 105, 118, 126, 131,
 132, 144, 146, 151, 163, 176, 179, 186,
 190, 206, 208, 220, 242, 255, 256, 262,
 280, 282, 331
brookii, 45, 53, 146, 151, 176, 179,
 207, 282
b. brooki, 45
b. haitianus, 53, 146, 151
garnotii, 282
haitianus, 105, 118, 126, 132, 146,
 151, 163, 255
mabouia, 43, 53, 105, 126, 131, 132,
 144, 151, 179, 186, 206, 207, 208, 282
palaichthus, 186, 206, 208, 282
turcicus, 45, 53, 282
t. turcicus, 45
Herpestes javanicus, xxx, 165, 257, 268
Herpetodryas carinatus, 217
Heterodon, 322
Hispaniolus, 156
Homalochilus, 157
Homonota, 4, 14
Hydrodynastes, 315
Hyla, 1, 3, 17, 65, 70, 75, 76, 79, 84, 86,
 90, 100, 120, 121, 139, 142, 163, 178, 187,
 188, 242, 256, 270, 271, 272, 361
albomarginata, 272
barbudensis, 188, 191
boans, 272
brunnea, 86
cinerea, 178, 272
crucifer, 41, 270
heilprini, xxiv, 70, 100, 120, 121, 142,
 272, 361
lichenata, 17, 79, 86, 272
marianae, 65, 79, 90, 271, 272
pulchrilineata, 70, 90, 100, 120, 121,
 139, 142, 163, 272
shrevei, 84
squirella, 272
vasta, 70, 90, 100, 120, 121, 139, 142,
 272
wilderi, xxiv, 65, 79, 84, 90, 271, 272
Hylodes, 77, 81, 143, 187
martinicensis, 77
oxyrhynchus, 143
Hypsiboas, 142
Hypsilophus, 152
ricordii, 152
Hypsirhynchus, 4, 5, 21, 114, 130, 136,
 146, 158, 164, 243, 252, 253, 255, 312,
 315, 316, 317, 319, 333, 362
ferox, 21, 114, 130, 136, 146, 158,
 164, 252, 253, 255, 319
f. exedrus, 114, 130
f. ferox, 114
f. paracrousis, 115, 130
f. scalaris, 115
scalaris, 158

I

- Ialtris*, 4, 21, 115, 130, 136, 146, 158, 165, 243, 256, 315, 317, 319, 334, 362
agyrtes, 115, 130, 136, 158, 165
dorsalis, 21, 115, 130, 136, 146, 158, 317
parishi, 115, 130, 136, 158, 165
vultuosa, 158
Iguana, 3, 13, 152, 179, 186, 190, 191, 195, 196, 219, 220, 221, 242, 256, 286, 287, 288, 331, 354
delicatissima, 13, 186, 195, 196, 221, 286, 354
iguana, 13, 179, 186, 195, 196, 286, 288, 353, 354
tuberculata, 13, 186, 195, 196
t. rhinolopha, 195, 196
Imantodes, 312

J

- Jaltris*, 158

K

- Kentropyx*, 186, 190, 199, 201, 243, 366
borckiana, 186, 201
Kinosternon, 326, 363
scorpiodes, 326
Klauberina, 305

L

- Lacerta*, 12, 13, 14, 15, 144, 152, 195, 209
bimaculatus, 13, 14
iguana, 13
occidua, 15
scincoidea, 15
Lachesis, 2, 4
Laemanctus, 363
Lamprophis, 318
Leimadophis, 4, 5, 157, 217
alleni, 157
mariae, 217

parvifrons, 158

p. lincolni, 158

p. niger, 158

tortuganus, 157

Leiocephalus, 1, 3, 28, 50, 58, 59, 71, 112, 113, 118, 135, 138, 140, 144, 147, 156, 160, 162, 167, 172, 184, 186, 190, 197, 198, 223, 232, 237, 242, 247, 250, 251, 255, 256, 257, 261, 289, 291, 302, 303, 332, 351, 354, 361

altavelensis, 156

anonymous, 162

apertosulcus, 162

barahonensis, 112, 135, 156, 257

b. aureus, 112

b. barahonensis, 112

b. beatanus, 112

b. oxygaster, 112

carinatus, xxiii, 28, 50, 58, 217, 223, 237, 257

c. aquarius, 50, 58

c. carinatus, 50, 58, 217

c. cayensis, 50

c. labrossytus, 50, 58

c. microcyon, 50, 58

c. mogotensis, 50, 58

c. zayasi, 58

cubensis, 28, 50, 58

c. cubensis, 50, 58

c. gigas, 50, 58

c. minor, 50, 58

c. pambasileus, 50, 59

c. paraphrus, 50, 59

cuneus, 186, 190, 198, 354

endomychus, 113, 118, 135, 157

eremitus, 138, 140, 156, 159, 167

herminieri, 186, 190, 197, 354

jamaicensis, 71

lunatus, 113, 135, 156

macropus, 50, 59, 156

m. aegialus, 50, 59

m. asbolomus, 50, 59

m. felinoi, 50, 59

m. hoplites, 50, 59

m. hyacinthurus, 50, 59

m. immaculatus, 50, 59

m. koopmani, 50, 59

m. lenticulatus, 50, 59

- m. macropus*, 50, 59
m. phylax, 50, 59
m. torrei, 50, 59
melanochlorus, 113, 135, 138, 140, 156, 160
m. hypsisus, 113
m. melanochlorus, 113
onaneyi, 50, 59
personatus, 113, 135, 147, 156, 162
p. actites, 113
p. agraulus, 113
p. budeni, 113
p. elattoprosopon, 113
p. mentalis, 113
p. personatus, 113
p. poikilometes, 113
p. pyrrholaemus, 113
p. scalaris, 113
p. socoensis, 113
p. tarachodes, 113
p. trujilloensis, 113
pratensis, 113, 135, 156
p. chimarus, 113
p. pratensis, 113
raviceps, 28, 50, 59
r. delavarai, 50, 59
r. jaumei, 50, 59
r. klinikowskii, 50, 59
r. raviceps, 50, 59
r. uzzelli, 50, 59
rhutidira, 113, 135, 157
schreibersii, xxv, 113, 135, 144, 237, 257
s. nesomorus, 113
s. schreibersii, 113
semilineatus, 113, 135, 156
stictigaster, 50, 59
s. astictus, 50, 59
s. celeustes, 50, 60
s. exothetus, 50, 59
s. gibarensis, 51, 59
s. lipomator, 51, 59
s. lucianus, 51, 59
s. naranjoi, 51, 59
s. ophiplacodes, 51, 59
s. parasphex, 51, 59
s. septentrionalis, 51, 59
s. sierrae, 51, 59
s. stictigaster, 50, 59
trigeminatus, 156
vinculum, 113, 135, 156
v. altavelensis, 113
v. vinculum, 113
Lepidophyma, 305, 361, 363
Leptodactylus, 1, 3, 70, 101, 120, 123, 140, 143, 172, 174, 175, 176, 185, 187, 188, 189, 242, 246, 255, 257, 273, 331, 354
albilabris, 70, 120, 143, 172, 174, 175, 176, 273
bufonius, 273
dominicensis, 101, 120, 123, 143, 176, 273
fallax, 185, 255, 257, 273, 354
flavopictus, 273
fuscus, 273
insularum, 273
knudseni, 273
labialis, 273
labyrinthicus, 273
latinasus, 273
mystaceus, 273
pentadactylus, 188, 273
validus, 185, 187, 188, 189
wagneri, 188, 189, 273
Leptodeira, 312, 313, 314
Leptophis, 157, 314
Leptotyphlops, 2, 4, 5, 115, 131, 136, 138, 159, 186, 215, 243, 320, 362
asbolepis, 115, 131, 136, 321
bilineata, 131, 186, 215, 321
calypso, 115, 131, 136, 321
columbi, 320, 362
goudotii, 320
leptipilepta, 115, 131, 136, 321
pyrites, 115, 131, 136, 138, 159, 321
tenella, 186, 215, 320
Lineatriton, 1
Liocephalus [sic], 156
Liolemus, 156
Liophis, 4, 14, 159, 186, 216, 217, 219, 243, 256, 312, 314, 316, 334, 363
cursor, 14, 186, 216, 217, 314
juliae, 186, 217, 314
j. copeae, 217
j. mariae, 217
melanotus, 186, 217, 273, 314

ornatus, 186, 217, 314
perfuscus, 186, 217, 219, 314
putnami, 14, 217
taeniatus, 314
triscalis, 314
viridis, 312
Lithodytes lentus, 77
Litoria luteola, 78, 84
Loxocemus, 322
Lygodactylus, 286

M

Mabuya, 1, 4, 9, 12, 66, 102, 111, 128, 131, 135, 138, 144, 154, 174, 175, 176, 186, 190, 212, 213, 238, 240, 242, 256, 299, 364
bistriata, 102, 111, 128, 131, 135, 138, 144, 154, 299
b. sloanei, 102, 111, 128, 155
lanceolata, 213
lineolata, 111, 128, 135, 138, 154, 299
mabouia sloanei, 10
mabouya, 154, 174, 175, 176, 186, 212, 213, 238, 240, 29
m. mabouya, 176
m. sloanei, 176
metallica, 213
sloanei, 12, 18, 212
Macrotus waterhousii, 351
Margarops fuscatus, 257
Mastigodryas, 4, 186, 217, 243, 310, 334
bruesi, 186, 217, 310
Megalocercus, 158
Melanerpes portoricensis, 257
Metopoceros, 152
Micrurus, 243, 259, 260, 306, 320, 334
alleni, 320
diastema, 320
distans, 320
fulvius, 320
nigrocinctus, 320
ruatanus, 259, 260, 306, 320
Mimus gilvus, 257
Mormoops blainvillei, 351
Mus, 263, 267

musculus, 268
Myiarchus tyrannulus, 257

N

Natrix capistrata, 76, 227
Neotarentola, 286
Nerodia, 51, 60, 61, 243, 310, 334, 364
clarki, 51, 61, 310, 364
c. compressicauda, 51, 61
fasciata, 310
sipedon, 310
Norops, 1, 2, 3, 153, 291
ophiolepis, 1

O

Oedipina, 363
Oedipus lineolus, 1
Oligoctenus otleyi, 257
Olios antiguensis, 257
Ololygon, 272
rubra, 187, 272
Ophisaurus ventralis, 279
Oryzomys, 263
Osteocephalus, 272
Osteopilus, xxiv, xxv, 3, 7, 20, 40, 42, 65, 70, 86, 89, 90, 100, 119, 121, 123, 139, 142, 163, 178, 185, 187, 242, 244, 245, 249, 251, 255, 256, 257, 266, 271, 272, 361
brunneus, xxiii, 7, 20, 65, 89, 90, 271, 272
dominicensis, 70, 100, 119, 123, 139, 142, 244, 249, 255, 257, 272
septentrionalis, 40, 42, 70, 178, 185, 187, 249, 255, 257, 272
Otus nudipes, 257
Oxybelis, 240
Oxyrhopus, 312

P

Palaeoxantusia, 305
Panolopus, 149
Pelamis platurus, 259, 260
Pelophilus, 146, 157

Pelorius, 42, 143, 277, 360
Peltaphryne, 3, 142
Peltophryne, xxii, 118, 142, 249, 270, 271, 361
 cataulaciceps, 271
 empusa, 271
 fluviatica, 271
 guentheri, 271
 gundlachi, 271
 lemur, 271
 longinasa, 271
 peltacephala, 271
 taladai, 271
Pelusios, 185, 218, 326
 subniger, 185, 218, 326
Phelsuma, 239
Phenacosaurus, 289
Philodryas, 158, 316, 317
Phrynopus, 2
Phrymus longipes, 257
Phyllobates, 1, 2, 3
 limbatus, 1, 2
Phyllodactylus, 3, 5, 27, 105, 126, 132, 148, 151, 173, 176, 186, 190, 206, 212, 242, 280, 282, 3
 hispaniolae, 27
 martini, 282
 pulcher, 27, 151, 186, 212, 282
 sommeri, 27
 spatulatus, 212
 underwoodi, 212
 ventralis, 212
 wirshingi, 27, 105, 126, 132, 148, 151, 173, 176, 282
 w. hispaniolae, 105, 126, 151
 w. sommeri, 105, 151
Phyllonycteris major, 351
Phylodryas, 146
Pinus, 38
 caribea, 38
 cubensis, 38
 maestrensis, 38
 tropicalis, 38
Pipa, 365
Placopsis ocellata, 19, 76
Platymantis, 143
Plectrohyla, 361, 363
Pleurodema, 365

Pliocercus, 313
Polychrus, 289, 291
Pristinotus, 144, 156
 schreibersii, 156
Psammophis antillensis, 216
Pseudacris crucifer, 38
Pseudemys, 147, 149, 327
Pseudoboa, 186, 217, 312, 334
 neuwiedi, 186, 217
 nigra, 312
Pseudogonatodes, 283
Pseustes, 314
Ptychohyla, 361, 363
Pumilia, 286
Python, 308, 322

Q

Quiscalus lugubris, 257

R

Ramphotyphlops braminus, 186, 215
Rana, 7, 12, 40, 44, 65, 86, 102, 119, 120, 139, 166, 178, 270, 363
 catesbeiana, 40, 44, 65, 86, 102, 119, 120, 139, 166, 178
 grylio, 270
Rattus, 263, 267
 rattus, 166, 257, 268
Rhadinaea, 313
Rhamphophryne, 271

S

Sanzinia, 308
Saphenophis, 318
 boursieri, 317
 sneideri, 318
Sauresia, 3, 5, 125, 144, 149, 150, 172, 279, 362
 sepsoides, 144, 150
Sauromalus, 286, 288
Saurothera, 257
 merlini, 257

- vieilloti*, 257
Scaphiodontophis, 363
Sceloporus, 363
Scinax, 178, 185, 187
 rubra, 178, 185
Scincus, 9, 10, 12, 144, 155
 bistriatus, 144
 sloanei, 9, 12
 sloanii, 144, 155
Scolecosa[ur]us alleni, 205
Scolecosaurus alleni parviceps, 205
Scolopendra alternans, 257
Semiurus, 291
Sibon, 313, 314
Smilisca, 363
Sminthillus, 2
 brasiliensis, 2
 peruvianus, 2
Speleperes infuscata, 1, 3
Sphaeriodactylus, 144
Sphaerodactylus, xxvi, 1, 3, 5, 10, 13, 19, 22, 28, 29, 33, 45, 46, 53, 54, 66, 70, 71, 75, 76, 78, 81, 82, 87, 92, 102, 105-107, 118, 126, 132-134, 138, 140, 144, 146-148, 151, 161, 163, 171, 173-177, 186, 190, 206, 209-212, 220, 231, 232, 236, 242, 246, 256, 257, 262, 264, 266, 280-285, 331, 348, 349, 361, 364, 366, 368, 369
 altavelensis, 105, 127, 132
 a. altavelensis, 105
 a. brevirostratus, 105
 a. enriquilloensis, 106
 a. lucioi, 106
 argus, 10, 19, 45, 54, 66, 82, 87, 92, 152, 177, 227, 284
 a. andresensis, 54
 a. argus, 45, 66
 a. henriquesi, 82
 armasi, 45, 54, 55
 armstrongi, 106, 133
 a. armstrongi, 106
 a. hypsinephes, 106
 asterulus, 106, 133
 beatyi, 176, 349
 becki, 138, 140, 161
 bromeliarum, 28, 45, 54, 55
 callocricus, 106, 133
 celicara, 45, 54, 55
 cinereus, 106, 133, 144, 152
 c. cinereus, 106
 c. stejnegeri, 106
 clenchi, 106, 133, 236
 c. apocoptus, 106
 c. clenchi, 106
 cochranae, 106, 133, 147
 copei, 106, 126, 133, 138, 140, 146, 147
 c. astreptus, 106
 c. cataplexis, 106
 c. copei, 106
 c. deuterus, 106
 c. enochrus, 106
 c. pelates, 106
 c. picturatus, 106
 c. polyommatus, 106
 c. websteri, 106
 copii, 210
 cricoderus, 45, 54, 55
 cryphius, 106, 126, 133
 dacnicolor, 66, 71, 78, 88
 darlingtoni, 106, 133
 d. bobilini, 106
 d. darlingtoni, 106
 d. mekistu, 106
 d. noblei, 106
 difficilis, 28, 106, 107, 127, 133, 163, 262, 283, 284
 d. anthracomus, 107
 d. difficilis, 107
 d. diolenius, 107
 d. euopter, 107, 127
 d. lycauges, 107
 d. peratus, 107
 d. typhlopous, 107, 127
 docimus, 45, 54, 55
 dommeli, 163
 elasmorhynchus, 107, 133, 152
 elegans, 45, 54, 102, 107, 126, 133, 144
 e. elegans, 54
 e. punctatissimus, 102, 107, 126
 elegantulus, xxvii, 186, 210, 211, 212
 epiurus, 107, 133
 fantasticus, 186, 210, 211, 212
 f. fuga, 210
 f. ligniservulus, 210

- festus*, 210
gaigeae, 173
gilvitorques, 66, 76, 87, 92
goniorhynchus, 66, 76, 92
grandisquamis, 283
intermedius, 45, 54
kirbyi, 186, 211
klauberi, 173, 257
ladae, xxv, 107, 133
lazelli, 107, 133
leucaster, 107, 133
lineolatus, 283
macrolepis, 173, 174, 176, 186, 209, 220, 236, 283, 349
 m. parvus, 176, 210
microlepis, 210, 211, 212
 m. thomasi, 211
micropithecus, 175
molei, 283
nicholsi, xxvi 173, 174
nigropunctatus, 45, 53, 54, 284
 n. alayoi, 45, 54
 n. granti, 46, 54
 n. lissodesmus, 46, 54
 n. ocujal, 46, 54
 n. strategus, 46, 54
notatus, 46, 54, 66, 92, 102, 105, 126, 150, 283
 n. atactus, 46, 54
nycteropus, 107, 133
ococae, 107, 133
oliveri, 46, 54, 55
omoglaux, 107, 126, 133
oxyrhinus, 19, 66, 70, 71, 88
parkeri, xxiv, 66, 81
parthenopion, 174
perissodactylius, 107, 133
picturatus, 147
pictus, 209
plummeri, 107, 133
ramsdeni, 46, 54, 55, 284
randi, 107, 127, 133
 r. methorius, 107
 r. randi, 107
 r. strahmi, 107
rhabdotus, 107, 133
richardi, 46, 54, 55
richardsoni, 75, 81, 92
 r. gossei, 66, 81, 92
 r. richardsoni, 66
roosevelti, 173, 177
ruibali, 46, 54, 55
sabanus, 186, 209, 211, 212
samanensis, 107, 118, 133
savagei, 107, 127, 133
 s. juanilloensis, 107, 127
 s. savagei, 107
scaber, 28, 46, 54, 55
schwartzi, 46, 54, 55
semasiops, xxiv, 66, 70, 71, 87
shrevei, 107, 133, 148
sommeri, 134, 151
sputator, 13, 14, 152, 186, 209, 211, 212
streptophorus, 107, 134
 s. sphenophanes, 107
 s. streptophorus, 107
storeyae, 46, 54, 55
thompsoni, 107, 127, 134
titan, 80
townsendi, 173, 175, 177
torrei, 46, 50, 53, 55, 59
 t. spielmani, 46, 55
 t. torrei, 46, 55
vincenti, 22, 28, 186, 210, 211, 212, 236,
 v. festus, 210
 v. monilifer, 210
williamsi, 107, 134
zygaena, 107, 134
Stasnia portoricensis, 257
Staurotypus, 363
Steironotus, 156
Stenocercus, 303
Syrrhophus, 274, 360

T

- Tarentola*, 2, 3, 46, 53, 242, 280, 286
 americana, 46
 a. americana, 46
Testudo, 12, 75
Thamnophis, 364
 couchii, 310

- Thecadactylus*, 179, 186, 190, 206, 208, 242, 280, 286, 366
 rapicauda, 179, 186, 208, 286
Tiliqua jamaicensis, 73
Tityus obtusus, 257
Todus mexicanus, 257
Tomodactylus, 274
Trachemys, 6, 14, 44, 45, 67, 75, 102, 103, 104, 124, 147, 149, 164, 171, 174, 176, 185, 218, 256, 326, 327, 330, 334, 364
 decorata, 103, 124, 147, 149, 326, 327
 decussata, 44, 45, 53, 149, 326, 327
 d. angusta, 45, 53
 d. decussata, 45, 53
 scripta, 149, 185, 218, 326, 327, 330
 stejnegeri, xxix, 104, 124, 149, 174, 176, 185, 218, 326, 327
 s. stejneri, 218
 s. vicina, 102, 104, 124, 149
 terrapen, 14, 67, 75, 149, 326, 327
Trachyboa, 321
Trachycephalus, 90, 272
 anochloros, 76
 lichenatus, 76
Tretanorhinus, 4, 51, 60, 61, 243, 260, 312, 313, 314, 334, 364
 moquardi, 314
 variabilis, 51, 52, 61, 314
 v. binghami, 52, 61
 v. insulaepinorum, 52, 61
 v. variabilis, 52, 61
 v. wagleri, 52, 61
Tretioscincus, 243
Trigonocephalus, 214
Tripriion, 361
Tropidophis, 2, 5, 28, 52, 62, 67, 82, 83, 115, 131, 136, 138, 140, 157, 159, 167, 232, 243, 256, 260, 267, 321, 322, 334, 362
 canus, 322
 conjunctus, 157
 feicki, 52, 62
 fuscus, 52, 62
 haetianus, 52, 62, 82, 115, 131, 136, 157, 160, 322
 h. haetianus, 52, 115, 157
 h. hemerus, 115
 h. tiburonensis, 115
 jamaicensis, 67
 maculatus, 28, 52, 62, 82, 83, 157
 m. jamaicensis, 82
 m. stullii, 82
 melanurus, xxiii, 52, 62, 138, 140, 160, 167
 m. bucculentus, 138, 167
 m. dysodes, 52, 62
 m. ericksoni, 52, 62
 m. melanurus, 52, 62
 nigriventris, 52, 62
 n. hardyi, 52, 62
 n. nigriventris, 52, 62
 pardalis, 28, 52, 62, 82, 83, 160
 p. stejneri, 82
 p. galacelidus, 52, 62
 pilsbryi, 52, 62
 p. pilsbryi, 52, 62
 semicinctus, 52, 62
 stejnegeri, 67
 stullae, 67, 83
 wrighti, 52, 62
Tropidurus, 156, 303
Turdus, 257
 nudigenis, 257
 plumbeus, 257
Typha domingensis, 38
Typhlops, 2, 4, 5, 12, 15, 31, 52, 62, 67, 75, 88, 116, 119, 131, 136, 137, 138, 140, 148, 159, 164, 167, 171, 173, 174, 176, 178, 186, 214, 215, 232, 238, 241, 243, 256, 260, 322, 323, 324, 334, 362, 368
 biminiensis, 52, 62
 b. biminiensis, 52
 bilineatus, 215
 caecatus, 323
 capitulatus, 116, 131, 136, 159
 catapontus, 174
 dominicana, 213, 214, 215
 dominicanus, 186
 gonavensis, 116, 131, 136, 159
 granti, 173
 guadeloupensis, 215
 hectus, 116, 131, 136, 159
 hypomethes, 173, 174, 178, 323
 jamaicensis, xxiv, 67, 88, 215, 323, 325
 lumbricalis, 12, 31, 52, 62, 159

monastus, 215, 325
m. geotomus, 215
platycephalus, 174, 176, 178, 323
pusillus, 116, 119, 131, 136, 159, 241
richardi, 174, 178, 215, 238
rostellatus, 173, 178
schwartzi, 116, 131, 136, 159
sulcatus, 116, 131, 136, 138, 140, 159, 167
syntherus, 116, 131, 136, 159, 164
tasymicris, 186, 215
tetrathyreus, 116, 131, 136, 159
titanops, 116, 131, 137, 159
Tyrannus dominicensis, 257
Tyto alba, 257

U

Ungalia bucculentus, 159, 160
Ungaliophis, 157
Ungualia haetiana, 157
Uranoscodon, 303
Uromacer, 4, 5, 25, 115, 130, 136, 144, 146, 147, 158, 164, 243, 244, 252, 253, 255, 256, 267, 312, 315, 316, 317, 319, 334, 362
catesbyi, xxv, 25, 115, 130, 136, 144, 158, 164, 244, 319
c. catesbyi, 115
c. cereolineatus, 115, 130
c. frondicolor, 115
c. hariolatus, 115
c. inchausteguii, 115, 130
c. insulaevaccarum, 115, 130
c. pampineus, 115
dorsalis, 158
frenatus, 115, 130, 136, 147, 157, 244, 252, 319
f. chlorauges, 115
f. dorsalis, 115, 130
f. frenatus, 115
f. wetmorei, 115, 130
oxyrhynchus, 115, 130, 136, 158, 244, 253, 255, 319
scandax, 158
wetmorei, 158
Urotheca, 313

W

Wetmorena, 3, 5, 27, 125, 149, 150, 172, 279, 362
haetiana, 150

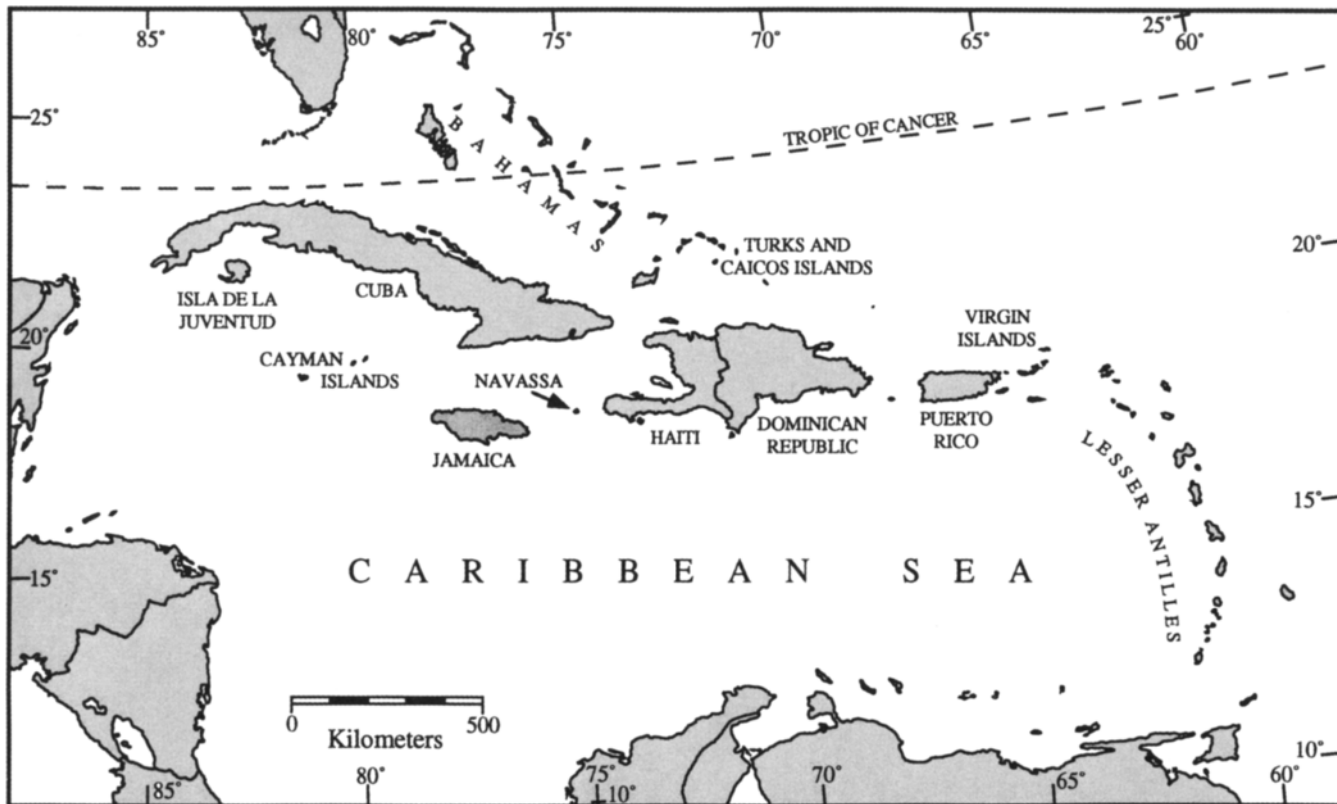
X

Xantusia, 305, 306
riversiana, 305
Xenoboa cropanii, 308
Xenodon, 363
Xiphocercus, 1, 2, 3, 153
darlingtoni, 153
valencienni, 1
Xiphosurus, 76

Z

Zamenis, 157

Color Plates



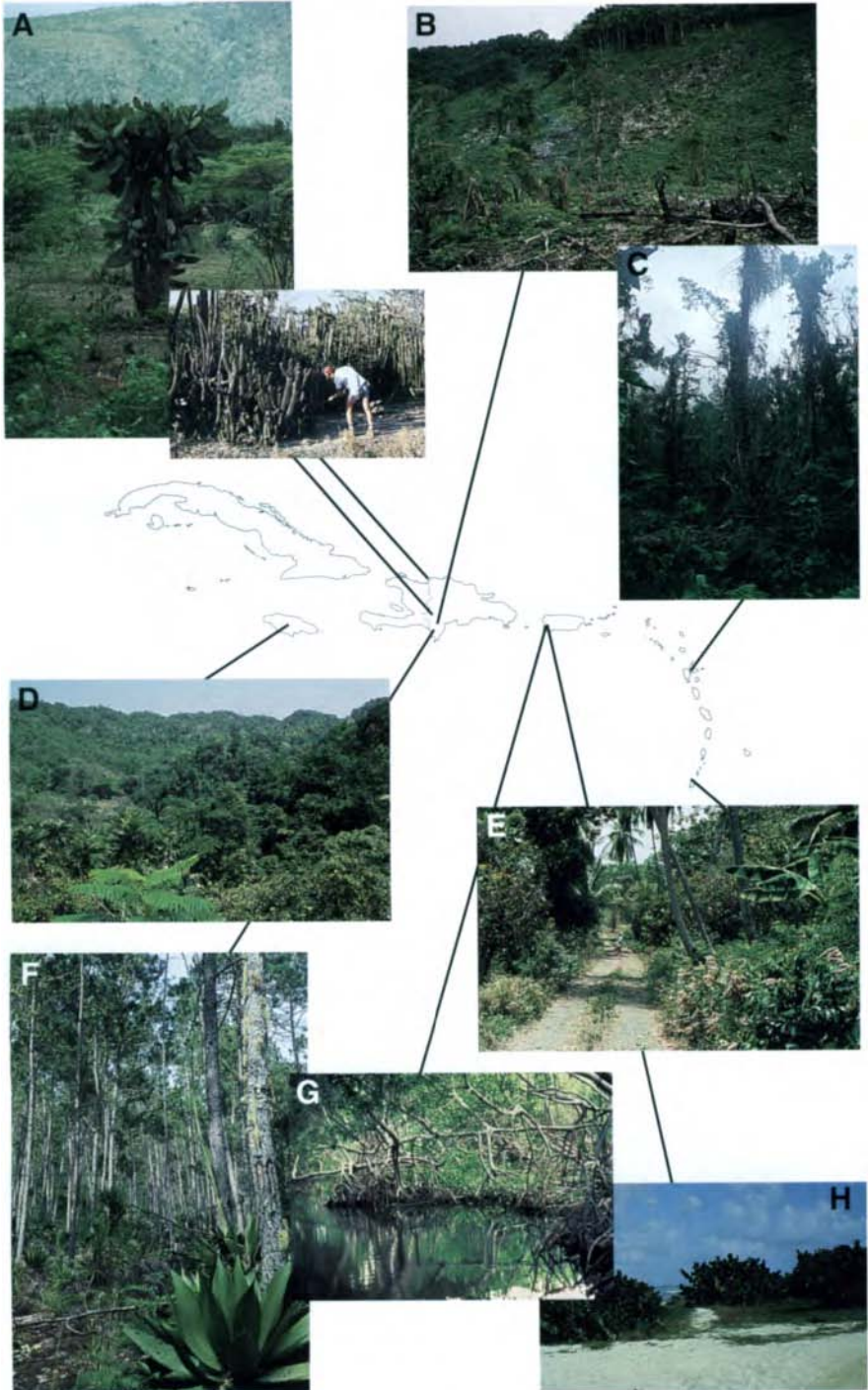


Plate 1



Plate 2



Plate 3



Plate 4



Plate 5



Plate 6



Plate 7



Plate 8